

## Review Article

# Thyroid Hormone Signaling in Muscle Development, Repair and Metabolism

Jang-Won Lee<sup>1</sup>, Nam-Ho Kim<sup>1</sup> and Anna Milanesi<sup>2\*</sup><sup>1</sup>Department of Neurosurgery, Cedars-Sinai Medical Center, USA<sup>2</sup>Department of Medicine, Division of Endocrinology, VA Greater Los Angeles Healthcare System, USA

Special Issue on

## Role of Thyroid Hormone in Metabolic Homeostasis

\*Corresponding author

Anna Milanesi, Division of Endocrinology, VA Greater Los Angeles Healthcare System and Department of Medicine, David Geffen School of Medicine at UCLA 90073, Email: amilanesi@mednet.ucla.edu

Submitted: 12 June 2014

Accepted: 17 July 2014

Published: 19 July 2014

ISSN: 2333-6692

Copyright

© 2014 Milanesi et al.

OPEN ACCESS

### Keywords

- Thyroid hormone
- Muscle development
- Muscle repair
- Muscle function
- Muscle metabolism

### Abstract

Skeletal muscle is a plastic organ made by highly specialized fibers with specific and different structure, function and metabolism. Skeletal muscle fibers can adapt, change, recover/regenerate after injury in response to various stimulators including hormones. Thyroid hormones are important players in the homeostasis of several tissues including skeletal muscle and their genomic action mostly depend on the tissue T3 bioavailability and on the distribution of the thyroid receptor isoforms which act as transcription factors and are modulated by T3. Changing in contractile and metabolic properties of the muscle fibers has been described in experimental models of hyper and hypothyroidism. Animal models with disruption of thyroid hormone signaling showed different and specific skeletal muscle phenotypes. By focusing on thyroid hormone signaling in skeletal muscle homeostasis, we review T3 specific action on skeletal muscle development, postnatal growth, function and metabolism.

## INTRODUCTION

Thyroid Hormones (THs) act in various tissues during development and post-natal life by modulating genes expression [1-3]. Triiodothyronine (T3) has actions in virtually every tissue, including skeletal muscle [4-7]. Myopathic changes have been found in the majority of the patient with hypothyroidism [8] including muscle weakness and pseudohypertrophy, myasthenic syndrome and rhabdomyolysis. On the other hand different degrees of muscle weakness and atrophy are also well known in hyperthyroid patients [9]. Even the mechanisms are not fully understood, the myopathy secondary to thyroid dysfunctions outlines the importance of THs and their signaling in skeletal muscle phenotypes and functions.

The genomic actions of T3 are mediated by thyroid hormone nuclear receptors (TRs), which act regulating gene transcription [10]. The two TR isoforms, TR $\alpha$  and TR $\beta$ , are expressed in specific temporal and spatial patterns during development with relative expression varying in different tissues and cell types [11]. Both TRs are present in the skeletal muscle with predominant expression of TR $\alpha$  [12]. In addition, it has been identified in the mitochondria a 43-KD truncated form of the nuclear receptor TR $\alpha$ 1 (p43), important in skeletal muscle development and function, which is stimulated by T3 [13]. Moreover, tissue action of T3 is regulated by deiodinase enzymes that are able to activate T4 to T3 (type 2 deiodinase; DIO2) or inactivate both T4 and T3 (type 3 deiodinase; DIO3) [14]. The presence of DIO2 in skeletal

muscle [15] indicates a possible critical role of thyroid hormone signaling in muscle phenotype and homeostasis [16].

Here, we will review the role of THs and their signaling in skeletal muscle development, postnatal growth, function and metabolism.

## THYROID HORMONES AND SKELETAL MUSCLE DEVELOPMENT

T3 is considered a regulator of muscle development *in vivo*. During embryonic development muscle progenitor cells are induced to differentiate by signaling from the near tissues such as notochord, and dorsal and lateral endoderm through activation of myogenic regulatory factors (MRFs). THs stimulate expression of several MRFs, including myogenin and myoblast determination protein 1 (MyoD1) [17,18].

It is well known that T3 stimulates skeletal muscle growth by increasing number and diameter of the muscle fiber [19]. Moreover T3 plays a role in the transition of neonatal to adult myosin isoforms. Immunocytochemical and biochemical studies showed that lack of THs results in a persistence of embryonic myosin in fetal skeletal muscle in rats [20] and excessive THs lead to accumulate myosin heavy chains resulted in precocious muscle maturation [21] of human fetal skeletal muscles (7-40 weeks of gestation). However, the absence of embryonic and fetal MHC isoform in TRs-deficient mice suggests that the transition from neonatal to adult myosin isoforms is not mediated by

only thyroid hormone [22]. In addition, hypothyroidism does not block but delay transitioning from embryonic/fetal to adult myosin [23].

Mice with knockdown of both TR  $\alpha$  and  $\beta$  (TR $\alpha\beta$ <sup>-/-</sup>) has significant lower muscle weight at birth resulting from smaller and fewer muscle fibers, indicating a role of TH on skeletal muscle development and possible skeletal muscle wasting [22] (Table 1).

### Thyroid hormone and postnatal muscle growth and repair

Maintenance and repair of skeletal muscles during post-natal life primarily rely on specialized myofiber-associated mononuclear cells called Satellite Cells (SCs). SCs are located in close contact with muscle fibers and beneath the basal lamina [24], and express early myogenic transcription factors, such as paired box protein 7(Pax7). Proliferation and differentiation of SCs are crucial to maintain normal muscle mass during adult life and regenerate new muscle fibers after injury [25]. To be activated SCs require the induction of MyoD and Myf5. Activation of SC consists of both proliferation and differentiation, which is prerequisite to supply new fully differentiated myofibers as well as maintain a pool of SCs with ‘Stemness’ [26]. Any perturbation in this process will result in the impaired regeneration of skeletal muscles in response to injury and skeletal muscle wasting overtime. Mice carrying mutation of the Pax7 gene are the critical example. Mice with Pax7 deletion can be born alive with normal but smaller skeletal muscles and then they initiate developing severe and lethal muscle wasting [27-29].

T3 has a crucial role in post-natal muscle growth. The level of T3 and the DIO2 activity significantly increase immediately after birth in mice [15], which corresponds to the switch from fetal to adult muscle fiber. Addition of T3 to the myogenic culture medium stimulates myoblast toward terminal differentiation [30] which can be mediated by cell cycle exit of myoblasts [30]. It is well known that myogenin and MyoD are induced by T3 [17,18]. Moreover, in proliferating avian myoblasts TR $\alpha$  transcriptional activity is directly stimulated by MYOD expression, and vice versa TR $\alpha$  represses MyoD activity [31]. Considering that MyoD is an

important transcription factor during myoblast proliferation and onset of myoblast differentiation, there should be a feedback mechanism between TR $\alpha$  and MYOD that could potentially play a critical role in regulating myoblast homeostasis. In addition to the inhibition of MyoD, TR $\alpha$  inhibits AP-1 activity [32], which is a strong inhibitor of differentiation. These observations raise the possibility that T3 regulates both myoblast proliferation and differentiation, depending on the stage of proliferation/differentiation processes.

Interestingly, mice with knockout of the predominant enzyme in muscle that activates T4 by converting it to T3, 5'-Deiodinase 2 (DIO2), showed impaired differentiation of muscle derived stem cells to myotubes *in vitro* and defective *in vivo* muscle regeneration after injury [15] (Table 1).

A mouse model with deletion of the mitochondrial T3 receptor p43 developed muscle hypertrophy [33] (Table 1). On the other hand, over expressing p43 [13] leads to muscle wasting with aging [34] (Table 1), suggesting a possible toxic effect of a prolonged stimulation of mitochondrial activity that leads to deficit of new skeletal muscle fiber replacement and differentiation overtime.

### Thyroid hormones and skeletal muscle function

Functional muscle alterations are common in both hyper- and hypothyroidism. A number of genes critical for muscle function and metabolism are T3 regulated [35].

In skeletal muscle, myosin is a critical protein necessary for the production of mechanic work and motion. There are 10 Myosin Heavy Chain (MHC) isoforms in the striated muscle of mammals. All members of MHC family respond to T3 [36,37], but the response is muscle and muscle fiber specific [38], ie., even the same myosin can be stimulated or inhibited depending on muscles [38]. The skeletal muscle has 4 major muscle fibers from the slowest contraction speed (type I muscle fiber) to progressively higher speeds (type IIa, IIx, IIb). Increased frequency of Type 2 fibers has been reported in muscle biopsy of patients with hyperthyroidism [39,40]. In hypothyroid mice, it has been described a mild switch from fast to slow fiber and

**Table 1:** Animal models with altered thyroid hormone signaling.

Study	species	model	Development	Repair/ Maintenance	Function	Metabolism
Pessemesse et al. [33]	Mice	p43 <sup>-/-</sup>	Muscle hypertrophy			↑ glycolytic metabolism
Casas et al. [34,44]	Mice	p43 over expression		Muscle atrophy with aging	Switch to type I MHC	↑ oxidative ↓glycolytic metabolism
Yu [22]	Mice	TR $\alpha$ <sup>-/-</sup> $\beta$ <sup>-/-</sup>	Muscle hypotrophy		Switch to type I MHC	↑ oxidative ↓glycolytic metabolism
Yu [22]	Mice	TR $\alpha$ <sup>-/-</sup>			Switch to type I MHC	↑ oxidative ↓glycolytic metabolism
Dentice [15]	Mice	DIO2 <sup>-/-</sup>		Impaired muscle regeneration		Insulin resistance
Fonseca [63]	Mice	SKM-D2KO				No metabolic phenotype

**Abbreviations:** MHC: Myosin Heavy Chain; TR: Thyroid Receptor; SKM-D2KO: Skeletal Muscle- Deiodinase 2 Knockout Mice

an increase of hybrid fibers [41]. Muscle with the prevalence of slow fibers has been shown to respond more dramatically to hypothyroidism up to a complete switch from fast to slow fibers [4]. Moreover, T3 directly stimulates the expression of sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase 1 (SERCA1) that is crucial for the fast skeletal muscle fibers [42], and reduces expression and activity of calcineurin that is involved in the slow muscle fiber phenotype [35].

Mice lacking of thyroid receptor  $\alpha$  (TR $\alpha$ <sup>-/-</sup>) showed the significantly reduced number of fast type II fibers and relatively increase of slow type I fibers comparing with both normal and TR $\beta$  knockout mice (TR $\beta$ <sup>-/-</sup>). Mice lacking of both TRs (TR $\alpha$ <sup>-/-</sup>) showed even more pronounce phenotype than the TR $\alpha$ <sup>-/-</sup> [22] (Table 1). Skeletal muscles from TR $\alpha$ <sup>-/-</sup> mice had a 20-60% longer contraction and relaxation time compared with ones from TR $\beta$ <sup>-/-</sup> and wild type animals [43]. This data can be explained by distribution patterns of TRs in tissue types [12] and/or tissue specific effect of TR $\alpha$  and TR $\beta$ .

On the other hand, mice over expressing p43, the mitochondrial T3 receptor, showed increased mitochondrial biogenesis and oxidative metabolism in the skeletal muscle, switching from the fast MHC II $\alpha$  to the slow type I fiber in soleus muscles [44]. All of these data delineate the crucial contribution of thyroid hormones to skeletal muscle plasticity.

### Thyroid hormones and metabolism

T3 is a well-known regulator of thermogenesis and lipid metabolism. TR $\alpha$  has been shown to be important for the metabolic phenotype [45-47]. Interestingly, the increased metabolic rate in skeletal muscle has been described in humans with resistance to TR $\beta$ , which is characterized by elevated levels of circulating thyroid hormones and unopposed TR $\alpha$  action [48].

THs regulate glycolytic and oxidative pathways in skeletal muscle. T3 induces an overall shift to faster muscle fibers, displaying a reduced mitochondrial density with the predominant glycolytic metabolism. T3-dependent stimulation of SERCA and myosin expression increases energy turnover and generation of heat during activity [35]. At the same time T3 regulates proliferator-activated receptor  $\gamma$  coactivator 1 $\alpha$  (PGC-1 $\alpha$ ) that is crucial for mitochondrial biogenesis through a specific mitochondrial thyroid receptor isoforms (p43) [49,50]. Over expression of p43 *in vivo* increases mitochondrial DNA synthesis, respiration and a switch to slow muscle fibers with reduced glycolytic metabolism and increased oxidative phenotype [44].

Mitochondrial Uncoupling Protein 3 (UCP3) has been suggested to promote energy expenditure in skeletal muscle and is stimulated by thyroid hormone [51]. T3 activates the expression of UCP3 in mice and humans [52]. Administration of thyroid hormone increases UCP3 expression in skeletal muscle and raises the resting metabolic rate in mice [53]. Moreover, T3 directly stimulates the muscle glycerol-3-phosphate dehydrogenase that is highly present in mitochondria of the fast muscle fibers [54] and crucial for T3-mediated stimulation of oxygen consumption.

In addition to metabolic rate and energy expenditure, THs regulate glucose homeostasis. Both hyperthyroidism and

hypothyroidism has been linked to insulin resistance [55]. GLUT4 is the major glucose transporter in the skeletal muscle [56] and plays a pivotal role in modulating insulin-stimulated glucose transporter. Interestingly it has been reported that GLUT4 gene contains a thyroid hormone receptor element in its promoter region [57]. Congenital hypothyroidism impairs GLUT4 expression in skeletal muscle [58], which can be reversed by T3 administration resulting in increase of GLUT4 expression and translocation to the plasma membrane in rat skeletal muscle [59]. In addition, T3 induces uptake of fatty acid in skeletal muscle that can be translated to the increased utilization of fatty acid and reduced deposits of tissue triglyceride [60].

On the other hand the role of DIO2 in skeletal muscle metabolism is contradictory. DIO2 (-/-) mice (table 1) has been shown to be insulin resistant [61] and myoblasts with DIO2 deficiency show impaired action of insulin [62]; however a mouse model that lacks of DIO2 only in the skeletal muscle did not show any significant changes of metabolic phenotype compared with wild type animals, indicating that DIO2 in muscle does not have a significant metabolic role [63].

### CONCLUSION

Thyroid hormones are important regulators of gene expression and the importance of thyroid hormone signaling in skeletal muscle physiology is well known. Classically, thyroid hormone action in skeletal muscle is mostly recognized for gene regulation and metabolism. With knowledge gained from animal models with altered thyroid hormone signaling, a more complex picture can be merged where thyroid hormones are crucial regulator of skeletal muscle homeostasis from development to post-natal maintenance, regeneration and functions.

### ACKNOWLEDGMENT

This work was supported by NIH grant NIDDKD 1K08DK097295 to A.M.

### REFERENCES

1. Mishra A, Zhu XG, Ge K, Cheng SY. Adipogenesis is differentially impaired by thyroid hormone receptor mutant isoforms. *J Mol Endocrinol.* 2010; 44: 247-255.
2. Qiu J, Ma XL, Wang X, Chen H, Huang BR. Insulin-like growth factor binding protein-6 interacts with the thyroid hormone receptor alpha1 and modulates the thyroid hormone-response in osteoblastic differentiation. *Mol Cell Biochem.* 2012; 361: 197-208.
3. Chatonnet F, Picou F, Fauquier T, Flamant F. Thyroid hormone action in cerebellum and cerebral cortex development. *J Thyroid Res.* 2011; 2011: 145762.
4. Nwoye L, Mommaerts WF, Simpson DR, Seraydarian K, Marusich M. Evidence for a direct action of thyroid hormone in specifying muscle properties. *Am J Physiol.* 1982; 242: R401-408.
5. Muller A, van der Linden GC, Zuidwijk MJ, Simonides WS, van der Laarse WJ, van Hardeveld C. Differential effects of thyroid hormone on the expression of sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase isoforms in rat skeletal muscle fibers. *Biochem Biophys Res Commun.* 1994; 203: 1035-1042.
6. Clément K, Viguier N, Diehn M, Alizadeh A, Barbe P, Thalamas C, et al. *In vivo* regulation of human skeletal muscle gene expression by thyroid hormone. *Genome Res.* 2002; 12: 281-291.

7. Brent GA. Mechanisms of thyroid hormone action. *J Clin Invest.* 2012; 122: 3035-3043.
8. Udayakumar N, Rameshkumar AC, Srinivasan AV. Hoffmann syndrome: presentation in hypothyroidism. *J Postgrad Med.* 2005; 51: 332-333.
9. Ramsay ID. Muscle dysfunction in hyperthyroidism. *Lancet.* 1966; 2: 931-934.
10. Cheng SY. Multiple mechanisms for regulation of the transcriptional activity of thyroid hormone receptors. *Rev Endocr Metab Disord.* 2000; 1: 9-18.
11. Forrest D, Sjöberg M, Vennström B. Contrasting developmental and tissue-specific expression of alpha and beta thyroid hormone receptor genes. *EMBO J.* 1990; 9: 1519-1528.
12. White P, Dauncey MJ. Differential expression of thyroid hormone receptor isoforms is strikingly related to cardiac and skeletal muscle phenotype during postnatal development. *J Mol Endocrinol.* 1999; 23: 241-254.
13. Casas F, Rochard P, Rodier A, Cassar-Malek I, Marchal-Victorion S, Wiesner RJ, et al. A variant form of the nuclear triiodothyronine receptor c-ErbAalpha1 plays a direct role in regulation of mitochondrial RNA synthesis. *Mol Cell Biol.* 1999; 19: 7913-7924.
14. Callebaut I, Curcio-Morelli C, Mornon JP, Gereben B, Buettner C, Huang S, et al. The iodothyronine selenodeiodinases are thioredoxin-fold family proteins containing a glycoside hydrolase clan GH-A-like structure. *J Biol Chem.* 2003; 278: 36887-36896.
15. Dentice M, Marsili A, Ambrosio R, Guardiola O, Sibilio A, Paik JH, et al. The FoxO3/type 2 deiodinase pathway is required for normal mouse myogenesis and muscle regeneration. *J Clin Invest.* 2010; 120: 4021-4030.
16. Salvatore D, Simonides WS, Dentice M, Zavacki AM, Larsen PR. Thyroid hormones and skeletal muscle--new insights and potential implications. *Nat Rev Endocrinol.* 2014; 10: 206-214.
17. Downes M, Griggs R, Atkins A, Olson EN, Muscat GE. Identification of a thyroid hormone response element in the mouse myogenin gene: characterization of the thyroid hormone and retinoid X receptor heterodimeric binding site. *Cell Growth Differ.* 1993; 4: 901-909.
18. Muscat GE, Mynett-Johnson L, Dowhan D, Downes M, Griggs R. Activation of myoD gene transcription by 3,5,3'-triiodo-L-thyronine: a direct role for the thyroid hormone and retinoid X receptors. *Nucleic Acids Res.* 1994; 22: 583-591.
19. Sugie H, Verity MA. Postnatal histochemical fiber type differentiation in normal and hypothyroid rat soleus muscle. *Muscle Nerve.* 1985; 8: 654-660.
20. Butler-Browne GS, Herlicoviez D, Whalen RG. Effects of hypothyroidism on myosin isozyme transitions in developing rat muscle. *FEBS Lett.* 1984; 166: 71-75.
21. Butler-Browne GS, Barbet JP, Thornell LE. Myosin heavy and light chain expression during human skeletal muscle development and precocious muscle maturation induced by thyroid hormone. *Anat Embryol (Berl).* 1990; 181: 513-522.
22. Yu F, Göthe S, Wikström L, Forrest D, Vennström B, Larsson L. Effects of thyroid hormone receptor gene disruption on myosin isoform expression in mouse skeletal muscles. *Am J Physiol Regul Integr Comp Physiol.* 2000; 278: R1545-1554.
23. Adams GR, McCue SA, Zeng M, Baldwin KM. Time course of myosin heavy chain transitions in neonatal rats: importance of innervation and thyroid state. *Am J Physiol.* 1999; 276: R954-961.
24. Mauro A. Satellite cell of skeletal muscle fibers. *J Biophys Biochem Cytol.* 1961; 9: 493-495.
25. Zammit PS, Heslop L, Hudon V, Rosenblatt JD, Tajbakhsh S, Buckingham ME, et al. Kinetics of myoblast proliferation show that resident satellite cells are competent to fully regenerate skeletal muscle fibers. *Exp Cell Res.* 2002; 281: 39-49.
26. Chargé SB, Rudnicki MA. Cellular and molecular regulation of muscle regeneration. *Physiol Rev.* 2004; 84: 209-238.
27. Seale P, Sabourin LA, Girgis-Gabardo A, Mansouri A, Gruss P, Rudnicki MA. Pax7 is required for the specification of myogenic satellite cells. *Cell.* 2000; 102: 777-786.
28. Sambasivan R, Yao R, Kissenpfennig A, Van Wittenberghe L, Paldi A, Gayraud-Morel B, et al. Pax7-expressing satellite cells are indispensable for adult skeletal muscle regeneration. *Development.* 2011; 138: 3647-3656.
29. Lepper C, Partridge TA, Fan CM. An absolute requirement for Pax7-positive satellite cells in acute injury-induced skeletal muscle regeneration. *Development.* 2011; 138: 3639-3646.
30. Marchal S, Cassar-Malek I, Pons F, Wrutniak C, Cabello G. Triiodothyronine influences quail myoblast proliferation and differentiation. *Biol Cell.* 1993; 78: 191-197.
31. Busson M, Daury L, Seyer P, Grandemange S, Pessemesse L, Casas F, et al. Avian MyoD and c-Jun coordinately induce transcriptional activity of the 3,5,3'-triiodothyronine nuclear receptor c-ErbAalpha1 in proliferating myoblasts. *Endocrinology.* 2006; 147: 3408-3418.
32. Daury L, Busson M, Casas F, Cassar-Malek I, Wrutniak-Cabello C, Cabello G. The triiodothyronine nuclear receptor c-ErbAalpha1 inhibits avian MyoD transcriptional activity in myoblasts. *FEBS Lett.* 2001; 508: 236-240.
33. Pessemesse L, Schlernitzauer A, Sar C, Levin J, Grandemange S, Seyer P, et al. Depletion of the p43 mitochondrial T3 receptor in mice affects skeletal muscle development and activity. *FASEB J.* 2012; 26: 748-756.
34. Casas F, Pessemesse L, Grandemange S, Seyer P, Baris O, Gueguen N, et al. Overexpression of the mitochondrial T3 receptor induces skeletal muscle atrophy during aging. *PLoS One.* 2009; 4: e5631.
35. Simonides WS, van Harveldt C. Thyroid hormone as a determinant of metabolic and contractile phenotype of skeletal muscle. *Thyroid.* 2008; 18: 205-216.
36. Diffie GM, Haddad F, Herrick RE, Baldwin KM. Control of myosin heavy chain expression: interaction of hypothyroidism and hindlimb suspension. *Am J Physiol.* 1991; 261: C1099-1106.
37. Gambke B, Lyons GE, Haselgrove J, Kelly AM, Rubinstein NA. Thyroidal and neural control of myosin transitions during development of rat fast and slow muscles. *FEBS Lett.* 1983; 156: 335-339.
38. Izumo S, Nadal-Ginard B, Mahdavi V. All members of the MHC multigene family respond to thyroid hormone in a highly tissue-specific manner. *Science.* 1986; 231: 597-600.
39. Ruff RL, Weissmann J. Endocrine myopathies. *Neurol Clin.* 1988; 6: 575-592.
40. Kim TJ, Lee HS, Shin JY, Kim DG, Kim SM, Sung JJ, et al. A case of thyrotoxic myopathy with extreme type 2 fiber predominance. *Exp Neurol.* 2013; 22: 232-234.
41. Caiozzo VJ, Haddad F, Baker M, McCue S, Baldwin KM. MHC polymorphism in rodent plantaris muscle: effects of mechanical overload and hypothyroidism. *Am J Physiol Cell Physiol.* 2000; 278: C709-717.
42. Simonides WS, Brent GA, Thelen MH, van der Linden CG, Larsen PR, van

- Hardevelde C. Characterization of the promoter of the rat sarcoplasmic endoplasmic reticulum Ca<sup>2+</sup>-ATPase 1 gene and analysis of thyroid hormone responsiveness. *J Biol Chem.* 1996; 271: 32048-32056.
43. Johansson C, Lännergren J, Lunde PK, Vennström B, Thorén P, Westerblad H. Isometric force and endurance in soleus muscle of thyroid hormone receptor-alpha(1)- or -beta-deficient mice. *Am J Physiol Regul Integr Comp Physiol.* 2000; 278: R598-603.
44. Casas F, Pesseme L, Grandemange S, Seyer P, Gueguen N, Baris O, et al. Overexpression of the mitochondrial T3 receptor p43 induces a shift in skeletal muscle fiber types. *PLoS One.* 2008; 3: e2501.
45. Vennström B, Mittag J, Wallis K. Severe psychomotor and metabolic damages caused by a mutant thyroid hormone receptor alpha 1 in mice: can patients with a similar mutation be found and treated? *Acta Paediatr.* 2008; 97: 1605-1610.
46. Liu YY, Brent GA. Thyroid hormone crosstalk with nuclear receptor signaling in metabolic regulation. *Trends Endocrinol Metab.* 2010; 21: 166-173.
47. Ribeiro MO, Bianco SD, Kaneshige M, Schultz JJ, Cheng SY, Bianco AC, et al. Expression of uncoupling protein 1 in mouse brown adipose tissue is thyroid hormone receptor-beta isoform specific and required for adaptive thermogenesis. *Endocrinology.* 2010; 151: 432-440.
48. Mitchell CS, Savage DB, Dufour S, Schoenmakers N, Murgatroyd P, Befroy D, et al. Resistance to thyroid hormone is associated with raised energy expenditure, muscle mitochondrial uncoupling, and hyperphagia. *J Clin Invest.* 2010; 120: 1345-1354.
49. Weitzel JM, Iwen KA, Seitz HJ. Regulation of mitochondrial biogenesis by thyroid hormone. *Exp Physiol.* 2003; 88: 121-128.
50. Psarra AM, Solakidi S, Sekeris CE. The mitochondrion as a primary site of action of steroid and thyroid hormones: presence and action of steroid and thyroid hormone receptors in mitochondria of animal cells. *Mol Cell Endocrinol.* 2006; 246: 21-33.
51. Lanni A, Moreno M, Lombardi A, Goglia F. Thyroid hormone and uncoupling proteins. *FEBS Lett.* 2003; 543: 5-10.
52. Solanes G, Pedraza N, Calvo V, Vidal-Puig A, Lowell BB, Villarroya F. Thyroid hormones directly activate the expression of the human and mouse uncoupling protein-3 genes through a thyroid response element in the proximal promoter region. *Biochem J.* 2005; 386: 505-513.
53. Gong DW, He Y, Karas M, Reitman M. Uncoupling protein-3 is a mediator of thermogenesis regulated by thyroid hormone, beta3-adrenergic agonists, and leptin. *J Biol Chem.* 1997; 272: 24129-24132.
54. Dümmler K, Müller S, Seitz HJ. Regulation of adenine nucleotide translocase and glycerol 3-phosphate dehydrogenase expression by thyroid hormones in different rat tissues. *Biochem J.* 1996; 317: 913-918.
55. Chidake A, Mentuccia D, Celi FS. Peripheral metabolism of thyroid hormone and glucose homeostasis. *Thyroid.* 2005; 15: 899-903.
56. Zorzano A, Palacín M, Gumà A. Mechanisms regulating GLUT4 glucose transporter expression and glucose transport in skeletal muscle. *Acta Physiol Scand.* 2005; 183: 43-58.
57. Torrance CJ, Devente JE, Jones JP, Dohm GL. Effects of thyroid hormone on GLUT4 glucose transporter gene expression and NIDDM in rats. *Endocrinology.* 1997; 138: 1204-1214.
58. Ramos S, Goya L, Alvarez C, Martín MA, Agote M, Escrivá F, et al. Different role of insulin in GLUT-1 and -4 regulation in heart and skeletal muscle during perinatal hypothyroidism. *Am J Physiol Endocrinol Metab.* 2001; 281: E1073-1081.
59. Brunetto EL, Teixeira Sda S, Giannocco G, Machado UF, Nunes MT. T3 rapidly increases SLC2A4 gene expression and GLUT4 trafficking to the plasma membrane in skeletal muscle of rat and improves glucose homeostasis. *Thyroid.* 2012; 22: 70-79.
60. Lombardi A, De Matteis R, Moreno M, Napolitano L, Busiello RA, Senese R, et al. Responses of skeletal muscle lipid metabolism in rat gastrocnemius to hypothyroidism and iodothyronine administration: a putative role for FAT/CD36. *Am J Physiol Endocrinol Metab.* 2012; 303: E1222-1233.
61. Marsili A, Aguayo-Mazzucato C, Chen T, Kumar A, Chung M, Lunsford EP, et al. Mice with a targeted deletion of the type 2 deiodinase are insulin resistant and susceptible to diet induced obesity. *PLoS One.* 2011; 6: e20832.
62. Grozovsky R, Ribich S, Rosene ML, Mulcahey MA, Huang SA, Patti ME, et al. Type 2 deiodinase expression is induced by peroxisomal proliferator-activated receptor-gamma agonists in skeletal myocytes. *Endocrinology.* 2009; 150: 1976-1983.
63. Fonseca TL, Werneck-De-Castro JP, Castillo M, Bocco BM, Fernandes GW, McAninch EA, et al. Tissue-specific inactivation of type 2 deiodinase reveals multilevel control of fatty acid oxidation by thyroid hormone in the mouse. *Diabetes.* 2014; 63: 1594-1604.

## Cite this article

Lee JW, Kim NH, Milanesi A (2014) Thyroid Hormone Signaling in Muscle Development, Repair and Metabolism. *J Endocrinol Diabetes Obes* 2(3): 1046.