

## Review Article

# Rasmussen's Encephalitis: Clinical Features and Mechanisms Advances

Tianfu Li<sup>1,3,4\*</sup>, Qing Gao<sup>3</sup> and Guoming Luan<sup>2,3,4</sup><sup>1</sup>Department of Neurology, Capital Medical University, China<sup>2</sup>Department of Neurosurgery, Capital Medical University, China<sup>3</sup>Beijing Key laboratory of Epilepsy, China<sup>4</sup>Center of Epilepsy, Beijing Institute for Brain Disorders, China

## \*Corresponding author

Tianfu Li, Beijing Sanbo Brain Hospital, Capital Medical University, Xiangshan Yikesong 50, Haidian district, Beijing, 100093, China, Tel: 86 1062856761; Fax: 86-10-62856902; Email: tianfuli66@126.com

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## Abstract

Rasmussen's encephalitis (RE) is neurological disorder of childhood characterized by uni-hemispheric inflammation, intractable focal epilepsy and progressive cognitive and neurological deficits. Currently, hemispherectomy is the only effective method to control the seizures associated with RE. Although this disease has been heavily investigated, the pathogenesis of RE with unilateral cortex atrophy and focal seizure is still enigmatic. Neuropathological and immunological studies support the hypothesis that destruction of neurons and astrocytes by cytotoxic CD8 T cells as a pathogenic mechanism underlying this enigmatic disorder. Recently data indicated that intrinsic activation of endogenous pro-inflammation high-mobility group box-1 (HMGB1) and Toll-like receptor (TLR) pathways, and dysregulation of adenosinergic mechanism are involved in the development of epilepsy, which suggest the specific targets in the treatment of epilepsy, inflammation and cognitive deterioration associated with epilepsy in RE patients.

## INTRODUCTION

Rasmussen's encephalitis (RE) was initially described by neurosurgeon Theodore Rasmussen and his colleagues from the Montreal Neurological Institute in the late 1950s [1]. From then on, extensive research on the clinical features, neuropathology, mechanisms and therapy of RE was carried out, and the 2005 European consensus on pathogenesis, diagnosis, and treatment of RE was achieved and still remains accepted guideline for evaluative criteria [2,3]. RE is a very rare chronic progressive inflammatory neurological disorder of uncertain etiology affecting mostly children and associated with hemispheric atrophy, pharmacoresistant focal epilepsy (epilepsia partialis continua), cognitive deterioration and progressive cognitive and neurological deficits, resulting from progressive loss of function subserved by the involved cerebral hemisphere [1,2,4,5]. An intriguing feature of RE is the restriction of the inflammatory process to one brain hemisphere, setting it apart from any other inflammatory disease of the CNS. The aetiology and pathogenesis of RE, in particular, the factors responsible for the characteristic of asymmetry are still elusive. The neuropathological hallmarks of RE consist of lymphocytic infiltrates (perivascular lymphocytic cuffing), microglial nodules, neuronal destruction, and gliosis of the affected hemisphere [6,7]. Currently, targeted therapeutic strategies remain elusive and hemispherectomy is the only

effective method to control the seizures associated with RE. Recent results demonstrate that activation of endogenous high-mobility group box-1 (HMGB1) and Toll-like receptor [8], and adenosine system dysfunction in epileptic tissue [9], which may play a role in the generation of seizures, and possibly epileptogenesis itself in RE. Here, we will review the current basic and clinical research associated with RE patients.

## Pathogenesis of RE

Over fifty years, extensive studies have been carried out to attempt to elucidate the pathogenesis of RE. The main hypothesis on the pathogenesis of RE as follows:

**Virus Infection:** The hypothesis on virus infection in RE suggested by Rasmussen, is that RE may be associated with constituents of the immune reaction in the brain following a virus infection, such as lymphocyte infiltration and microglial nodules. Support of Rasmussen's idea comes from studies linking cytomegalovirus and herpes simplex virus [1]. Nevertheless, using the method of polymerase chain reaction (PCR) and/or in situ hybridization, results from other research groups demonstrate that the virus DNA sequence existed within the tissue of epilepsy patients associated with other central nervous system (CNS) diseases such as focal cortical dysplasia, gangliocytoma and cerebromalacia [7,10]. Several viruses,

including Epstein-Barr virus, cytomegalovirus, herpes simplex virus, and enterovirus have been tested for the presence in brain tissue from patients with RE [11-13]. None of these studies was able to show a causal association between Rasmussen's encephalitis and a specific virus. To establish a reliable method to identify the specific virus in RE is challenging, for infected cells under cytotoxic T-cell attack can be injured and later engulfed by microglia and macrophages, in which case the chances of finding a virus most likely occur early in the disease when surgical sampling is less frequent [5]. It is important to name Russian spring-summer meningoencephalitis here, also referred to as the Far Eastern variant of tick-borne encephalitis (TBE-FE). Tick-borne encephalitis virus (TBEV) causes a febrile syndrome that can often be complicated by neurological symptoms. It often leads to a form of encephalitis that affects young people in the summer. Both MRI and EEG abnormalities are often a specific and not diagnostic. Abnormalities in MRI are seen in up to 18% with lesions usually confined to the thalamus, cerebellum, brainstem and caudatus [14]. EEG was abnormal in 65% of cases with evidence of slow abnormalities (theta and delta waves) in leads the center-front [15].

**Antibody-mediated CNS degeneration:** Anti-glutamate receptors GluR3, was the first circulating auto-antibodies identified in RE in 1994 (Rogers et al., 1994). From then on, antibody against GluR3, munc-18 and NMDA receptor-mediated mechanisms dominated RE research for more than 10 years [7, 16-23]. However, the follow-up studies demonstrate that the circulating auto-antibodies, even those highly-specific cell surface-directed antibodies can occasionally be present in patients with neurodegenerative disease, in which case these antibodies are probably secondary to the pathology rather than causative. None of the auto-antibodies have been found in more than a small number of patients with RE, and responses to plasma exchange are unpredictable. Therefore, the role of circulating auto-antibodies in the pathogenesis of RE is still elusive [5].

**T-cell cytotoxicity:** Recent studies have suggested that in contrast to a random attraction of cells as part of a secondary immune response, CD8+ T cell-mediated attack against neurons and astrocytes in the central nervous system is a major component of the pathogenesis of RE [24-26]. Because neurons and astrocytes are attacked by cytotoxic T cells, there may be an auto-antigen, as a driving antigen expressed by both neurons and astrocytes. However, to identify the driving antigen of cytotoxic T cells is still elusive [5].

## Advance of Pathogenesis

**Dysfunction of adenosine system in RE:** Adenosine is an endogenous purine nucleoside that modulates a wide range of physiological functions [27]. Most notable among its many roles is its importance in controlling inflammation [28-30] and inhibiting seizures [31-37] and restoring cognitive function related with epilepsy [38]. Recent evidence illustrated that the adenosine dysfunction in astrogliosis including decrease in the density of A1R [39]. Extracellular levels of adenosine are regulated largely by an astrocyte-based adenosine cycle and astrocytic ADK is the major adenosine-removing enzyme [40]. Minor changes in ADK activity translate rapidly into major changes in adenosine [41]. Therefore, up-regulation of ADK in astrogliosis leads to reducing

the "tone" of ambient adenosine leading to insufficient activation of adenosine receptors [42]. Neuronal excitability in the brain is modulated by activation of G protein-coupled adenosine receptors (A1, A2A, A2B, A3) [27,43]. The receptor expression levels and availability of endogenous adenosine to activate the receptors play a crucial role in neuronal excitability [38]. Adenosine is a neuro-modulator that has been proved to be a major endogenous anticonvulsant acting via A1R. In the brain, adenosine modulates neuronal activity by decreasing presynaptic release of various neurotransmitters, and the most dramatic inhibitory actions are on the glutamatergic system [44]. In addition, adenosine acting through postsynaptic A1Rs may activate K<sup>+</sup> channels, leading to hyperpolarization of postsynaptic neurons and promoting NMDA receptor inhibition [45]. Recent work demonstrated that increased expression of major adenosine-removing enzyme adenosine kinase (ADK) in RE patients plays an important role in the epileptogenesis of RE [9]. Focal astrogliosis and marked expression of ADK were observed in the lesions of RE. Significantly greater ADK expression in RE versus controls was demonstrated by Western blot, and greater enzymatic activity for ADK was demonstrated using an enzyme-coupled bioluminescent assay. ADK has been highlighted as a diagnostic marker to predict epileptogenesis as well as a potential target for anti-epileptogenesis or disease modification [31,32,34,38,46-50]. Therefore, up-regulation of ADK in RE is considered as a common pathologic hallmark of RE and that ADK might be a target in the treatment of epilepsy associated with RE [9]. Neuronal adenosine A1 receptor (A1R) in the lesion area of RE was also observed (unpublished data). Activation of A1R has been proved to prevent the spatial spread of seizures [31,34,39,51-55]. Activation of A1R signaling may be related to the clinical features of the RE:

i) unilateral epileptic discharge in RE patients with or without uni-hemispheric slowing in long-term video EEG, and EPC is not always accompanied by visually recognizable ictal surface EEG activity; ii) both seizures and inflammation atrophy are seen only in one cerebral hemisphere, rarely spreading to the contra-hemisphere, only four out of the roughly 200-300 published cases of Rasmussen's encephalitis had evidence of bilateral disease on histopathology; iii) most of the clinical seizure types are focal seizures (SPS or CPS) with or without EPC, and notably, secondary generalized epilepsy continua from EPC has rarely been reported. Thereby, adenosine augmenting therapeutic strategies with anti-convulsion, anti-inflammation, and anti-cognitive dysfunction effects might be the ideal treatment for RE.

**Activation of HMGB1-TLR-RAGE signaling in RE:** Novel evidence of intrinsic activation of pro-inflammation signaling, endogenous high-mobility group box-1 (HMGB1), toll-like receptor (TLR) and RAGE in human Rasmussen's encephalitis has been reported recently [8]. HMGB-1 is a 29 kDa DNA-binding protein with a highly conserved structure in several species [56]. HMGB-1 participates in nucleosome formation and regulation of gene transcription [57,58]. Including proinflammatory genes [59]. In response to inflammatory stimuli, HMGB-1 is secreted by activated macrophages [60], natural killer cells [61], myeloid dendritic cells [62] and astrocytes [63] binding to the receptor for advanced glycation end products (RAGE) and other receptors, including TLR2 and TLR4 [64,65]. HMGB1 acts as a "danger signal" and alerts the immune system to damaged or dying cells.

The hyper acetylated form of HMGB1 regulates transcription of various pro-inflammatory cytokines through binding to TLR2, TLR4 and also to RAGE [59,66]. HMGB1-TLR-RAGE may represent a novel pro-inflammatory axis following sterile brain injury [67].

In addition to exert a pro-inflammatory role, HMGB1-TLR-RAGE signaling plays a critical role in determining the pathological outcomes epilepsy [63, 68-71]. Although HMGB1 release and signaling may be a general feature of all epilepsies [63,69,71], expression of HMGB1, TLR2, TLR4 and RAGE was more markedly increased in perivascular areas, and endothelial cells in walls of blood vessels within the lesions cortex displayed immune reactivity in RE patients [8] (Figure 1). These findings are concomitant with the reactive astrogliosis, neuron loss and inflammation (i.e. CD8-positive, CD3-positive T lymphocyte). The evidence further supported the role of HMGB1-TLR pathway in activation of immune and endothelial cells in the pathogenesis of RE. Anti-inflammatory agents targeting on HMGB1-TLRs-RAGE may also prove beneficial in alleviating some of the common comorbidities associated with chronic epilepsy, including cognitive dysfunction and memory deficits [72-74], which will provide benefits for RE in three ways: anti-inflammation, anti-epilepsy and improve cognitive dysfunction associated with epilepsy [8].

### Clinical features

Rasmussen's encephalitis is a progressive disease characterized by drug-resistant focal epilepsy, progressive

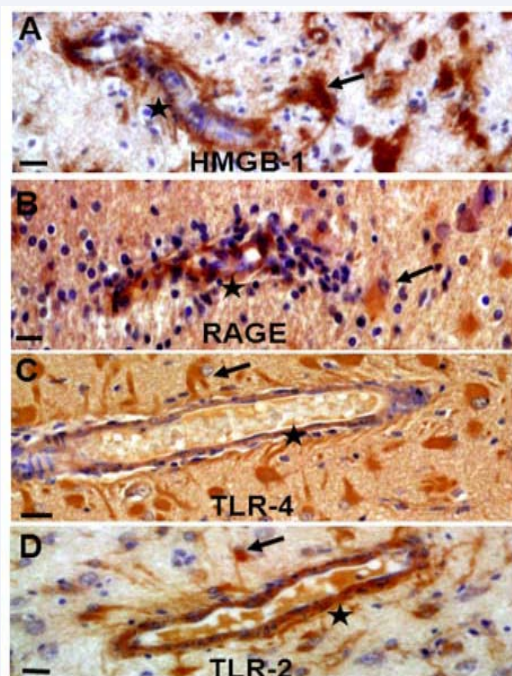
hemiplegia, and cognitive deterioration, with unihemispheric brain atrophy [3-5, 75,76]. The disorder is rare and affects mostly children or young adults, and the median age of onset is 6 years and the range from infancy to adulthood [4,77]. The typical clinical course of RE has been proposed last century [4]. Stage 1 (prodromal phase) is characterized by a non-specific low seizure frequency and, in rare instances, some degree of hemiparesis with a average duration of 7.1 months (range 0 to 8.1 years) and even longer duration in adolescent and adult patients compared with the children. Stage 2 (acute stage) is characterized by frequent simple partial motor seizures, and EPC in 69% of cases. The median duration of this stage was 8 months (range 4-8 months). During the acute stage the neurological deficits appeared including progressive hemiparesis, cognitive deterioration, and aphasia if dominant hemisphere affected. The last stage (residual stage) is characterized by continuing seizures with a decrease in seizure frequency, notably, with permanent and stable neurological deficit [4].

### Neuroimaging of RE

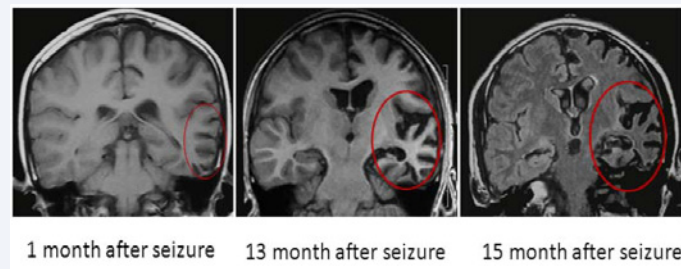
Magnetic resonance imaging (MRI) of the brain has become a key tool for diagnostic assessment, especially provides the evidence of the diagnosis in the early stages and the progression of RE [78]. MRI findings of RE patients usually indicate a progressive unihemispheric focal cortical atrophy with mild or severe enlargement of the lateral ventricle (Figure 2). The majority of patients exhibit unilateral enlargement of the inner and outer CSF compartments in the insular and periinsular regions during the early stage [78,79]. In majority of RE patients, ipsilateral moderate atrophy of the head of the caudate nucleus is a typical alteration of MRI and regarded as a sign for early stage of RE, but not an invariable accompanying feature of hemispheric atrophy [78]. Even in the early stage of RE, normal findings of MRI scans or scans with gadolinium enhancement are very rare in RE patients [4,78,80,81]. Functional studies using  $^{18}\text{F}$ -FDG positron emission tomography (PET) illustrates diffuse unilateral cerebral hypo metabolism with in the affected hemisphere [82]. In cases of RE with no significant finding in brain MRI, PET and SPECT may provide helpful information to select the right site for brain biopsy.

### EEG

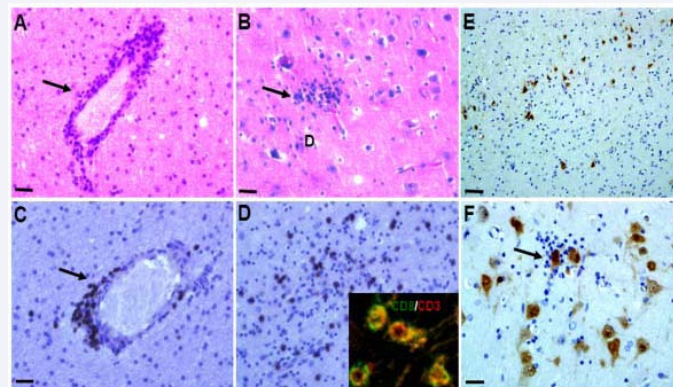
Overall, no specific EEG changes identified to differentiate RE from other causes of focal epilepsy such as focal cortical dysplasia [83]. Long-term video EEG results were abnormal in all cases and generally expressed slow waves in the affected hemisphere. Polymorphic delta waves and multifocal unilateral epileptic discharge interictal EEG were mainly observed in temporal and central locations. In the majority of patients, contra lateral asynchronous slow waves and epilepticform discharges occurred [84]. However, few ictal patterns were ever recorded from contra lateral electrodes [85]. Up to now, only four patients have been reported to have bilateral RE [86]. Epilepticform abnormalities are frequent and they often develop into electrographic seizures, however, EPC in RE is not always accompanied by rhythmic EEG discharges on surface EEG [5]. Emerging persistent delta activity over the affected hemisphere with contra lateral normal background rhythms, followed in due



**Figure 1** HMGB-1, RAGE, TLR-4 and TLR-2 immuno reactivity in RE. Nuclear and cytoplasmic staining showed in perivascular glial cells (A, arrows) and marked cytoplasmic staining of RAGE (B, arrows), TLR-4 (C, arrows), TLR-2 (D, arrows). Endothelial cells within the lesions cortex displayed HMGB1 (A, stars), RAGE (B, stars), TLR-4 (C, stars) and TLR-2 immuno reactivity (D, stars). Scale bar: 12.5 μm.



**Figure 2** Neuroimaging in RE. (A-C): The brain MRI displayed a typical progressive abnormal signal area expansion and atrophy of the left hemisphere, with mild enlargement of the left lateral ventricle (1 months, 13 months, 15 months after the first seizure onset).



**Figure 3** Neuropathological features in RE. (A): Perivascular lymphocytic cuffing in the cerebral cortex (arrows, HE staining). (B): formation of microglial nodules and diffuse microglial activation (arrows, HE staining). (C): Perivascular lymphocytic cuffing in the cerebral cortex (arrows, CD8 staining). (D): parenchymal lymphocytic in the cerebral cortex (arrows, CD8 staining). Inset in D: co-localization of CD3 and CD8. (E): neuronal loss in the lesion cortex of RE (NeuN staining). (F): Cortical neuron with neuronophagia (arrows, NeuN staining). Scale bar: A, B, C, D, F: 25  $\mu$ m; E: 100  $\mu$ m.

course by independent interictal epileptiform abnormalities over the unaffected hemisphere highly indicate the diagnosis of RE as the condition evolves, and highlight as the marker of overall cognitive deterioration [83].

### Characteristic of neuropathology

The neuropathologic hallmarks of RE include inflammation (perivascular lymphocytic cuffing, microglial nodules, leptomeningeal infiltrates), neuronal loss, and astrogliosis confined to one cerebral hemisphere (Figure 3) [1,6,7,87]. Microglial and lymphocytic nodules and perivascular cuffing, neuronal death, and neuronophagia are the most common pathological features. T cells aggregating around the perivascular and infiltrating the meninges are as the majority of CD3<sup>+</sup>CD8<sup>+</sup> cells. During the residual stage the pathological features include cortical cavitation, marked astrogliosis, and neuronal cell loss. Recently evidence illustrates dual pathology, with the finding of focal cortical dysplasia, tuberous sclerosis, low grade tumor, vascular abnormalities or old ischaemic lesions in association with Rasmussen's encephalitis [4,88-92]. Inflammation is usually multifocal within the hemisphere and progressive, with the characteristic of an area of pronounced cortical damage is often surrounded by normal cerebral cortex or milder stages of inflammation, which illustrate the reason why a biopsy results sometimes mislead in the diagnosis [5].

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### REFERENCES

1. Rasmussen T, Olszewski J, Lloydsmith D. Focal seizures due to chronic localized encephalitis. *Neurology*. 1958; 8: 435-445.
2. Bien CG, Granata T, Antozzi C, Cross JH, Dulac O, Kurthen M, et al. Pathogenesis, diagnosis and treatment of Rasmussen encephalitis: a European consensus statement. *Brain*. 2005; 128: 454-471.
3. Olson HE, Lechpammer M, Prabhu SP, Ciarlini PD, Poduri A, Gooty VD, et al. Clinical application and evaluation of the Bien diagnostic criteria for Rasmussen encephalitis. *Epilepsia*. 2013; 54: 1753-1760.
4. Bien CG, Widman G, Urbach H, Sassen R, Kuczaty S, Wiestler OD, et al. The natural history of Rasmussen's encephalitis. *Brain*. 2002; 125: 1751-1759.
5. Varadkar S, Bien CG, Kruse CA, Jensen FE, Bauer J, Pardo CA, et al. Rasmussen's encephalitis: clinical features, pathobiology, and

- treatment advances. *Lancet Neurol.* 2014; 13: 195-205.
6. Farrell MA, Droogan O, Secor DL, Poukens V, Quinn B, Vinters HV, et al. Chronic encephalitis associated with epilepsy: immunohistochemical and ultrastructural studies. *Acta Neuropathol.* 1995; 89: 313-321.
  7. Rogers SW, Andrews PI, Gahring LC, Whisenand T, Cauley K, Crain B, et al. Autoantibodies to glutamate receptor GluR3 in Rasmussen's encephalitis. *Science.* 1994; 265: 648-51.
  8. Luan G, Gao Q, Zhai F, Chen Y, Li T. Upregulation of HMGB1, toll-like receptor and RAGE in human Rasmussen's encephalitis. *Epilepsy Res.* 2016; 123: 36-49.
  9. Luan G, Gao Q, Guan Y, Zhai F, Zhou J, Liu C, et al. Upregulation of adenosine kinase in Rasmussen encephalitis. *J Neuropathol Exp Neurol.* 2013; 72: 1000-1008.
  10. Jay V, Becker LE, Blaser S, Hwang P, Hoffman HJ, Humphreys R, et al. Pathology of chronic herpes infection associated with seizure disorder: a report of two cases with tissue detection of herpes simplex virus 1 by the polymerase chain reaction. *Pediatr Pathol Lab Med.* 1995; 15: 131-146.
  11. Walter GF, Renella RR. Epstein-Barr virus in brain and Rasmussen's encephalitis. *Lancet.* 1989; 1: 279-280.
  12. Friedman H, Ch'ien L, Parham D. Virus in brain of child with hemiplegia, hemiconvulsions, and epilepsy. *Lancet.* 1977 Sep 24; 2: 666.
  13. Power C, Poland SD, Blume WT, Girvin JP, Rice GP. Cytomegalovirus and Rasmussen's encephalitis. *Lancet.* 1990; 336: 1282-1284.
  14. Marjelund S, Tikkakoski T, Tuisku S, Räisänen S. Magnetic resonance imaging findings and outcome in severe tick-borne encephalitis. Report of four cases and review of the literature. *Acta Radiol.* 2004; 45: 88-94.
  15. Zambito MS, Pistacchi M, Gioulis M, Mel R, Marchini C, Francavilla E, et al. Neurological complications of tick borne encephalitis: the experience of 89 patients studied and literature review. *Neurol Sci.* 2014; 35: 15-21.
  16. Yang R, Puranam RS, Butler LS, Qian WH, He XP, Moyer MB, et al. Autoimmunity to munc-18 in Rasmussen's encephalitis. *Neuron.* 2000; 28: 375-383.
  17. Alvarez-Barón E, Bien CG, Schramm J, Elger CE, Becker AJ, Schoch S, et al. Autoantibodies to Munc18, cerebral plasma cells and B-lymphocytes in Rasmussen encephalitis. *Epilepsy Res.* 2008; 80: 93-97.
  18. Granata T, Fusco L, Gobbi G, Freri E, Ragona F, Broggi G, et al. Experience with immunomodulatory treatments in Rasmussen's encephalitis. *Neurology.* 2003; 61: 1807-1810.
  19. Mantegazza R, Bernasconi P, Baggi F, Spreafico R, Ragona F, Antozzi C, et al. Antibodies against GluR3 peptides are not specific for Rasmussen's encephalitis but are also present in epilepsy patients with severe, early onset. *J Neuroimmunol.* 2002; 131: 179-185.
  20. Watson R, Jepson JE, Bermudez I, Alexander S, Hart Y, McKnight K, et al. Alpha7-acetylcholine receptor antibodies in two patients with Rasmussen encephalitis. *Neurology.* 2005; 65: 1802-1804.
  21. Watson R, Jiang Y, Bermudez I, Houlihan L, Clover L, McKnight K, et al. Absence of antibodies to glutamate receptor type 3 (GluR3) in Rasmussen encephalitis. *Neurology.* 2004; 63: 43-50.
  22. Wiendl H, Bien CG, Bernasconi P, Fleckenstein B, Elger CE, Dichgans J, et al. GluR3 antibodies: prevalence in focal epilepsy but no specificity for Rasmussen's encephalitis. *Neurology.* 2001; 57: 1511-1514.
  23. Greiner H, Leach JL, Lee KH and Krueger DA. Anti-NMDA receptor encephalitis presenting with imaging findings and clinical features mimicking Rasmussen syndrome. *Seizure.* 2011; 20: 266-270.
  24. Schwab N, Bien CG, Waschbisch A, Becker A, Vince GH, Dornmair K, et al. CD8+ T-cell clones dominate brain infiltrates in Rasmussen encephalitis and persist in the periphery. *Brain.* 2009; 132: 1236-1246.
  25. Bien CG, Bauer J, Deckwerth TL, Wiendl H, Deckert M, Wiestler OD, et al. Destruction of neurons by cytotoxic T cells: a new pathogenic mechanism in Rasmussen's encephalitis. *Ann Neurol.* 2002; 51: 311-318.
  26. Bauer J, Elger CE, Hans VH, Schramm J, Urbach H, Lassmann H, et al. Astrocytes are a specific immunological target in Rasmussen's encephalitis. *Ann Neurol.* 2007; 62: 67-80.
  27. Fredholm BB, Ijzerman AP, Jacobson KA, Linden J, Müller CE. International Union of Basic and Clinical Pharmacology. LXXXI. Nomenclature and classification of adenosine receptors--an update. *Pharmacol Rev.* 2011; 63: 1-34.
  28. Mills JH, Kim DG, Krenz A, Chen JF, Bynoe MS. A2A adenosine receptor signaling in lymphocytes and the central nervous system regulates inflammation during experimental autoimmune encephalomyelitis. *J Immunol.* 2012; 188: 5713-5722.
  29. Blackburn MR, Vance CO, Morschl E, Wilson CN. Adenosine receptors and inflammation. *Handb Exp Pharmacol.* 2009; 215-269.
  30. Wen J, Jiang X, Dai Y, Zhang Y, Tang Y, Sun H, et al. Increased adenosine contributes to penile fibrosis, a dangerous feature of priapism, via A2B adenosine receptor signaling. *FASEB J.* 2010; 24: 740-749.
  31. Masino SA, Li T, Theofilas P, Sandau US, Ruskin DN, Fredholm BB, et al. A ketogenic diet suppresses seizures in mice through adenosine A<sub>1</sub> receptors. *J Clin Invest.* 2011; 121: 2679-2683.
  32. Li T, Lytle N, Lan JQ, Sandau US, Boison D. Local disruption of glial adenosine homeostasis in mice associates with focal electrographic seizures: a first step in epileptogenesis. *Glia.* 2012; 60: 83-95.
  33. Li T, Ren G, Kaplan DL, Boison D. Human mesenchymal stem cell grafts engineered to release adenosine reduce chronic seizures in a mouse model of CA3-selective epileptogenesis. *Epilepsy Res.* 2009; 84: 238-241.
  34. Li T, Ren G, Lusardi T, Wilz A, Lan JQ, Iwasato T, et al. Adenosine kinase is a target for the prediction and prevention of epileptogenesis in mice. *J Clin Invest.* 2008; 118: 571-582.
  35. Li T, Steinbeck JA, Lusardi T, Koch P, Lan JQ, Wilz A, et al. Suppression of kindling epileptogenesis by adenosine releasing stem cell-derived brain implants. *Brain.* 2007; 130: 1276-1288.
  36. Wilz A, Pritchard EM, Li T, Lan JQ, Kaplan DL, Boison D, et al. Silk polymer-based adenosine release: therapeutic potential for epilepsy. *Biomaterials.* 2008; 29: 3609-3616.
  37. Li T, Lan JQ, Boison D. Uncoupling of astrogliosis from epileptogenesis in adenosine kinase (ADK) transgenic mice. *Neuron Glia Biol.* 2008; 4: 91-99.
  38. Boison D. Adenosinergic signaling in epilepsy. *Neuropharmacology.* 2016; 104: 131-139.
  39. Glass M, Faull RL, Bullock JY, Jansen K, Mee EW, Walker EB, et al. Loss of A<sub>1</sub> adenosine receptors in human temporal lobe epilepsy. *Brain Res.* 1996; 710: 56-68.
  40. Boison D, Chen JF, Fredholm BB. Adenosine signaling and function in glial cells. *Cell Death Differ.* 2010; 17: 1071-1082.
  41. Boison D. Adenosine kinase, epilepsy and stroke: mechanisms and therapies. *Trends Pharmacol Sci.* 2006; 27: 652-658.
  42. Boison D. The adenosine kinase hypothesis of epileptogenesis. *Prog Neurobiol.* 2008; 84: 249-62.

43. Fredholm BB, Chen JF, Masino SA, Vaugeois JM. Actions of adenosine at its receptors in the CNS: insights from knockouts and drugs. *Annu Rev Pharmacol Toxicol.* 2005; 45: 385-412.
44. Reppert SM, Weaver DR, Stehle JH, Rivkees SA. Molecular cloning and characterization of a rat A1-adenosine receptor that is widely expressed in brain and spinal cord. *Mol Endocrinol.* 1991; 5: 1037-48.
45. Wardas J. Neuroprotective role of adenosine in the CNS. *Pol J Pharmacol.* 2002; 54: 313-326.
46. Luan G, Gao Q, Zhai F, Zhou J, Liu C, Chen Y, et al. Adenosine kinase expression in cortical dysplasia with balloon cells: analysis of developmental lineage of cell types. *J Neuropathol Exp Neurol.* 2015; 74: 132-147.
47. Aronica E, Sandau US, Iyer A, Boison D. Glial adenosine kinase--a neuropathological marker of the epileptic brain. *Neurochem Int.* 2013; 63: 688-695.
48. Boison D. Adenosine dysfunction in epilepsy. *Glia.* 2012; 60: 1234-1243.
49. Boison D. Adenosine kinase: exploitation for therapeutic gain. *Pharmacol Rev.* 2013; 65: 906-943.
50. Li T, Quan Lan J, Fredholm BB, Simon RP, Boison D. Adenosine dysfunction in astrogliosis: cause for seizure generation? *Neuron Glia Biol.* 2007; 3: 353-366.
51. Hamil NE, Cock HR, Walker MC. Acute down-regulation of adenosine A(1) receptor activity in status epilepticus. *Epilepsia.* 2012; 53: 177-188.
52. Fedele DE, Li T, Lan JQ, Fredholm BB, Boison D. Adenosine A1 receptors are crucial in keeping an epileptic focus localized. *Exp Neurol.* 2006; 200: 184-190.
53. Klaft ZJ, Hollnagel JO, Salar S, Caliřkan G, Schulz SB, Schneider UC, et al. Adenosine A1 receptor-mediated suppression of carbamazepine-resistant seizure-like events in human neocortical slices. *Epilepsia* 2016; 57:746-756.
54. Wagner AK, Miller MA, Scanlon J, Ren D, Kochanek PM, Conley YP, et al. Adenosine A receptor gene variants associated with post-traumatic seizures after severe TBI. *Epilepsy Res.* 2010; 90: 259-272.
55. Kochanek PM, Vagni VA, Janesko KL, Washington CB, Crumrine PK, Garman RH, et al. Adenosine A1 receptor knockout mice develop lethal status epilepticus after experimental traumatic brain injury. *J Cereb Blood Flow Metab.* 2006; 26: 565-575.
56. Thomas JO. HMG and 2: architectural DNA-binding proteins. *Biochem Soc Trans.* 2001; 29: 395-401.
57. Park JS, Arcaroli J, Yum HK, Yang H, Wang H, Yang KY, et al. Activation of gene expression in human neutrophils by high mobility group box 1 protein. *Am J Physiol Cell Physiol.* 2003; 284: 870-879.
58. Stros M, Ozaki T, Bacikova A, Kageyama H, Nakagawara A. HMGB1 and HMGB2 cell-specifically down-regulate the p53- and p73-dependent sequence-specific transactivation from the human Bax gene promoter. *J Biol Chem.* 2002; 277: 7157-7164.
59. Bianchi ME, Manfredi AA. Immunology. Dangers in and out. *Science.* 2009; 323: 1683-1684.
60. Bonaldi T, Talamo F, Scaffidi P, Ferrera D, Porto A, Bachi A, et al. Monocytic cells hyperacetylate chromatin protein HMGB1 to redirect it towards secretion. *EMBO J.* 2003; 22: 5551-5560.
61. Semino C, Angelini G, Poggi A, Rubartelli A. NK/iDC interaction results in IL-18 secretion by DCs at the synaptic cleft followed by NK cell activation and release of the DC maturation factor H... *Blood.* 2005; 106: 609-616.
62. Lotze MT, Tracey KJ. High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nat Rev Immunol.* 2005; 5: 331-342.
63. Maroso M, Balosso S, Ravizza T, Liu J, Aronica E, Iyer AM, et al. Toll-like receptor 4 and high-mobility group box-1 are involved in ictogenesis and can be targeted to reduce seizures. *Nat Med.* 2010; 16: 413-419.
64. Parker LC, Whyte MK, Vogel SN, Dower SK, Sabroe I. Toll-like receptor (TLR) 2 and TLR4 agonists regulate CCR expression in human monocytic cells. *J Immunol.* 2004; 172: 4977-4986.
65. Scaffidi P, Misteli T, Bianchi ME. Release of chromatin protein HMGB1 by necrotic cells triggers inflammation. *Nature.* 2002; 418: 191-195.
66. Maroso M, Balosso S, Ravizza T, Liu J, Bianchi ME, Vezzani A. Interleukin-1 type 1 receptor/Toll-like receptor signalling in epilepsy: the importance of IL-1beta and high-mobility group box 1. *J Intern Med.* 2011; 270: 319-326.
67. Walker L, Sills GJ. Inflammation and epilepsy: the foundations for a new therapeutic approach in epilepsy? *Epilepsy Curr.* 2012; 12: 8-12.
68. Rodgers KM, Hutchinson MR, Northcutt A, Maier SF, Watkins LR, Barth DS, et al. The cortical innate immune response increases local neuronal excitability leading to seizures. *Brain.* 2009; 132: 2478-2486.
69. Iori V, Maroso M, Rizzi M, Iyer AM, Vertemara R, Carli M, et al. Receptor for Advanced Glycation Endproducts is upregulated in temporal lobe epilepsy and contributes to experimental seizures. *Neurobiol Dis.* 2013; 58: 102-114.
70. Riazi K, Galic MA, Pittman QJ. Contributions of peripheral inflammation to seizure susceptibility: cytokines and brain excitability. *Epilepsy Res.* 2010; 89: 34-42.
71. Zurolo E, Iyer A, Maroso M, Carbonell C, Anink JJ, Ravizza T, et al. Activation of Toll-like receptor, RAGE and HMGB1 signalling in malformations of cortical development. *Brain.* 2011; 134: 1015-32.
72. Costello DA, Watson MB, Cowley TR, Murphy N, Murphy Royal C, Garlanda C, et al. Interleukin-1alpha and HMGB1 mediate hippocampal dysfunction in SIGIRR-deficient mice. *J Neurosci.* 2011; 31: 3871-9.
73. Mazarati A, Maroso M, Iori V, Vezzani A, Carli M. High-mobility group box-1 impairs memory in mice through both toll-like receptor 4 and Receptor for Advanced Glycation End Products. *Exp Neurol.* 2011; 232: 143-8.
74. Vezzani A, Aronica E, Mazarati A, Pittman QJ. Epilepsy and brain inflammation. *Exp Neurol.* 2013; 244: 11-21.
75. Pulsifer MB, Brandt J, Salorio CF, Vining EP, Carson BS, Freeman JM. The cognitive outcome of hemispherectomy in 71 children. *Epilepsia.* 2004; 45: 243-254.
76. Hoffman CE, Ochi A, Snead OC 3rd, Widjaja E, Hawkins C, et al. Rasmussen's encephalitis: advances in management and patient outcomes. *Childs Nerv Syst.* 2016; 32: 629-640.
77. Granata T, Gobbi G, Spreafico R, Vigeveno F, Capovilla G, Ragona F, et al. Rasmussen's encephalitis: early characteristics allow diagnosis. *Neurology.* 2003; 60: 422-5.
78. Chiapparini L, Granata T, Farina L, Ciceri E, Erbetta A, Ragona F, et al. Diagnostic imaging in 13 cases of Rasmussen's encephalitis: can early MRI suggest the diagnosis? *Neuroradiology.* 2003; 45: 171-83.
79. Chinchilla D1, Dulac O, Robain O, Plouin P, Ponsot G, Pinel JF, et al. Reappraisal of Rasmussen's syndrome with special emphasis on treatment with high doses of steroids. *J Neurol Neurosurg Psychiatry.* 1994; 57: 1325-1333.
80. Lee JS, Juhász C, Kaddurah AK, Chugani HT. Patterns of cerebral glucose metabolism in early and late stages of Rasmussen's syndrome.

- J Child Neurol. 2001; 16: 798-805.
81. Kaiboriboon K, Cortese C, Hogan RE. Magnetic resonance and positron emission tomography changes during the clinical progression of Rasmussen encephalitis. *J Neuroimaging*. 2000; 10:122-125.
82. Fiorella DJ, Provenzale JM, Coleman RE, Crain BJ, Al-Sugair AA. F-fluorodeoxyglucose positron emission tomography and MR imaging findings in Rasmussen encephalitis. *AJNR Am J Neuroradiol* 2001; 22:1291-1299.
83. Longaretti F, Dunkley C, Varadkar S, Vargha-Khadem F, Boyd SG, Cross JH, et al. Evolution of the EEG in children with Rasmussen's syndrome. *Epilepsia*. 2012; 53: 1539-45.
84. Granata T, Gobbi G, Spreafico R, Vigeveno F, Capovilla G, Ragona F, et al. Rasmussen's encephalitis: early characteristics allow diagnosis. *Neurology*. 2003; 60: 422-425.
85. Andrews PI, McNamara JO, Lewis DV. Clinical and electroencephalographic correlates in Rasmussen's encephalitis. *Epilepsia*. 1997; 38: 189-194.
86. Guan Y, Luan G, Zhou J, Liu X. Bilateral Rasmussen encephalitis. *Epilepsy Behav*. 2011; 20: 398-403.
87. Pardo CA, Vining EP, Guo L, Skolasky RL, Carson BS, Freeman JM. The pathology of Rasmussen syndrome: stages of cortical involvement and neuropathological studies in 45 hemispherectomies. *Epilepsia*. 2004; 45: 516-526.
88. Palmer CA, Geyer JD, Keating JM, Gilliam F, Kuzniecky RI, Morawetz RB, et al. Rasmussen's encephalitis with concomitant cortical dysplasia: the role of GluR3. *Epilepsia*. 1999; 40: 242-7.
89. Hart YM, Andermann F, Robitaille Y, Laxer KD, Rasmussen T, Davis R, et al. Double pathology in Rasmussen's syndrome: a window on the etiology? *Neurology*. 1998; 50: 731-5.
90. Takei H, Wilfong A, Malphrus A, Yoshor D, Hunter JV, Armstrong DL, et al. Dual pathology in Rasmussen's encephalitis: a study of seven cases and review of the literature. *Neuropathology*. 2010; 30: 381-391.
91. Thom M, Moran NF, Plant GT, Stevens JM, Scaravilli F. Cortical dysplasia with angiodysgenesis and chronic inflammation in multifocal partial epilepsy. *Neurology*. 1999; 52: 654-657.
92. Iyer A, Zurolo E, Spliet WG, van Rijen PC, Baayen JC, Gorter JA, et al. Evaluation of the innate and adaptive immunity in type I and type II focal cortical dysplasias. *Epilepsia*. 2010; 51: 1763-1773.

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