**⊘**SciMedCentral

#### **Review Article**

# An Embryonic Gene (Cripto-1) in Cancer Stem Cells

Malgorzata Klauzinska, Nadia P. Castro, M. Cristina Rangel, Daniel C. Bertolette and David Salomon\* Department of Tumor Growth Factor section, National Cancer Institute, USA

#### Abstract

Cripto-1 (TDGF-1) is a cell surface glycosylphosphatidylinositol-linked glycoprotein which functions as an obligatory co-receptor for transforming growth factor- $\beta$  (TGF- $\beta$ ) family members. Cripto-1 is essential for early embryonic development and maintenance of embryonic stem cells. Cripto-1 has been shown to be activated in numerous human tumors. Notably, it is expressed in a population of cancer stem cells (CSCs) and facilitates the epithelial to mesenchymal transition (EMT) program. Furthermore, Cripto-1 can significantly enhance tumor cell migration, invasion and angiogenesis. Collectively, these facts suggest that Cripto-1 may be an attractive target in the diagnosis, prognosis and therapy aiming at cancer cell subpopulations with stem-like properties within aggressive tumors.

# **ABBREVIATIONS**

TDGF-1: Teratocarcinoma-Derived Growth Factor 1; EGF: Epidermal Growth Factor; TGF- $\beta$ : The Transforming Growth Factor- $\beta$ ; EMT: Epithelial to Mesenchymal Transition; CSCs: Cancers Stem Cells.

### **INTRODUCTION**

The treatment of cancer is undergoing evolutionary changes as new information about the biology of cancer emerges. Nevertheless, one in four deaths are estimated to result from cancer according to data from 2013 [1]. Conventional therapies such as cytotoxic chemotherapy, radiotherapy and surgical resection can temporarily shrink and restrain the primary tumor but generally the tumor tends to relapse. New targeted therapies, which act on specific molecular targets that are associated with cancer such as Trastuzumab (a monoclonal antibody specifically targeting HER2/neu-over expressing breast tumors), Bevacizumab (a monoclonal antibody therapy against VEGF) or Gefitinib (a tyrosine kinase inhibitor targeting the EGFR, used to treat advanced non-small cell lung cancer), increase the effectiveness and precision of treatment, survival and quality of the patient's life. Unfortunately, even targeted therapies do have limitations. Mainly, cancer cells can become resistant to them. Resistance to chemotherapy and molecularly-targeted therapies is one of the major causes which under lies cancer treatment failure [2]. Ginsburg and Willard [3] have reported that chemoresistance and treatment effects depend on the distinct patterns of genes associated with stemness/differentiation pathways. Indeed, tumors are a heterogeneous mixture consisting of epithelial non-stem cells and cancer stem cells (CSCs) with mixed epithelial to mesenchymal phenotypes [4]. Reports have shown a link between stem-like properties and therapy resistance in

# Journal of Cancer Biology & Research

#### \*Corresponding author

David Salomon, Tumor Growth Factor Section, Mouse Cancer Genetics Program, Center for Cancer Research, National Cancer Institute, Building 560, Room 12-67, Frederick, MD 21702-1201, USA; Tel: 301-228-4770, Email: salomond@mail.nih.gov

Submitted: 01 November 2014

Accepted: 27 January 2015

Published: 29 January 2015

Copyright

© 2015 Salomon et al.

OPEN ACCESS

glioblastoma, colon cancer, breast cancer, acute myelogenous leukemia and numerous other tumors demonstrating that CSCs are more resistant to therapy compared to non-CSCs [5-8]. This minority tumor subpopulation of CSCs, due to features like over expression of ABC transporters, a slow rate of self-renewal, and an active DNA repair capacity, is probably responsible for chemoresistance in cancer and the reason why treatment fails [9]. Interestingly, many current drug treatments result in an enrichment of CSCs in the tumor [10] e.g. oxaliplatin treatment of colon cancers actually boosted the abundance of CSCs by more than 10 times [11]. It is clear that a more complete understanding of the properties and characteristics of CSCs is a key to future success in cancer treatment. To improve the outcome of cancer treatments, CSCs must be effectively targeted and eradicated.

#### **CANCER STEM CELLS**

In adult organisms, normal stem cells can be found in different tissue types and function as an internal repair system, dividing to replenish specialized cells and also maintaining normal turnover of regenerative organs, such as blood, skin, or intestinal tissues [12]. Similar to normal stem cells, CSCs have been suggested to maintain the tumors and the abnormal regulation of CSCs is implicated in the generation and progression of malignant tumors. CSCs are defined by their ability to efficiently regenerate the original phenotype of the tumor upon inoculation into immune deficient mice [13]. This functional definition is often complemented by including the expression of cell-surface markers that are also expressed by the normal stem cells in the tissue of origin [14,15]. CSCs, also called tumor-initiating cells, were initially identified in the hematopoietic system [16,17]. Later, they were also found in solid tumors, including those arising in the breast, lung, prostate, colon, brain, head and neck, and pancreas [14,16,18,19]. Likewise, epithelial-mesenchymal

Cite this article: Klauzinska M, Castro NP, Rangel, Bertolette DC, Salomon D (2015) An Embryonic Gene (Cripto-1) in Cancer Stem Cells. J Cancer Biol Res 3(1): 1056.

### **⊘**SciMedCentral-

transition (EMT), which is critical during normal development, fibrosis and wound healing, has also been implicated as a means by which transformed epithelial cells can acquire the abilities to invade, resist apoptosis, and disseminate, thus contributing to tumor invasion and metastasis[20-23]. Recently, the link between activation of the EMT program and the genesis and maintenance of cells with stem cell-like properties has been confirmed [24-26]. This connection may generate more aggressive cell behaviors, hence demanding the development of therapeutic strategies designed to interfere with EMT and CSC activity within tumors.

### **CRIPTO-1 AND EMT**

The Teratocarcinoma-Derived Growth Factor-1, Tdgf-1 or *Cripto-1* gene, a member of the TGF- $\beta$  super family, plays a fundamental role in normal development as well as during the regulation of self-renewal and pluripotency of mouse and human embryonic stem cells. Furthermore, Cripto-1 represents a clear example of an embryonic signaling molecule which when reactivated in an uncontrolled manner can drive cell transformation and tumor progression in adult tissues [27]. Interestingly Cripto-1 effects multiple signaling pathways known to be EMT triggers such as transforming growth factor (TGF)-β, fibroblast growth factors (FGFs), Wnts and Notch [28]. Multiple studies showed that Cripto-1 over expression in mammary epithelial cells and multiple cancer cell lines leads to their enhanced migration and invasion capacity. HC-11 mouse mammary epithelial cells over expressing Cripto-1 undergo EMT, as shown by a decrease in E-cadherin expression and an increase in vimentin, N-cadherin, and Snail expression [29]. Also over expression of Cripto-1 in MCF-7 breast cancer cells, Caski human cervical carcinoma cells and LS174-T colon cancer cells show a significant increase in their migration and invasion behaviors compared with parental cell lines. Our previous study showed that mammary tumors from MMTV-Cripto-1 transgenic mice exhibit areas of morphological changes associated with EMT such as reduction of intercellular adhesion proteins, e.g. E-cadherin, and an increase in the expression of mesenchymal markers, including N-cadherin and vimentin. Moreover, we detected increased expression of several integrins, including integrins  $\beta$ 3,  $\beta$ 5,  $\beta$ 1 and β4 [29], also linked to EMT and cell spreading. Markers of EMT could also be detected in uterine leiomyosarcomas that develop in approximately 20% of nulliparous or multiparous MMTV-Cripto-1 mice [30].Since Cripto-1 has been found to promote EMT in vitro and in vivo in mouse mammary epithelial cells, cancer cell lines and mouse mammary tumors [28], it is possible that this gene is involved in driving an EMT program in breast cancer stem-like cells thereby supporting their self-renewal and metastatic abilities [31].

Cripto-1 expression is not restricted to malignant breast tissues.Cripto-1 is over expressed in a variety of human tumors and its expression has been associated with more aggressive behavior in several types of cancers [28]. Increased Cripto-1 expression and decreased E-cadherin expression have been positively associated with tumor progression, poor prognostic factors (e.g. tumor size, depth of invasion, lymph node metastasis, liver metastasis, and TNM stage) in patients with gastric, breast and colon cancers [32,33].Several reports have showed that Cripto-1 might also play a role in human melanomas [34, 35]. Cripto-1 expression was detected in 43% of primary human cutaneous melanomas and in 57% of melanoma cell lines. In melanomas, Cripto-1 strongly enhanced the motility of melanoma cell lines and blocking of Cripto-1 expression using small-interfering RNAs significantly inhibited growth and the invasive ability of melanoma cells [34].

# **CRIPTO-1 AND CANCER STEM CELLS**

Cripto-1 is known to be involved in stem cell maintenance and pluripotency [31]. Furthermore, Cripto-1 contributes to the tumorigenicity of invasive cancer cells with stem-like characteristics [36] and multiple groups have detected stem cell markers in Cripto-1-positive human cancer cells. Cripto-1 has been demonstrated to be a potential marker for the identification of CSCs in human malignant melanomas [37]. Strizzi et al, using Fluorescence-activated Cell Sorting (FACS), isolated from a human melanoma cell line a subpopulation of cells that expresses Cripto-1 on the cell surface and possesses stem-like characteristics [37]. Although the Cripto-1-positive subpopulation represented a relatively small fraction of the population, it showed a more spindle-shaped morphology, and exhibited increased expression of embryonic stem cell-associated transcription factors, such as Oct-4 and Nanog, as compared to the parental melanoma cells [37]. In another study, Cripto-1 was identified together with Oct-4 and SUZ-12 in a small subpopulation of stem-like cells in both hormone-responsive and non-responsive human prostate tumor cells [38]. Furthermore, Watanabe and colleagues reported that Cripto-1 is heterogeneously expressed in human embryonal carcinoma (EC) cells, which are pluripotent stem cells derived from germ cell teratocarcinomas (the malignant counterpart of embryonic human stem cells). In accordance with Cripto-1 expression on the cell surface, two populations of EC cells were isolated that expressed high and low levels of Cripto-1. The Cripto-1 high subpopulation formed significantly more tumor spheres in vitro under serum-free conditions than the Cripto-1 low-expressing cells. Additionally, by injecting Cripto-1 high expressing cells subcutaneously into nude mice, the authors observed that these cells were able to generate significantly larger tumors with shorter latency period when compared with tumors derived from Cripto-1 low expressing cells[39]. In addition, Cripto-1/GRP78 (heat shock 78kDa glucose-regulated protein) signaling has been suggested as an important pathway that regulates hematopoietic stem cell quiescence [40]. Under hypoxic conditions, Cripto-1 expression was essential in regulating the lineage specification of a CD34/GRP78 myeloid progenitor population in the hematopoietic stroma niche of the bone marrow [40]. Finally, the Nodal/Cripto-1 signaling pathway has been shown to maintain self-renewal and *in vivo* oncogenic capacity of pancreatic CSCs [41]. Inhibition of this signaling by blocking the Alk4/7 receptor reversed the chemoresistance of orthotopically engrafted pancreatic CSCs, suggesting a novel therapeutic approach to target pancreatic CSCs. All these novel findings suggest that Cripto-1 may represent an important marker for the identification of a cancer cell subpopulation with stem-like properties within aggressive tumors that are resistant to conventional therapy.

# **MULTIPLE MODALITIES TO TARGET CRIPTO-1**

As Cripto-1 may be expressed in a population of CSCs and contribute to CSC self-renewal and EMT, it is potentially an attractive therapeutic target. Beyond the potential role of Cripto-1 in the induction or maintenance of CSCs, the role of Cripto-1 in

### **⊘**SciMedCentral-

the tumor microenvironment makes it a potential therapeutic target since it also assists in sustaining the niche in which the tumor thrives. Our laboratory and others have successfully targeted Cripto-1 in vitro in various types of cancer cell lines and in vivo in several types of xenograft tumor model systems thereby abrogating its effects on tumor growth and metastasis [36]. There are various methods that have been shown to counteract the oncogenic effects of Cripto-1 by targeting either Cripto-1 mRNA or protein. Interference of Cripto-1 expression has been effective by using antisense oligonucleotides against Cripto-1 in vitro in breast, colon and ovarian cancer cells, and in vivo in colon cancer xenografts [42-45]. Recently, it has been shown that micro RNAs (miRNA), small non-coding RNA molecules that inhibit mRNA translation, are involved in the regulation of Cripto-1 expression. In particular, miR-15a -16 was shown to downregulate Cripto-1 expression and disrupt Cripto-1-mediated invasion of non-small cell lung cancer cells (NSCLC) in vitro [46]. Also, miR-15a -16 was able to reduce the tumor volume of NSCLC xenograft tumors [46]. Using proper delivery systems, this strategy could prove useful in targeting Cripto-1 expressing cells refractory to normal therapy.Cripto-1-neutralizing monoclonal antibodies and small molecule compounds that target the various domains of Cripto-1 have shown to be efficacious by inhibiting the proliferation of multiple cancer cell lines, blocking the growth of tumor xenografts, enhancing the effects of chemotherapeutic regimens and inducing apoptosis [47-51]. Antibodies directed against the EGF-like domain of Cripto-1 have prevented the activation of Smad signaling by interfering with Nodal-mediated Alk4 activation [48]. Antibody neutralization of the EGF-domain and its interactions can also impair the activation of Akt leading to apoptosis as seen in a multi-drug resistant leukemia model [49]. An antibody directed against GRP78 is also able to prevent Cripto-1-mediated activation of MAPK and Akt [52]. Antagonists of Cripto-1's CFC domain, such as monoclonal antibodies[47], Alantolactone (a natural small molecule compound) [53], and a synthetic Cripto-1-binding peptide [54] have been shown to disrupt Cripto-1/Nodal binding and suppress Cripto-1-mediated inhibition of TGF- $\beta$ /Active in growth inhibition. Also, a kinasedeficient soluble version of Alk4 has recently been shown to be efficacious in blocking Cripto-1 activation of Akt [55]. Cripto-1 as a possible marker for CSCs provides the potential of specifically delivering nanomedicines to Cripto-1-expressing cells using nonneutralizing Cripto-1 antibodies, similar to a recently concluded phase I clinical trial using a cytoxin-conjugated antibody directed against the amino-terminal region of Cripto-1[50], thereby eliminating Cripto-1-positive CSCs.

Taken together, these data show that targeting Cripto-1 in a therapeutic setting can be quite advantageous due to its role in multiple processes of tumor formation and metastasis. The combination of therapies directed against Cripto-1 along with treatments against the bulk tumor could provide not only success against a primary tumor, but prevent metastasis and relapse.

#### REFERENCES

- Siegel R, Naishadham D, Jemal A. Cancer statistics, 2013. CA Cancer J Clin. 2013; 63: 11-30.
- 2. Holohan C, Van Schaeybroeck S, Longley DB, Johnston PG. Cancer drug resistance: an evolving paradigm. Nat Rev Cancer. 2013; 13: 714-726.

- 3. Ginsburg GS, Willard HF. Genomic and personalized medicine: foundations and applications. Transl Res. 2009; 154: 277-287.
- 4. Pattabiraman DR, Weinberg RA. Tackling the cancer stem cells what challenges do they pose? Nat Rev Drug Discov. 2014; 13: 497-512.
- Bao S, Wu Q, McLendon RE, Hao Y, Shi Q, Hjelmeland AB, et al. Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. Nature. 2006; 444: 756-760.
- Diehn M, Cho RW, Lobo NA, Kalisky T, Dorie MJ, Kulp AN, et al. Association of reactive oxygen species levels and radioresistance in cancer stem cells. Nature. 2009; 458: 780-783.
- Ishikawa F, Yoshida S, Saito Y, Hijikata A, Kitamura H, Tanaka S, et al. Chemotherapy-resistant human AML stem cells home to and engraft within the bone-marrow endosteal region. Nat Biotechnol. 2007; 25: 1315-1321.
- Zhang M, Atkinson RL, Rosen JM. Selective targeting of radiationresistant tumor-initiating cells. Proc Natl Acad Sci U S A. 2010; 107: 3522-3527.
- O'Connor ML, Xiang D, Shigdar S, Macdonald J, Li Y, Wang T, et al. Cancer stem cells: A contentious hypothesis now moving forward. Cancer Lett. 2014; 344: 180-187.
- 10.De Sousa E Melo F, Vermeulen L, Fessler E, Medema JP. Cancer heterogeneity--a multifaceted view. EMBO Rep. 2013; 14: 686-695.
- 11.Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. Nat Rev Cancer. 2005; 5: 275-284.
- 12.Bendall SC, Stewart MH, Bhatia M. Human embryonic stem cells: lessons from stem cell niches in vivo. Regen Med. 2008; 3: 365-376.
- 13.Visvader JE, Lindeman GJ. Cancer stem cells in solid tumours: accumulating evidence and unresolved questions. Nat Rev Cancer. 2008; 8: 755-768.
- 14. Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. Proc Natl Acad Sci USA. 2003; 100: 3983-3988.
- 15.Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011; 144: 646-674.
- 16.Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. Nat Med. 1997; 3: 730-737.
- 17. Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. Nature. 2001; 414: 105-111.
- 18. Lapidot T, Sirard C, Vormoor J, Murdoch B, Hoang T, Caceres-Cortes J, et al. A cell initiating human acute myeloid leukaemia after transplantation into SCID mice. Nature. 1994; 367: 645-648.
- 19. Singh SK, Hawkins C, Clarke ID, Squire JA, Bayani J, Hide T, et al. Identification of human brain tumour initiating cells. Nature. 2004; 432: 396-401.
- 20.Boyer B, Thiery JP. Epithelium-mesenchyme interconversion as example of epithelial plasticity. APMIS. 1993; 101: 257-268.
- 21.Zeisberg M, Neilson EG. Biomarkers for epithelial-mesenchymal transitions. J Clin Invest. 2009; 119: 1429-1437.
- 22.Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. J Clin Invest. 2009; 119: 1420-1428.
- 23. Micalizzi DS, Farabaugh SM, Ford HL. Epithelial-mesenchymal transition in cancer: parallels between normal development and tumor progression. J Mammary Gland Biol Neoplasia. 2010; 15: 117-134.
- 24. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, et al. The

# **⊘**SciMedCentral-

epithelial-mesenchymal transition generates cells with properties of stem cells. Cell. 2008; 133: 704-715.

- 25. Scheel C, Eaton EN, Li SH, Chaffer CL, Reinhardt F, Kah KJ, et al. Paracrine and autocrine signals induce and maintain mesenchymal and stem cell states in the breast. Cell. 2011; 145: 926-940.
- 26. Scheel C, Weinberg RA. Cancer stem cells and epithelial-mesenchymal transition: concepts and molecular links. Semin Cancer Biol. 2012; 22: 396-403.
- 27. de Castro NP, Rangel MC, Nagaoka T, Salomon DS, Bianco C. Cripto-1: an embryonic gene that promotes tumorigenesis. Future Oncol. 2010; 6: 1127-1142.
- 28.Rangel MC, Karasawa H, Castro NP, Nagaoka T, Salomon DS, Bianco C. Role of Cripto-1 during epithelial-to-mesenchymal transition in development and cancer. Am J Pathol. 2012; 180: 2188-2200.
- 29.Strizzi L, Bianco C, Normanno N, Seno M, Wechselberger C, Wallace-Jones B, et al. Epithelial mesenchymal transition is a characteristic of hyperplasias and tumors in mammary gland from MMTV-Cripto-1 transgenic mice. J Cell Physiol. 2004; 201: 266-276.
- 30.Strizzi L, Bianco C, Hirota M, Watanabe K, Mancino M, Hamada S, et al. Development of leiomyosarcoma of the uterus in MMTV-CR-1 transgenic mice. J Pathol. 2007; 211: 36-44.
- 31.Bianco C, Rangel MC, Castro NP, Nagaoka T, Rollman K, Gonzales M, et al. Role of Cripto-1 in stem cell maintenance and malignant progression. Am J Pathol. 2010; 177: 532-540.
- 32. Zhong XY, Zhang LH, Jia SQ, Shi T, Niu ZJ, Du H, et al. Positive association of up-regulated Cripto-1 and down-regulated E-cadherin with tumour progression and poor prognosis in gastric cancer. Histopathology. 2008; 52: 560-568.
- 33.Gong YP, Yarrow PM, Carmalt HL, Kwun SY, Kennedy CW, Lin BP, et al. Overexpression of Cripto and its prognostic significance in breast cancer: a study with long-term survival. Eur J Surg Oncol. 2007; 33: 438-443.
- 34. De Luca A, Lamura L, Strizzi L, Roma C, D'Antonio A, Margaryan N, et al. Expression and functional role of CRIPTO-1 in cutaneous melanoma. Br J Cancer. 2011; 105: 1030-1038.
- 35.Strizzi L, Margaryan NV, Gilgur A, Hardy KM, Normanno N, Salomon DS, et al. The significance of a Cripto-1 positive subpopulation of human melanoma cells exhibiting stem cell-like characteristics. Cell Cycle. 2013; 12: 1450-1456.
- 36. Klauzinska M, Castro NP, Rangel MC, Spike BT, Gray PC, Bertolette D, et al. The multifaceted role of the embryonic gene Cripto-1 in cancer, stem cells and epithelial-mesenchymal transition. Semin Cancer Biol. 2014; 29: 51-58.
- 37. Strizzi L, Abbott DE, Salomon DS, Hendrix MJ. Potential for cripto-1 in defining stem cell-like characteristics in human malignant melanoma. Cell Cycle. 2008; 7: 1931-1935.
- 38.Cocciadiferro L, Miceli V, Kang KS, Polito LM, Trosko JE, Carruba G. Profiling cancer stem cells in androgen-responsive and refractory human prostate tumor cell lines. Ann N Y Acad Sci. 2009; 1155: 257-262.
- 39. Watanabe K, Meyer MJ, Strizzi L, Lee JM, Gonzales M, Bianco C, et al. Cripto-1 is a cell surface marker for a tumorigenic, undifferentiated subpopulation in human embryonal carcinoma cells. Stem cells. 2010; 28:1303-1314.
- 40. Miharada K, Karlsson G, Rehn M, Rörby E, Siva K, Cammenga J, et al.

Cripto regulates hematopoietic stem cells as a hypoxic-niche-related factor through cell surface receptor GRP78. Cell Stem Cell. 2011; 9: 330-344.

- 41.Lonardo E, Hermann PC, Mueller MT, Huber S, Balic A, Miranda-Lorenzo I, et al. Nodal/Activin signaling drives self-renewal and tumorigenicity of pancreatic cancer stem cells and provides a target for combined drug therapy. Cell Stem Cell. 2011; 9: 433-446.
- 42. Casamassimi A, De Luca A, Agrawal S, Stromberg K, Salomon DS, Normanno N. EGF-related antisense oligonucleotides inhibit the proliferation of human ovarian carcinoma cells. Ann Oncol. 2000; 11: 319-325.
- 43. De Luca A, Arra C, D'Antonio A, Casamassimi A, Losito S, Ferraro P, et al. Simultaneous blockage of different EGF-like growth factors results in efficient growth inhibition of human colon carcinoma xenografts. Oncogene. 2000; 19: 5863-5871.
- 44. De Luca A, Casamassimi A, Selvam MP, Losito S, Ciardiello F, Agrawal S, et al. EGF-related peptides are involved in the proliferation and survival of MDA-MB-468 human breast carcinoma cells. Int J Cancer. 1999; 80: 589-594.
- 45. Normanno N, De Luca A, Maiello MR, Bianco C, Mancino M, Strizzi L, et al. CRIPTO-1: a novel target for therapeutic intervention in human carcinoma. Int J Oncol. 2004; 25: 1013-1020.
- 46. Chen F, Hou SK, Fan HJ, Liu YF. MiR-15a-16 represses Cripto and inhibits NSCLC cell progression. Mol Cell Biochem. 2014; 391: 11-19.
- 47. Adkins HB, Bianco C, Schiffer SG, Rayhorn P, Zafari M, Cheung AE, et al. Antibody blockade of the Cripto CFC domain suppresses tumor cell growth in vivo. J Clin Invest. 2003; 112: 575-587.
- 48.Bianco C, Salomon DS. Targeting the embryonic gene Cripto-1 in cancer and beyond. Expert Opin Ther Pat. 2010; 20: 1739-1749.
- 49.Hu XF, Li J, Yang E, Vandervalk S, Xing PX. Anti-Cripto Mab inhibit tumour growth and overcome MDR in a human leukaemia MDR cell line by inhibition of Akt and activation of JNK/SAPK and bad death pathways. Br J Cancer. 2007; 96: 918-927.
- 50. Sanicola-Nadel M, Williams KP, Schiffer SG, Rayhorn P: Cripto blocking antibodies and uses thereof. In. Edited by USPTO, vol. 7,888,052. US: Biogen-Idec; 2011.
- 51.Xing PX, Hu XF, Pietersz GA, Hosick HL, McKenzie IF. Cripto: a novel target for antibody-based cancer immunotherapy. Cancer Res. 2004; 64: 4018-4023.
- 52.Kelber JA, Panopoulos AD, Shani G, Booker EC, Belmonte JC, Vale WW, et al. Blockade of Cripto binding to cell surface GRP78 inhibits oncogenic Cripto signaling via MAPK/PI3K and Smad2/3 pathways. Oncogene. 2009; 28: 2324-2336.
- 53.Shi Y, Bao YL, Wu Y, Yu CL, Huang YX, Sun Y, et al. Alantolactone inhibits cell proliferation by interrupting the interaction between Cripto-1 and activin receptor type II A in activin signaling pathway. J Biomol Screen. 2011; 16: 525-535.
- 54. Lonardo E, Parish CL, Ponticelli S, Marasco D, Ribeiro D, Ruvo M, et al. A small synthetic cripto blocking Peptide improves neural induction, dopaminergic differentiation, and functional integration of mouse embryonic stem cells in a rat model of Parkinson's disease. Stem Cells. 2010; 28:1326-1337.
- 55.Spike BT, Kelber JA, Booker E, Kalathur M, Rodewald R, Lipianskaya J, et al. CRIPTO/GRP78 signaling maintains fetal and adult mammary stem cells ex vivo. Stem Cell Reports. 2014; 2: 427-439.

#### Cite this article

Klauzinska M, Castro NP, Rangel, Bertolette DC, Salomon D (2015) An Embryonic Gene (Cripto-1) in Cancer Stem Cells. J Cancer Biol Res 3(1): 1056.

J Cancer Biol Res 3(1): 1056 (2015)