

## Research Article

# Temporal Effects of Elevated Carbon Dioxide and Ozone on Soil Carbon and Nitrogen Stoichiometry in a No-till Soybean-Wheat Agro-ecosystem

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## Abstract

The effects of elevated CO<sub>2</sub> and O<sub>3</sub> on plants have been studied widely, but their possible effects on carbon: nitrogen stoichiometry of soil organic matter (SOM) has received much less attention. We conducted a five-year field experiment to address such effects on SOM. We used open-top chambers to expose a no-till (NT) soybean-wheat system to a factorial combinations of two CO<sub>2</sub> (360 & 500 μmol mol<sup>-1</sup>) and two O<sub>3</sub> (25 & 70 nmol mol<sup>-1</sup>) treatments daily from April to October under ambient light and temperature conditions. Geo-referenced composite soil samples were collected annually and analyzed for soil total organic C (TOC), N (TN), hydrogen (H) concentrations, and bulk density. TOC increased through time under both CO<sub>2</sub> levels, but the increase was significantly greater under elevated CO<sub>2</sub> (585-kg ha<sup>-1</sup> y<sup>-1</sup>) compared with the ambient CO<sub>2</sub> (18-kg ha<sup>-1</sup> y<sup>-1</sup>). TN decreased over time under both levels of the CO<sub>2</sub> and O<sub>3</sub> treatments. The rate of decrease in TN was greater under elevated CO<sub>2</sub> (216-kg N ha<sup>-1</sup> y<sup>-1</sup>) compared to ambient CO<sub>2</sub> (152-kg N ha<sup>-1</sup> y<sup>-1</sup>), but it was smaller under elevated O<sub>3</sub> (172-kg N ha<sup>-1</sup> y<sup>-1</sup>) as compared to low O<sub>3</sub> (197-kg N ha<sup>-1</sup> y<sup>-1</sup>). The C:N was increased significantly over time under both levels of CO<sub>2</sub> or O<sub>3</sub> treatments. Results suggest that increased TOC sequestration at elevated CO<sub>2</sub> could elicit a progressive soil N deficiency. In contrast, the impact of elevated O<sub>3</sub> (+O<sub>3</sub> and O<sub>3</sub>+CO<sub>2</sub>) may cause a small accumulation of recalcitrant C, which in the long-term could affect SOM lability. Increased soil N under elevated O<sub>3</sub> may enhance N leaching from soil.

## INTRODUCTION

Tropospheric CO<sub>2</sub> and O<sub>3</sub> concentrations have been increasing and will continue to increase in response to human activities [1-4]. At least in the short term, elevated CO<sub>2</sub> can increase plant photosynthesis, water- and nutrient-use efficiencies, net primary production, root production, rhizosphere deposition, and mycorrhizal associations [5], but alter plant biomass composition by increasing C:N stoichiometry with higher proportions of nonstructural carbohydrates, lignin and phenolic compounds [6]. However, it has been suggested that accumulation of lignin and phenolic compounds in plant biomass may be due to nutrient limitations rather than the direct effect of elevated CO<sub>2</sub> [7].

Near-surface O<sub>3</sub> concentration is the product of photochemical reactions of carbon monoxide, methane and other hydrocarbons

in the presence of nitrogen oxides [1]. O<sub>3</sub> is a phytotoxic pollutant that affects the integrity and permeability of plant-cell membranes and decreases metabolic efficiency [1,8,9]. Elevated O<sub>3</sub> may decrease crop yield and reduce below-ground carbon allocation [10]. However, plants may acclimate to, or resist, the effects of elevated O<sub>3</sub> through formation of antioxidants [11] and/or changing structural and cellular composition of plant biomass [12]. The latter can result in plant litters that are relatively recalcitrant for soil microorganisms [13,14].

Plant biomass is the main source of SOM formation [15,16]. Effects of elevated CO<sub>2</sub> and/or O<sub>3</sub> on plant biomass may have immediate and variable effects on soil food webs, microbial decomposition of organic matter and biological efficiency, and consequently on the soil's ability as a sink or source of C and other

nutrients by influencing SOM lability in response to agricultural management practices [17-19].

No-till (NT) is an agricultural practice that allows surface accumulation of crop residues. Reduced contact between soil microorganisms and crop residues under NT favors dominance of fungal food webs to utilize N poor un-fragmented crop residues [20-22]. Extensive fungal hyphae-plant root associations may aid in the protection of C especially particulate organic matter (POM) in soil macro-aggregates [22]. Furthermore, microbial preference for easily decomposable C-H-O enriched root exudates under elevated CO<sub>2</sub> may slow-down and/or reduce decomposition of high C:N crop residues and subsequent accumulation of C and other nutrients in SOM [22-24]. However, biological processes associated with crop residue decomposition are often influenced by N availability [25]. Decomposition of high C:N crop residue may increase microbial demand for N and, in turn, increase mineralization of low C:N native SOM [26]. Therefore, we hypothesized that no-till surface accumulation of high C:N crop residue produced under elevated CO<sub>2</sub> would be converted to C-H enriched but N-deficient SOM.

Soil functional processes that are mediated by microbes are often C-limited. Therefore, reduced crop biomass with greater recalcitrant composition at elevated O<sub>3</sub> could lead to an intense competition among the heterotrophic microbes for labile C metabolism [27,28]. An intense competition for labile C may favor the dominance of energy inefficient and generalist microbial feeders (e.g. bacteria) under labile C-limited ecosystems [16,21]. As a result, microbes may be forced to mineralize native SOM or autolyse their cells to fulfill increasing demand for labile C [27]. Therefore, we hypothesized that soils with growing crops under elevated O<sub>3</sub> will lose labile C faster but accumulate a small amount of recalcitrant C over time.

Soil organic matter is a major reservoir of nutrients, especially C and N that are stoichiometrically linked. This link is often influenced by H contents to maintain quality, stability and biogeochemistry of SOM [28,29]. A greater availability of high C:N crop residue or a lack of easily decomposable C-H enriched substrates for metabolism under elevated CO<sub>2</sub> or O<sub>3</sub> may be a key factor for C and N sequestration or depletion [30]. Knowledge of how SOM formation and lability will respond to long-term effects of elevated CO<sub>2</sub> or O<sub>3</sub> on plants is important to understand C and N dynamics.

Several studies have evaluated long-term effects of elevated CO<sub>2</sub> and O<sub>3</sub> on below ground responses and soil C dynamics for different plants [4,6,17-19,30]. Soil ability to serve as a sink for C under elevated CO<sub>2</sub> may be limited [6]. Significant variable CO<sub>2</sub> × O<sub>3</sub> × time interactions on below ground processes, residue decomposition, and soil C dynamics have been reported [6,19,29-32]. However, there is no long-term research conducted to date examining CO<sub>2</sub> × O<sub>3</sub> × time interaction on soil C and N dynamics under continuous NT soybean-wheat agro ecosystems. Therefore, the objective of our study was to address temporal effects of elevated CO<sub>2</sub> and O<sub>3</sub>, individually and in combination, on TOC, TN and H contents of SOM under NT soybean (*Glycine max*, L.) - wheat (*Triticumaestivum*, L.) rotation. More specifically, we used analysis of covariance and repeated-measures analysis of variance to address: (1) the temporal change in the responses

(i.e., test of the equality of the slope of the temporal trends to zero); (2) the interactive effects of CO<sub>2</sub> and O<sub>3</sub> on the responses studied; and (3) the interactive effects of temporal change and air quality treatments (i.e., test of interaction between time and the main or simple effects of CO<sub>2</sub> and O<sub>3</sub>).

## MATERIALS AND METHODS

The study was conducted at the USDA-ARS Climate Stress Laboratory field research facility at Beltsville, MD, USA. The soil is a moderately well drained Codorus silt loam (fine sand, acid, mesic, Fluvaquentic Dystrochrept), which is formed from recently deposited alluvium washed from soils developed from acid crystalline rocks of the Piedmont [33]. Initial studies on the soil properties and climatic conditions have been described elsewhere [34].

### Experimental treatments and cultural practices

Air quality treatments consisted of a 2<sup>2</sup> factorial combinations of two CO<sub>2</sub> (ambient at 365±5 vs. elevated at 500±5 μmol mol<sup>-1</sup>) and two O<sub>3</sub> (charcoal filtered air at 25±5 vs. elevated at 70±10 nmol mol<sup>-1</sup>) levels. Each treatment combination was replicated twice, using eight open-top chambers (OTC). The OTC's (3-m diameter) were placed on 4-m × 4-m plots in which wheat and soybeans were grown in rotation from seed to maturity under continuous no-till. The CO<sub>2</sub> treatment was supplied daily from a bulk tank for 18-hr (0300 – 2100-hr EST). The O<sub>3</sub> was produced from cylinder O<sub>2</sub> using a Griffin O<sub>3</sub> generator (Lodi, NJ) and supplied daily for 7 h (0900 – 1600-hr EST). All OTC's were equipped with sprinkler irrigation system to maintain near field moisture capacity (16 to 18%, dry weight) of soil [35,36].

Wheat was planted in the fall with split applications of standard N-P-K fertilization in both fall and spring. Wheat was harvested by the 3<sup>rd</sup> week of June. Short-day soybean seedlings were transplanted to the plots in rows 60-cm apart between plants and 10-cm apart within the rows immediately following wheat harvest. Weeding was performed manually by hand. To control insects, Diazinon 4E (Ciba, Greensboro, NC) @ 1-mL L<sup>-1</sup> solution was applied. The plots were also sprayed for powdery mildew control on wheat one or more times each spring using Benomyl (Dupont Agricultural Products, Wilmington, DE) at the rate of 0.3-kg ha<sup>-1</sup>. Following wheat harvest, grasses and weeds in the plots within OTC's were killed with Glyphosphate (Monsanto Chemical, St. Louis, MO) using a 1.5% solution as foliar spray. The treatments were terminated in late October prior to harvesting soybean.

### Soil collection, processing and analysis

Soil sampling and analysis started two years after the establishment of plots and 18 months after the initiation of gaseous treatments and continued for four years. Fourteen soil cores (1.9-cm dia.) from 15-cm depth were randomly collected between the rows of plants after fall harvest in late October or early November. Soil samples were then gently sieved through a 2-mm sieve, oven-dried at 105±2°C for 24-h to determine antecedent moisture content for correction of weight when calculating bulk density. A sub-sample of the oven-dried soil was ground with a ceramic mortar and pestle to pass through a 200-μm sieve before analysis.

A 150 to 200-mg sample of air-dried ground soil was used to analyze for TOC, TN and H contents using the LECO® dry combustion method. The thermal oxidation of SOM by LECO dry combustion not only produces CO<sub>2</sub> and N<sub>2</sub> but also H<sub>2</sub>O, as a measure of H [28], according to the equation: C<sub>6</sub>H<sub>12</sub>O<sub>6</sub> + 6O<sub>2</sub> → 6CO<sub>2</sub> + 6H<sub>2</sub>O. Therefore, all H is derived from thermal oxidation of SOM [32]. The stocks of C, N, and H were determined by multiplying with the concurrently measured soil bulk density (pb). The pb was calculated using the oven-dried weight of a known volume of soil as: pb (g cm<sup>-3</sup>) = (πr<sup>2</sup> \* l \* n) w<sup>-1</sup>. Where r is the internal radius of the soil core sampler, l is the length of the soil core, n is the number of soil cores, and w is the total weight (g) of oven-dried soil.

### Statistical analysis

Analysis of covariance (ANCOVA) was performed in which the response variables were TOC, TN, H, C:N, and H:N, and explanatory variables were CO<sub>2</sub> and O<sub>3</sub> as class variables, and time (year after the initiation of the CO<sub>2</sub> and O<sub>3</sub> treatments) as a continuous covariate. ANCOVA allowed to detect whether or not: (1) there was a significant temporal change in the response variable being studied (test of the equality of the slopes of the temporal changes to zero), (2) the rate of the temporal changes depended on the treatments (test of interaction between time and class factors—CO<sub>2</sub> or O<sub>3</sub>), and (3) there was an interaction between the effects of CO<sub>2</sub> and O<sub>3</sub>. The MIXED procedure of the SAS System was used for statistical analysis. This routine has appropriate options (RANDOM and/or REPEATED) to allow for appropriate calculation of errors, F and P values for the factors involved (CO<sub>2</sub>, O<sub>3</sub>, time), and their interactions. Temporal trends were compared for the main-effects of CO<sub>2</sub> and O<sub>3</sub> when there was no significant interaction between CO<sub>2</sub> and O<sub>3</sub>. In case of a significant CO<sub>2</sub>-by-O<sub>3</sub> interaction (CO<sub>3</sub> × O<sub>3</sub>), the temporal trends were compared for the four simple effects (four combinations of two CO<sub>2</sub>-by-two O<sub>3</sub> levels).

## RESULTS AND DISCUSSION

The main effects of CO<sub>2</sub> or O<sub>3</sub> on TOC and TN contents, and the C:N and H:N are reported and discussed because the related CO<sub>2</sub>-by-O<sub>3</sub> interactions were insignificant. The CO<sub>2</sub> × O<sub>3</sub> effect was significant for H content (P<0.01) and for the C:H (P<0.01), and thus, the simple effects are reported and discussed for these variables.

Intercepts of the temporal trend for the main- or simple-effects of CO<sub>2</sub> or O<sub>3</sub> for TOC, TN and H contents and their ratios did not differ significantly (Figures 1–9). Since the experiment was established on a single type of soil, we assumed that there was no significant difference in soil properties before the initiation of the CO<sub>2</sub> and O<sub>3</sub> treatments, and that any temporal difference in TOC, TN and H contents should be in response to applied treatments over time. A lack of significant differences among the intercepts of the temporal trends supports these assumptions.

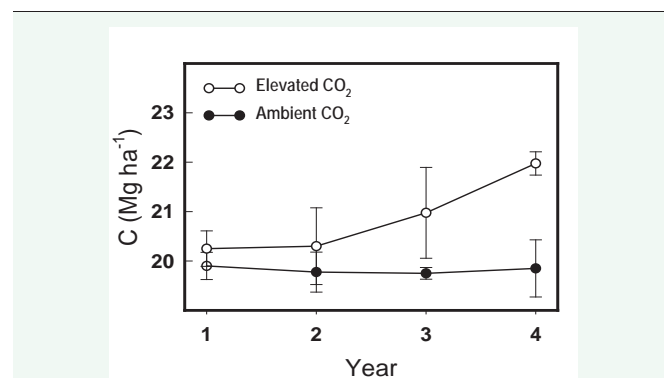
A significant CO<sub>2</sub> × time effect on TOC content (P<0.05) shows a clear difference between the slopes of the temporal trends for elevated and ambient CO<sub>2</sub> levels (Figure 1).

While TOC content increased (585-kg C ha<sup>-1</sup> y<sup>-1</sup>) under elevated CO<sub>2</sub>, it increased marginally (18-kg C ha<sup>-1</sup> y<sup>-1</sup>) under

ambient CO<sub>2</sub> concentrations. Increased soil C sequestration under elevated CO<sub>2</sub> was anticipated as a result of greater inputs of high C:N crop residues [6,37], slower decomposition [23], fungi dominated microbial processes [15], and greater bio-physical and/or biochemical protection [22,38,39]. Since decreased soil N may limit microbial utilization of C, greater availability of N-poor substrates under elevated CO<sub>2</sub> might reduce decomposition of crop residues. A slower decomposition implies a longer retention of crop residue and eventually greater accumulation of TOC. In addition to slower and higher C assimilation, mycorrhizal associations might contribute in retaining C as particulate organic matter (POM) by enhancing macro-aggregation through bio-physical enmeshing of micro-aggregates [20,22,40,41] or releasing polysaccharides to cement primary particles, POM, and micro-aggregates together to form or stabilize macro-aggregates [21,38,41,42]. A deposition of soybean and wheat roots and mycorrhizal hyphae within pore-spaces [38] and comminuted litter detritus, as POM, may also have incorporated in soil casts and macro-aggregates by the feeding and casting activities of earthworm and other faunas [21].

Soil acidification by legumes (e.g. soybeans) has promoted micro-aggregate formation through complex physico-chemical interactions of plant and microbial derived C compounds with di- and polyvalent metal ions and clays [29,38,39,43]. Since aggregate formation is directly influenced by plant root growth and exudations and indirectly by microbes-faunal activities [42], the increased accumulation and protection of C as POM within macro- and micro-aggregates, are the most important mechanisms enabling greater sequestration of TOC under elevated CO<sub>2</sub> concentrations.

Our results supports most of previous findings [6,31,32,43,44]. It has been reported that soil C sequestration asymptotically increased by 3.3% over a 3- year period [6]. A meta-analysis has shown that mineral soil acts as a sink of C in response to elevated CO<sub>2</sub> [32]. Under elevated CO<sub>2</sub>, prairie soil sequestered 336±96 kg C ha<sup>-1</sup> yr<sup>-1</sup> at 0 to 15-cm depth [33], which is slightly lower than our results, perhaps due to continuous NT soybean-wheat rotation we used. Since C and N are stoichiometrically linked in SOM, a higher input of biomass N from soybeans might contribute to higher soil C sequestration in our study.



**Figure 1** Carbon dioxide and time effects on total soil organic carbon (TOC) content.

Regardless of the mechanism of the effects, an important finding of our study is the elevated CO<sub>2</sub> impact on TOC under continuous NT cropping systems because an incomplete and slow decomposition of high C:N crop residue by fungal food webs may accelerate dissolved organic matter (DOM) production. Such an increase could be important in fueling soil denitrification processes.

Through time, soil TOC remained unchanged under reduced O<sub>3</sub>, but it showed an increasing trend that did not reach statistical significance (i.e., slope of temporal increase was not significant) (Figure 2). Moreover, the interaction (test of the difference between the two slopes) was not significant. However, averaged over time, soil TOC was significantly higher under elevated O<sub>3</sub> compared with reduced O<sub>3</sub>.

This could be due to greater fine root production and reduced decomposition of partially recalcitrant crop residue as influenced by O<sub>3</sub> fumigation under continuous NT [14,21,22]. Four year exposure of soybean-wheat to elevated O<sub>3</sub> might have increased plant O<sub>3</sub>-tolerance of the crops [46]. A less O<sub>3</sub>-responsive plants may cause a higher allocation of C to roots, leading to increased plant root biomass and soil TOC under elevated O<sub>3</sub> [19]. This is contrary to findings that elevated O<sub>3</sub> reduced soil C [2,46].

Although the effects of CO<sub>2</sub> × O<sub>3</sub> × time interaction on TOC were not significant, TOC was higher under the combination of elevated CO<sub>2</sub> and O<sub>3</sub> compared with their reduced levels. This could be due to the combined effects of high C:N litter inputs produced by CO<sub>2</sub> fertilization and partially recalcitrant leaf litters, increased fine root production under O<sub>3</sub> exposures, and slower decomposition of crop residues plus increased biophysical protection of POM under continuous NT over the years [6,13-15,22-24,38,39].

The temporal trends for soil TN differed for elevated CO<sub>2</sub> and O<sub>3</sub> when compared to ambient CO<sub>2</sub> and low O<sub>3</sub> treatments, respectively (Figures 3 and 4). These are indicated by the significance of CO<sub>2</sub> × time and O<sub>3</sub> × time interactions (P<0.01). Soil TN decreased over time under both CO<sub>2</sub> levels but the rate of decrease was significantly greater under elevated CO<sub>2</sub> (-216 kg N ha<sup>-1</sup> y<sup>-1</sup>) than under ambient CO<sub>2</sub> (-152 kg N ha<sup>-1</sup> y<sup>-1</sup>) treatment (Figure 3).

This implies that CO<sub>2</sub> fertilization reduced soil TN content @ 64 kg ha<sup>-1</sup> y<sup>-1</sup> as compared to ambient CO<sub>2</sub>. Soil TN decreased over time perhaps due to continuous harvest of N-enriched grain/seed or feed, which exceeds the amount of TN returned to soil through fertilization, recycling of plant residues and atmospheric inputs. Results from several previous studies indicate that CO<sub>2</sub> might enhance root growth and facilitate greater soil exploration, thereby increasing N and other nutrients uptake to maintain the CO<sub>2</sub> fertilization effects on plants [39,47]. Although soil N uptake may increase to fulfill a greater overall plant growth, foliar N may decrease in response to greater flux of C-H enriched compounds, leading to a decreased N in litters [29,37,48]. The C:N of plant residues produced at elevated CO<sub>2</sub> is higher than that of native SOM, and an additional amount of N is required to sustain the process of residue decomposition, especially under a net positive C balance in soil [22]. In response to progressive N deficiency, and to fulfill the increasing demand for N and other nutrients, soil microbes often forced to mineralize relatively low C:N native SOM instead of decomposing high C:N plant residue [6,25,26].

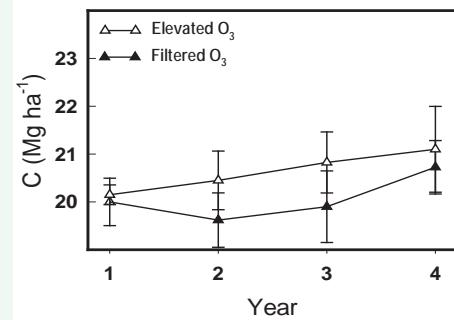


Figure 2 Ozone and time effects on total soil organic carbon (TOC) content.

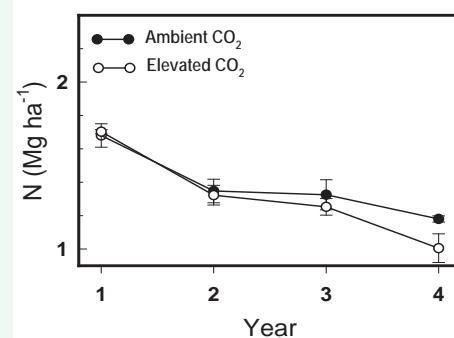


Figure 3 Carbon dioxide and time effects on soil total nitrogen (TN) content.

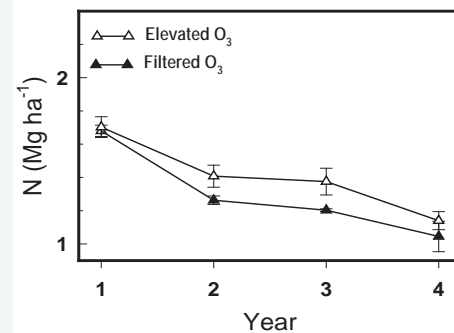


Figure 4 Ozone and time effects on total soil nitrogen (TN) content.

The mineralization of native SOM would eventually release soil N available to plants through microbial biomass turnover and a temporary positive feedback on plant growth, which would increase N uptake even further. Therefore, enhanced plant uptake of N and its removal from the site by harvest in combination with mineralization of native SOM for increased microbial N demand in response to greater amount of high C:N litters produced under elevated CO<sub>2</sub> may likely result in C-H enriched but N-deficient SOM over time through alterations in the stoichiometry of C, N and H. The effects of progressive decreases in N availability will have a detrimental effect on plant growth, and consequently on soil C storage [6].



In contrast, the temporal N depletion rate was smaller under elevated  $O_3$  ( $-172 \text{ kg N ha}^{-1} \text{ y}^{-1}$ ) compared to low  $O_3$  ( $-97 \text{ kg N ha}^{-1} \text{ y}^{-1}$ ) treatment (Figure 4).

The elevated  $O_3$  treatment resulted in an excess of  $25 \text{ kg}$  of  $N$   $\text{ha}^{-1} \text{ y}^{-1}$  remaining in the soil as compared to low  $O_3$  treatment. Again, reduced N over time by both  $O_3$  treatments might be due to increased N uptake by plants and continuous harvest of N-enriched grain/seed. Using SOM enriched in  $^{15}\text{N}$ , it has been reported that soybeans grown under elevated  $O_3$  obtained more N from soil than plants grown under low  $O_3$  due to reduction in N-fixation caused by decreased photosynthetic translocation of  $^{13}\text{C}$  to nodules [49,50]. However, reduced temporal N depletion under elevated  $O_3$  compared with low  $O_3$  in the current study is perhaps due to replenishment of N from incomplete breakdown of the litter fractions. Elevated  $O_3$  can cause plant foliage to age prematurely with proportionally more N to be bound strongly with recalcitrant lignin and phenolic compounds, and decrease N reabsorption before senescence and/or abscission of leaves. This is supported by the findings that prematurely abscised litters produced at elevated  $O_3$  had a higher N content [13]. Despite having a relatively higher N content, such plant litters can be more recalcitrant to microbial decomposition [14]. The recycling of N-enriched prematurely abscised plant litters in soil, may in turn, favor an accumulation of recalcitrant SOM, while increasing mineralization of native SOM due to progressive lacking of labile C to perform soil biological processes.

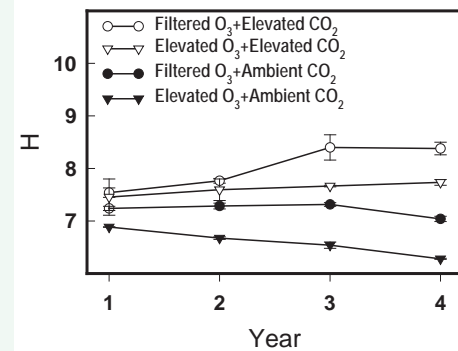
For the H content, all slopes of the temporal trends under the four combinations of two  $\text{CO}_2$ -by-two  $O_3$  levels were significantly ( $P < 0.01$ ) different from zero (Figure 5).

The H content increased over time under elevated  $\text{CO}_2$  irrespective of the  $O_3$  levels, which differed significantly from each other ( $316$  vs.  $91 \text{ kg H ha}^{-1} \text{ y}^{-1}$ , respectively). By contrast, the H content decreased under ambient  $\text{CO}_2$  irrespective of the  $O_3$  regimes, which also differed significantly from each other ( $-195$  vs.  $-57 \text{ kg H ha}^{-1} \text{ y}^{-1}$ , respectively). Greater translocation of C-H-O enriched compounds belowground through fine root productions, exudations, sloughed cells, and lysate turnover [23,24,50], accounts for much of the increase in H content ( $316 \text{ kg H ha}^{-1} \text{ y}^{-1}$ ) under elevated  $\text{CO}_2$  + low  $O_3$  treatment than under elevated  $\text{CO}_2$  + elevated  $O_3$  ( $91 \text{ kg H ha}^{-1} \text{ y}^{-1}$ ) treatment. Higher proportion of lignin and other structural carbohydrates in litters as reported elsewhere [13] may be the most important factor in explaining relatively higher depletion of H in native SOM under ambient  $\text{CO}_2$  + elevated  $O_3$  ( $-195 \text{ kg H ha}^{-1} \text{ y}^{-1}$ ) than ambient  $\text{CO}_2$  + low  $O_3$  ( $-57 \text{ kg H ha}^{-1} \text{ y}^{-1}$ ) treatment.

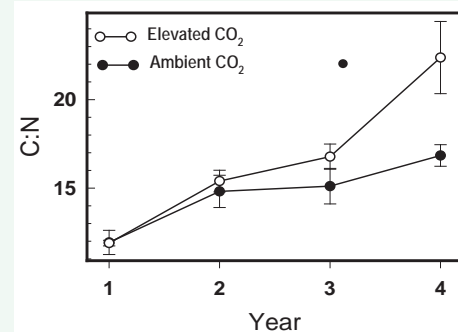
As indicated by the significant  $\text{CO}_2 \times \text{time}$  and  $O_3 \times \text{time}$  effects ( $P < 0.01$ ) on the C:N, the temporal increase in C:N was greater under elevated  $\text{CO}_2$  compared to ambient  $\text{CO}_2$  (Figure 6) and under low  $O_3$  compared to elevated  $O_3$  (Figure 7).

Similar patterns were observed for the H:N, but the slope differences were even greater (Figures 8 and 9).

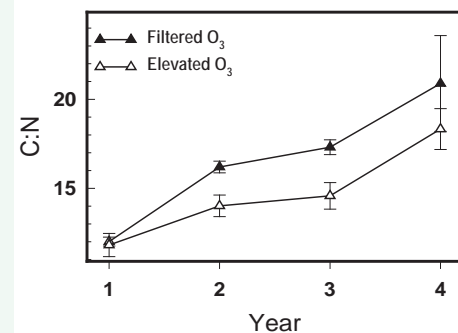
Greater flux of C-H enriched compounds with simultaneous dilution of N in plant litters, mineralization of native SOM and subsequent harvest of large amounts of N in grain/seed, may have translated into the resulting high C:N and H:N under



**Figure 5** Carbon dioxide, ozone and time effects on total soil hydrogen (H) content.



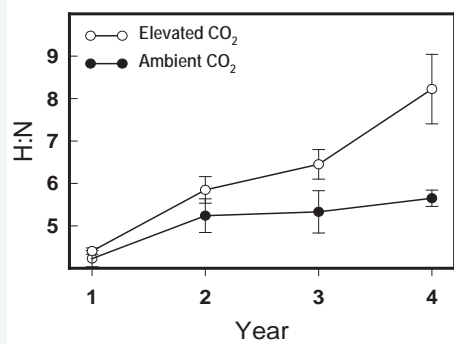
**Figure 6** Carbon dioxide and time effects on soil carbon:nitrogen (C:N).



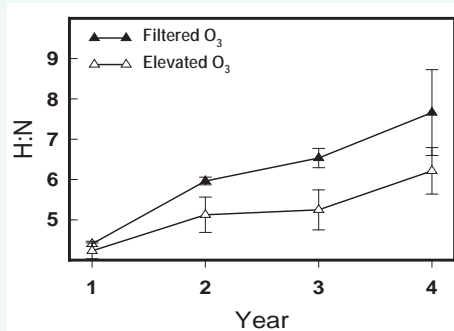
**Figure 7** Ozone and time effects on soil carbon:nitrogen (C:N).

elevated  $\text{CO}_2$  (Figures 6-8, respectively). Relatively smaller temporal increase in the C:N and H:N under elevated  $O_3$  (Figures 7-9, respectively) may be due to recycling of phenolic-N enriched prematurely abscised plant litters recalcitrant to incomplete microbial decomposition.

A significant  $\text{CO}_2 \times O_3$  effect ( $P < 0.01$ ) was detected for the C:H, but multiple mean comparisons showed no significant difference among the temporal changes in the C:H under the four simple effects (data not shown). The long-term exposure of plants to elevated  $\text{CO}_2$  and  $O_3$  concentrations may have produced neutralizing effect on C:H due to protective roles of elevated  $\text{CO}_2$  against  $O_3$  induced damage on plants and conversely, reducing the



**Figure 8** Carbon dioxide and time effects on soil hydrogen:nitrogen (H:N).



**Figure 9** Ozone and time effects on soil hydrogen:nitrogen (H:N).

beneficial effects of CO<sub>2</sub> on plant growth and biomass production by elevated O<sub>3</sub> treatment [35,36].

## CONCLUSION

After five years of elevated CO<sub>2</sub> and O<sub>3</sub> exposure treatments on no-till soybean-wheat rotation in the open top field chambers, the results showed that C, N and H dynamics of SOM have been partially altered. Soils with crops under long-term exposure to elevated CO<sub>2</sub> alone acted as temporary and/or transient sinks of C. Results suggest that soils with increases in C-H sequestration eventually become N-limited due to stoichiometric changes in C, N and H contents of the young and/or newly formed soil organic matter. In contrast, long-term effects of elevated O<sub>3</sub> (+O<sub>3</sub> and O<sub>3</sub>+CO<sub>2</sub>) on plants have caused a small accumulation of recalcitrant C in soil, which, in turn, would reduce the lability of C necessary to perform ecosystem services. Increasing concerns are raised whether more labile C-H with N deficient or recalcitrant C accumulation in new soil organic matter under elevated CO<sub>2</sub> or O<sub>3</sub> will cause negative agro ecosystem-level feedbacks and constrain plant productivity. Increased C sequestration, in particular, under elevated CO<sub>2</sub> could elicit a greater formation and leaching of dissolved organic matter and consequently fueling soil denitrification processes.

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