

## Review Article

# Hypertension, Mechanical Force, and Renal Disease

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## Abstract

Hypertension or high blood pressure is a major public health problem and a leading cause of disability and death in the world. Mechanical forces generated from blood pressure and blood flow are responsible for cell metabolism, growth, proliferation, migration, differentiation, and morphology, but also lead cell detachment, injury, and apoptosis. Mechanical force-induced apoptosis could damage various organs including the heart, kidneys, lungs, brain and other vasculatures, and cause different pathogenesis such as premature vascular, renal and cardiovascular mortalities. One-quarter of the world's adult population has hypertension, and the increasing economic burden of hypertension has been attributed to economic progress lagging in the developing countries and quality of life in the developed countries. In this review, we summarize the recent progresses in understanding the molecular mechanisms of hypertension and renal pathogenesis, and exploring the regulation of signaling network in mechanical force stimulation. These advances have significantly increased our understanding of the connection between hypertension-mechanotransduction and mechanical stress-pathogenesis, and lead us to search for novel potential targets and strategies in the treatment of hypertension and other related diseases.

## Keywords

- Hypertension
- Mechanical force
- Cell signaling
- Gene expression
- Renal disease

## ABBREVIATIONS

All: Angiotensin II; AT<sub>1</sub>: All receptor type 1; AP-1: Activator Protein-1; CNP: c-type Natriuretic Peptide; COX: Cyclooxygenase; ECM: Extracellular Matrix; Egr-1: Early growth response gene-1; ENaC: Epithelial Sodium Channel; ERK: Extracellular Signal-regulated Kinase; ET-1: Endothelin-1; FAK: Focal Adhesion Kinase; G: G protein; GADD 153: Growth Arrest and DNA Damage-inducible gene 153; GEF: Guanine nucleotide Exchange Factor; GFs: Growth Factors; GPCRs: G Protein Coupled Receptors; HIF: Hypoxia-inducible Factor; ICAM: Intercellular Adhesion Molecule; IGF: Insulin-like Growth Factor; IL: Interleukin; JNK: c-Jun N-terminal protein Kinase; MAPK: Mitogen-activated Protein Kinase; MCP: Monocyte Chemotactic Protein; MMP: Matrix Metalloproteinase; MS: Mechanical Sensitive; mTOR: Mammalian Target of Rapamycin; NOS: Nitric Oxide Synthase; PCNA: Proliferation Cell Nuclear Antigen; PI-3K: Phosphatidylinositol-3 Kinase; PKA: Protein Kinase A; PKB: Protein Kinase B; PKC: Protein Kinase C; PLA<sub>2</sub>: Phospholipase A<sub>2</sub>; PLC: Phospholipase C; PLD: Phospholipase D; PTKs: Protein Tyrosine Kinases; REB: Response Element-Binding Protein; RGS: Regulator of G protein Signaling; RTK: Receptor Tyrosine Kinase; SGK: Serum-Glucocorticoid-induced protein Kinase; SM22: Smooth Muscle cell specific protein; SOD: Superoxide Dismutase; THA2: Thromboxane Synthase A2; TLR: Toll Like Receptor; TNF: Tumor Necrosis Factor; T-PA: Tissue Plasminogen Activator; TRAAK: TWIK-Related Arachidonic Acid-stimulated K<sup>+</sup> channel;

TREK: TWIK-Related K<sup>+</sup> channel; TRP: Transient Receptor Potential; VCAM: Vascular Cell Adhesion Molecule.

## INTRODUCTION

Hypertension or high blood pressure is a chronic medical condition characterized by a sustained increase in vasoconstriction and attenuated vasodilation in the face of elevated mechanical stress in the blood vessel wall [1]. This requires the heart to work harder than normal to circulate blood through the blood vessel, and generates more mechanical force pushing against the blood vessel wall. Mechanical forces are responsible for the modulation of blood vessel size and morphology in the development and for the regulation of cell signaling network in cell metabolism, growth, proliferation, migration, differentiation, detachment, injury, and apoptosis [2-4]. Hypertension is a progressive cardiovascular syndrome that can damage various organs including the heart, kidneys, lungs, brain and other vasculatures, and lead to premature morbidity and death [1-6]. The relationship between hypertension and chronic kidney disease remains enigmatic and a matter of considerable clinical and academic interest with evidence supporting that hypertension is both a cause and a consequence of chronic kidney disease [3-7]. On the one hand, high blood pressure predicts end-stage renal disease in diabetic and nondiabetic patients [5-7]. On the other hand, renal malfunction predicts later onset of hypertension [7,8]. In this brief review, we discuss how mechanical forces generated by high blood pressure

and fast blood flow in hypertensive condition regulate signaling networks to modulate cellular function, and lead to chronic kidney disease.

### Hypertension and mechanical force

The vascular wall is an integrated functional component of the circulatory system. Hypertension leads to a chronic increased mechanical force on the vessel wall due to high blood pressure and fast blood flow [1,2]. To adapt to increased mechanical forces, vascular endothelial cells, vascular smooth muscle cells, and the surrounding environment undergo structural and functional changes known as vascular remodeling. Multiple mechanisms underlie the remodeling process, including increased expression of humoral factors and their receptors, adhesion molecules, integrins and their receptors, and many enzymes as well as related proteins [9,10]. The remodeling process appears to collaborate and interact in the response to pressure elevation. The most relevant mechanical forces that influence vessel size and morphology are shear stress, circumferential stress, and axial stress [11,12].

Shear stress is the tangential force that a fluid (blood) exerts parallel to the vessel surface due to friction of the blood against the vessel wall. Circumferential stress describes the perpendicular force that the intraluminal pressure applies on the vessel wall. Axial stress in blood vessels is defined by the exerted longitudinal force, vessel radius, and wall thickness and governs length adaptations. Many commercially available devices have been used *in vitro* cell models to study the effect of different mechanical forces on cell responses. Exposure of cells cultured as a monolayer to these instruments (Flexercell strain unit with a uniaxial circular well device, a linear stretching tub, biaxial cyclic cell strain devices, a parallel-plate flow channel and perfused transcapillary co-culture system) provides different patterns of mechanical forces.

Upon long-term exposure to blood pressure and blood flow, the structural and functional properties of vessels are modified to accommodate these changes in pressure by vascular remodeling [10,13]. This remodeling encompasses increased cell growth, proliferation, migration, differentiation and apoptosis, as well as alteration of the extracellular matrix (ECM) and vessel wall shape and composition [10-13]. This remodeling also enables arteries to withstand the increased pressure load. Consequently, arteries become more rigid and have a reduced compliance than in their native state, which decreases their ability to dampen the cyclic changes in blood pressure [10]. The progressive stiffening ultimately results in altered gene expression, signal transduction and cell functions, and causes various clinical complications [4-14]. A raised peripheral resistance due to harder and more rigid arteries leads to more severe hypertension and hypertension-induced changes in different levels.

The kidneys are one of the key organs affected by mechanical forces in hypertension patients. The glomerulus is a highly specialized structure that is the site of plasma ultrafiltration and urine production. The glomerular capillary wall, composed of glomerular endothelial cells, glomerular basement membrane, and glomerular epithelial cells (podocytes), is exposed to mechanical forces *in vivo* arising from capillary blood pressure and blood

flow. Podocytes, consisting of a cell body, major processes and foot processes interlinked by slit diaphragms, may experience lateral stretch as a capillary dilates, force perpendicular to the plane of the basement membrane (vertical traction) as they balloon in response to increased filtrate flow, or shear force from increased filtrate flow [15,16]. In mechanically stressed podocytes, the processes of podocytes become thinner and more elongated, whereas the cell body size decreases. Through an actin-based contractile apparatus, podocytes counterbalance pressure within the underlying capillary to prevent outward ballooning of the vessel and to preserve the normal architecture of the cells [15-18]. How do the mechanosensitive cells convert physical stimuli to biochemical signals?

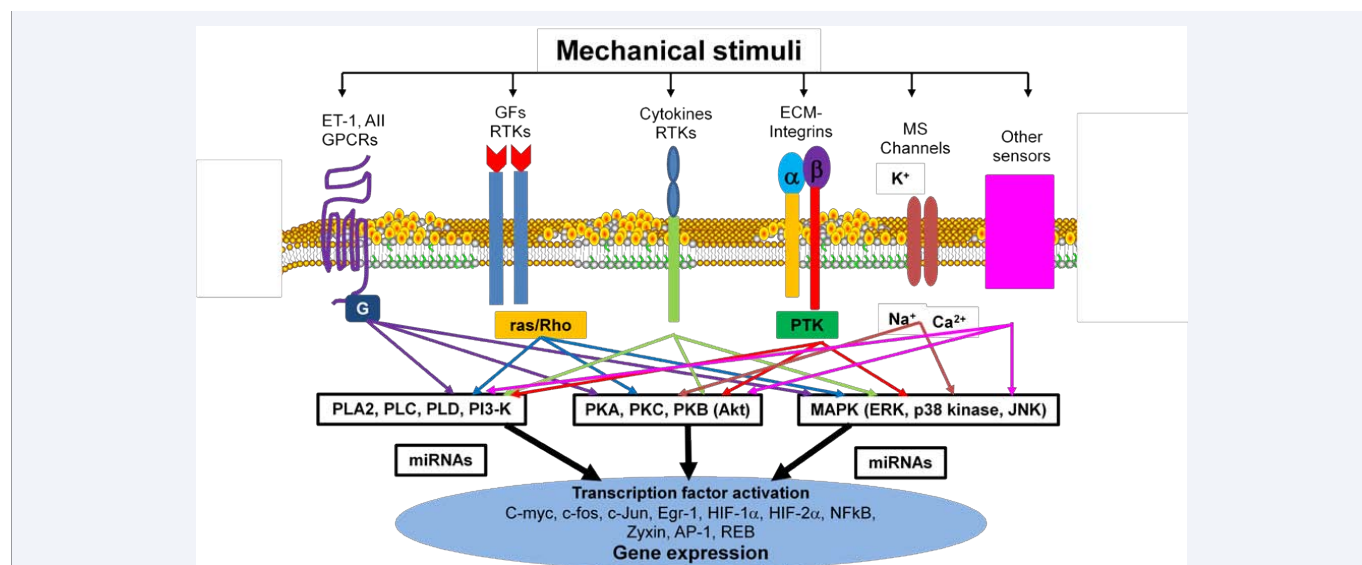
### Mechanical force regulates signaling network

As blood flows, cells on the vascular wall are constantly subjected to mechanical forces that appear to activate the same signaling pathways that are activated by hormones, growth factors, and inflammatory mediators. Mechanical stimuli can cause many important physiological responses which regulate cell functions in the chronic remodeling and development of organs. Changes in blood pressure and flow rate, such as hypertension, generate altered haemodynamic forces, and have been strongly implicated in the pathogenesis of cardiovascular disease and many other diseases [1,2]. Mechanical forces are sensed at the cell surface by sensors such as receptors, receptor tyrosine kinase (RTK), ion channels, and ECM-integrin complex, transduced through the cell by different signaling cascades, and activated different kinases or other enzymes that ultimately result in altered cell function (Figure 1) [19-23]. Although the molecular identity of the sensors has long remained elusive, it is clear that an increasing number of signaling pathways are involved in the responses to mechanical force.

### Membrane-mechanosensors

**Mechanoreceptors:** Cells are equipped with numerous receptors that allow them to detect and respond to micro-environment changes such as mechanical forces generated by blood pressure and blood flow. Exposure of vascular endothelial cells and smooth muscle cells to mechanical forces increase the synthesis of endothelin-1 (ET-1) and expression of ET receptor type B (ET<sub>B</sub> receptor) [23]. In podocytes, mechanical forces induce angiotensin II (AII) secretion and AII type 1 (AT<sub>1</sub>) receptor expression [24,25]. Both ET-1 and AII act through G protein coupled receptors (GPCR) to stimulate their downstream effectors. The regulation of blood pressure within the normal range requires precise kinetic control of GPCR signaling [26].

Many studies demonstrated that the levels of some growth factors and inflammatory mediators are also significantly increased when cells are exposed to mechanical forces [9,12-14]. Fluid shear stress differentially modulates the expression of genes encoding basic fibroblast growth factor (bFGF) and platelet-derived growth factor (PDGF) B chain in vascular endothelium and smooth muscle cells, and stimulates extracellular signal-regulated kinases (ERK) signaling pathway [27,28]. In human periodontal ligament cells mechanical stress induces immune response genes expression: cytokines (TNF- $\alpha$ , IL-1 $\beta$ ), chemokines (IL-8, CCL-20), human  $\beta$ -defensin-2 and-3, and Toll-



**Figure 1** Schematic representation of cell mechanotransduction. Mechanical forces induce the activation of mechanical sensors on the plasma membrane which can convert physical stimuli to biochemical signals. As a result, these signals further activate different signal pathways which lead to regulate gene expression and to modulate cellular function.

like receptors (TLR-2 and TLR-4) in a force- and time-dependent manner [29]. In mouse models, physical force regulates fibrosis through inflammatory focal adhesion kinase (FAK)-ERK-monocyte chemoattractant protein-1 (MCP-1) pathways [30,31]. Through their receptors, these growth factors and inflammatory mediators regulate cell growth, proliferation, differentiation and apoptosis via many different signaling pathways. Now, increasing evidence indicates that the activation of growth factor receptors and inflammatory mediator receptors in response to increased mechanical stress plays a critical role in cell signaling regulation of hypertension [32].

**Integrin-extracellular matrix:** Integrins exist as  $\alpha\beta$  pairings that interact with ECM components including fibronectin (ligand for  $\alpha5\beta1$  and  $\alpha\beta3$ ), vitronectin (ligand for  $\alpha\beta3$ ), and laminin (ligand for  $\alpha6\beta1$ ). Many of these components sense mechanical forces and their responses depend on specific integrin-ECM interactions [33-36]. Mechanical forces cause direct stretching of cell surface integrin binding sites. Force-induced conformational changes in the ECM may alter integrin structure and lead to activation of several signaling pathways within the cell [36]. Integrins connect the ECM to the cytoskeleton, and provide cells with mechanical anchorages and signaling platforms. Activation of these signaling pathways leads to altered regulation of genes that synthesize several enzymes and catabolize ECM proteins [9,10,13,14]. On the other hand, these pathways are also involved in the deformation of gap junctions to alter cell-to-cell communications [37]. The increased intracellular  $Ca^{2+}$  concentration is associated with mechanical stimulation in the cell membrane and intercellular gap junctions [38].

**Mechanosensitive ion channels:** Mechanical forces cause a change in membrane potential of the cell resulting from the opening of mechanosensitive ion channels [39,40]. As mechano-electrical molecular switches, these channels convert mechanical forces exerted on the cell membrane into electrical or biochemical signals in physiological processes. Mechanosensitive ion channels

exist in all cells, and respond to mechanical forces along the plane of the cell membrane. These channels can be either non-selective depolarizing, permeable to  $Na^+$ ,  $K^+$  and  $Ca^{2+}$ , or hyperpolarizing channels selective for  $K^+$  [41]. For instance, recent work has identified and electrophysiologically characterized two members of a new family of two-pore-domain, weakly inward-rectifying  $K^+$  channels: TWIK-Related  $K^+$  channel (TREK) and TWIK-Related Arachidonic Acid (AA)-stimulated  $K^+$  channel (TRAAK) [42]. TREK channels are polymodal  $K^+$  channels (i.e. gated by a variety of chemical and physical stimuli) expressed in a variety of tissues, but are particularly abundant in the brain and heart. TRAAK is similar to TREK in that it can be activated by membrane tension and arachidonic acid. TRAAK is widely expressed in the brain, spinal cord, and retina, which indicates that it has a function wider than mechanotransduction in neuronal excitability [43]. The epithelial sodium channel (ENaC) belongs to the superfamily of amiloride-sensitive  $Na^+$  channels of the transporting epithelia and degenerins (MEC/DEG channels), many of which are suspected to be directly gated by mechanical stimuli [44,45]. The MEC/DEG subfamily of degenerins is responsible for swelling-induced neuronal degeneration in nematodes. The subfamily includes the MEC-4, MEC-6 and MEC-10 proteins, which are thought to function as subunits of a mechanosensitive ion channel that might have a role in touch sensitivity [46,47]. Transient receptor potential (TRP) proteins constitute a large non-voltage-gated cation channel superfamily, activated polymodally by various physicochemical stimuli, and are implicated in a variety of cellular functions [48]. In mammals many TRP family members such as TRPC1, TRPC3, TRPC6, TRPM4, TRPM7, TRPN1, TRPA1, TRPY1, TRPP1, TRPP2, and notably, TRPV1, TRPV2 and TRPV4, have been reported to be involved in mechanotransduction [49]. TRPs are up-regulated in animal models of congestive heart failure or hypertension models [50]. TRPC6, TRPM6, and TRPP2 have been implicated in hereditary focal segmental glomerulosclerosis, hypomagnesemia with secondary hypocalcemia, and polycystic kidney disease, respectively [51]. In addition, TRPV5 contributes

to several acquired mineral dysregulations, such as diabetes mellitus, acid-base disorders, diuretics, immunosuppressant agents, and vitamin D analogues-associated  $\text{Ca}^{2+}$  imbalance, whereas TRPV4 may function as an osmoreceptor in the kidneys and participate in the regulation of sodium and water balance [52].

### Intracellular transducers and effectors

**G protein, Rho and GEF:** Mechanical forces increase the levels of AII in podocytes [24,53] and ET-1 in vascular smooth muscle cells [23] and endothelial cells [54]. These hormones bind to their receptor to activate heterotrimeric G protein and to promote their signaling pathways. Mechanical stimuli also induce conformational changes of mechanosensitive proteins. Instead of mechanical-induced AII release, Zou et al. [53] demonstrated that mechanical stress activates  $\text{AT}_1$  receptor without the involvement of AII in cardiomyocytes. The role of G protein in hypertension is also demonstrated by genetic studies. A genetic ablation of  $\text{G}\alpha_q$  in mice has been shown to result in a cardiac malformation and craniofacial defects [55], whereas an overstimulation of the  $\text{G}\alpha_q$  pathway in mice has been shown to result in the development of hypertrophic cardiomyopathy [56-58]. Regulator of G protein signaling 2 (RGS2) can selectively attenuate  $\text{G}\alpha_q$  signaling [59]. RGS2 knockout mice (RGS2<sup>+/-</sup> and RGS2<sup>-/-</sup>) exhibit a strong hypertensive phenotype, renovascular abnormalities, persistent constriction of the resistant vasculature, and prolong response of the vasculature to vasoconstrictors *in vivo* [60,61]. Many current pharmacological therapies of hypertension focus on the regulation of vascular resistance by blocking hormones such as catecholamines and AII to activate their receptors and G protein signaling.

Small GTPases such as the *ras* and Rho families are associated with actin-based cytoskeletal structure [62]. Rho proteins act as molecular switches to control cellular processes by cycling between an active GTP-bound state and an inactive GDP-bound state. Recent investigations have shown that shear stress mediates Rho-dependent cytoskeletal alignment and directional migration of the endothelial cells [63,64]. The inhibition of the Rho-associated kinase p160ROCK eliminates the shear-enhancement of migration speed [65]. We recently reported the role of Rho in mechanical force-activated renal epithelial phospholipase D [66]. All these data indicate that Rho, as a mechanosensitive motor, plays an important role in the regulation of cell function.

Some guanine nucleotide exchange factors (GEFs) specific for RhoA have been found to associate with the cytoskeleton and adhesions [67]. For example, vav2 was reported to be phosphorylated and activated in response to cyclic stretch in mesangial cells [68], and mechanical force activates RhoA through two GEFs, GEF-H1 and LARG in rat fibroblasts [69]. We recently found that mechanical force-activated phospholipase D is mediated by  $\text{G}\alpha_{12/13}$ -Rho [66]. It would be interesting to determine if p115RhoGEF activity is regulated by mechanical forces. Recent studies indicate that some RhoA GTPase activating proteins (GAPs) also play a role in the response to mechanical forces. In endothelial cells, shear stress regulates p190RhoGAP activity in a biphasic pattern [70].

**Protein kinases:** Cell mitogenic activities are stimulated by

growth factors and other mitogens which are mediated through mitogen-activated protein kinases (MAPK) in the regulation of cell function. Exposure of cells to mechanical forces increases the release of several growth factors in the cells, and these growth factors further induce ERK activation [71,72]. Parallel to the ERK signaling pathway, mechanical forces also elevate the levels of endotoxin and cytokines such as tumor necrosis factor (TNF)- $\alpha$  and interleukin (IL), and these stimuli activate p38 MAPK [73] and c-Jun NH<sub>2</sub>-terminal protein kinases (JNK, also called stress-activated protein kinases) [74]. ERK activation enhances the phosphorylation of the transcription factor *Elk1*, the acetylation of core histone, and the expression of the *Elk1* target gene, *c-fos* in the mediation of cell growth and proliferation, while JNK activation induces *c-Jun* phosphorylation in the regulation of cell programmed death.

A mechanical force-induced increase in cAMP content has been reported in fetal rabbit epithelial cells [75]. Increased cAMP concentration and induced protein kinase A (PKA) activation was also observed in lung tissues after partial pneumonectomy or mechanical ventilation [76]. Protein kinase B (PKB), also known as Akt, is a serine/threonine-specific protein kinase and a downstream effector of phosphatidylinositol-3 kinases (PI-3K). Mechanical stretch induces expression of insulin-like growth factor (IGF)-1 and its receptor, which activates IGF-1R/PI-3K/Akt in the regulation of proliferation of venous smooth muscle cells [77]. PI-3K/Akt pathway in mouse lung has significant protective effects in response to mechanical stress [78,79]. Protein kinase C (PKC) also controls the function of other proteins through the phosphorylation of their serine and threonine residues, and plays an important role in many of the pathologies of heart disease [80]. The activity of several major isozymes of PKC is regulated by the intracellular concentrations of free  $\text{Ca}^{2+}$  and diacylglycerol. Pulmonary arterial smooth muscle cells respond to mechanical force with a transient increase in inositol 1, 4, 5-trisphosphate and diacylglycerol leading to PKC activation [81]. PKC $\epsilon$  induces dramatic alterations in muscle cell shape, leading to an overall increase in cell length, length-to-width ratio, and perimeter-to-area ratio [82,83].

Protein tyrosine kinases (PTKs) are enzymes that catalyze the phosphorylation of tyrosyl residues. Two classes of PTKs are present in cells: the transmembrane receptor tyrosine kinases (RTKs) and the nonreceptor PTKs [84]. RTKs are induced by a large growth factor family, which enhance receptor catalytic activity or provide docking sites for downstream signaling proteins [85]. Many RTKs, including receptors for EGF, FGF, IGF and PDGF, have been proposed as mechanoreceptors in a variety of tissues [86,87]. Nonreceptor PTKs (e.g. FAK and c-Src) represent cellular enzymes that have intrinsic kinase activity but do not possess an extracellular domain. Both FAK and c-Src mediate cell migration by promoting membrane protrusion and focal adhesion turnover utilizing several signaling pathways. Desai et al. reported that mechanical stretch decreases FAK phosphorylation and reduces cell migration in airway epithelial cells [88]. We recently demonstrated that shear stress induces cell apoptosis via a c-Src in the cultured podocytes [89]. Shear stress also regulates endothelial nitric oxide synthase (NOS) expression through c-Src [90].

**Other enzymes:** Mechanical forces have been documented to stimulate phospholipase A<sub>2</sub> leading to lung inflammation and acute lung injury [91]. Exposure of rat lung cells to mechanical strain activates phospholipid turnover via phospholipase C, followed by PKC activation in the regulation of cell proliferation [92]. Mechanical stress stimulates phospholipase C activity and elevates intracellular calcium ion levels in neonatal rat cardiomyocytes [93]. Shear stress induces cell apoptosis via a c-Src-phospholipase D-mTOR signaling pathway in the cultured podocytes [89]. Mechanical stimulation of skeletal muscle induces phospholipase D activation, PA accumulation, and mTOR signaling [94]. Mechanical forces stimulate membrane phospholipid metabolism through different enzymes to generate several bioactive lipid molecules, which is the key step of the conversion of mechanical stimuli to biochemical signals in the cells [94,95]. Metabolism of the ECM and reorganization of the cytoskeleton are also responded to mechanical stimulation [9-14].

### Nucleus where mechanical forces regulate gene expression

There are many studies supporting that mechanical forces provide an important context for cell growth and differentiation, tissue genesis and maintenance, and organ development and function. All these processes depend on the regulation of signaling networks in the cells [9-14,19-22]. At the molecular level, mechanical forces can regulate a variety of gene expression [9,14]. During development, mechanical forces cause changes in size, shape, number and position of cells by regulating cell growth, proliferation, differentiation, and polarity, and lead to organ development by coordinating cell-cell interactions, tissue spreading, compression, condensation, and self-organization [9-14,19-22,97]. These changes are therefore integral to any morphogenetic processes. The key in these processes is that mechanical forces modulate gene expression at the molecular level and regulate growth, proliferation, differentiation, migration and apoptosis at the cellular level.

The genes encoding for many proteins including vasoactive substances, growth factors, adhesion molecules, chemoattractant molecules, coagulation factors, antioxidant factors, and proto-oncogenes can respond to mechanical force stimulation by modulating mRNA levels during tissue morphogenesis and pathophysiology such as hypertension [9,14]. Although mechanical force induced gene expression has been studied for many years, significant progress has been made only due to microarray technology [28,29]. The technique enables simultaneous measurement of the transcriptional response of thousands of genes.

Exposing *in vitro* cultured cells to mechanical forces generated by different commercial available devices and analyzing the changes with microarray technology, we can profile gene expression in response to mechanical forces. Gene expression is modulated by specific transcription factors (e.g. *c-myc*, *c-fos*, *c-Jun*, *Egr-1*, *AP-1*, *SP-1* and *NF-κB/Rel*) that bind to their target sequences in the promoter region of the gene. Using reporter systems, many laboratories have discovered mechanical force-inducible transcription factors, they are *c-fos*, *c-Jun*, *HIF-1α* and *2α*, *Egr-1*, *AP-1*, *SP-1*, *NF-κB/Rel*, *Zyxin*, and *REB* [9,14,97,98],

and these transcription factors are involved in force-induced gene expression. Mechanical forces increase the levels of *Angiotensin II*, *ET-1*, different growth factors (*EGF*, *FGF*, *PDGF*, *VEGF* and *TGF*), *nitric oxide*, *IL-6* and *8*, *CD44*, *TNF-α*, and *tissue factor* [9,98]. As agonists, these products can stimulate different signaling pathways in the regulation of the cell signaling networks.

Mechanical forces also modulate the mRNA levels of different receptors (*AT<sub>1</sub> receptor* and *ET-B receptor*), *ECM-integrin-cytoskeleton components* (*collagen*, *fibronectin*, *vitronectin*, *ICAM*, *VCAM*, *MCP*, *MMP*, *integrin*, *tensin*, *paxillin*, *actin*, *desmin*, and *calponin*) and *downstream effectors* (*COX-1*, *COX-2*, *SOD*, *FAK*, *catalase*, *SGK*, *NOS*, and *THA-2*) as well as many other proteins (*PCNA*, *GADD-153*, *SM22-a*, *CPR*, *MCP-1*, *t-PA*, *CNP*, *prostacyclin*, *adrenomedullin*, *thrombomodulin* and *tenasin*) [9,98]. Gene expression modulated by mechanical forces is also affected by different forms and magnitudes of forces. *Laminar pulsatile flow* and *non-laminar disturbed flow* leads to different gene expression profiles [97-99]. Some gene expression is increased at low shear stresses but decreased by moderate and high shear stresses [9,98]. Modulation of gene expression and activation of many enzymes in response to mechanical force stimulation play important roles in signaling networks and in the regulation of cellular functions.

Recent studies demonstrated that *microRNAs (miRNA)*, small noncoding RNAs, participate in the regulation of gene expression by interacting with the 3'-UTR of the target mRNA. This interaction results in mRNA degradation and/or inhibition/activation of protein translation [100]. *Turczyńska et al.* reported that *miRNA-145* is essential for stretch-induced L-type calcium channel expression in vascular smooth muscle contractile differentiation [101]. *MiRNA-146a* is also a mechanosensitive miRNA that is rapidly up-regulated by oscillatory pressure and plays an important role in regulating mechanically induced inflammation in lung epithelia [102]. Stretch-induced activation of *AMPK* in vascular smooth muscle is in part regulated by reduced levels of *miRNA-144/451* [103]. *MiRNAs* appear to play an important role in cell signaling regulated by mechanical forces. Altering *miRNA* expression levels can prevent and even reverse the acquisition of cell synthetic phenotype *in vivo*, thereby implicating *miRNAs* as exciting future therapeutic targets for vascular proliferative disease.

Mechanical forces play an important role in the regulation of cell signaling networks, however, which signaling pathway could be activated and how many signaling pathways would be activated are dependent on cell types, cell states (development or pathogenesis) and force patterns. On the other hand, mechanical forces lead to the alteration of agonists, receptors, transducers and downstream effectors, these changes could show cross effect in the regulation of cell signaling networks. Meanwhile, mechanical forces induce different signaling pathways at different time frames such as channels and *PI-PLC* activation in seconds, protein phosphorylation in minutes and gene expression in hours. This complex system remains largely unknown.

### Hypertension and chronic kidney disease

Hypertension represents a complex, multifactorial disease and contributes to the major causes of morbidity and mortality

in industrialized countries: ischemic and hypertensive heart disease, stroke, peripheral atherosclerosis and renal failure [1,4,104]. Under hypertensive conditions, mechanical stress induces the damage of podocytes which leads to cell hypertrophy, foot process effacement, cell body attenuation, pseudocyst formation, cytoplasmic overload with reabsorption droplets, and, finally, detachment from the glomerular basement membrane [105,106]. Following podocyte loss, the inability of podocytes to proliferate contributes to the development of glomerulosclerosis [15,17]. With loss of nephron mass, intraglomerular pressure increases and resident glomerular cells are exposed to increased mechanical forces. Glomerular capillary hypertension perpetuates further damage to resident podocytes, alters glomerular haemodynamics, fails to serve as a glomerular filtration barrier, and leads to proteinuria and progressive loss of kidney function resulting in glomerulosclerosis and end-stage renal failure [16,107-110].

Diabetic nephropathy is the most common cause of progressive chronic kidney disease in the developed countries. It is thought to result from interactions between metabolic (hyperglycaemia) and haemodynamic (glomerular hypertension) factors [107,111-114]. High glucose activates different signaling pathways within diabetic renal tissues. These signaling pathways induce oxidative stress, polyol pathway flux, hexosamine flux and accumulation of advanced glycated end-products as well as significant increases in angiotensinogen, AII and AT<sub>1</sub> receptor levels. Many of these signaling pathways also regulate podocyte apoptosis [112-114]. There is evidence from human disease that the number of podocytes is significantly reduced in both type 1 and type 2 diabetic patients [112-117]. Forces increases and damages resident glomerular cells

The mechanical properties of tissues and cells are commonly characterized by measuring their elasticity, their recoverable deformation in response to a force. Recently, many examples from clinical and whole animal studies have shown that changes in tissue stiffness are related to specific disease characteristics [118]. Using atomic force microscopy and a new technique, capillary micromechanics, to measure glomerular biomechanics in normal and two disease mouse models [Col4a3<sup>-/-</sup> mice (Alport model) and Tg26<sup>HIV/ml</sup> mice (HIV-associated nephropathy model)], one report demonstrated that the glomeruli from both disease models exhibited significant mechanical abnormalities (reduces 30% stiffness, the diseased renal glomeruli are getting soft) and were significantly more deformable than normal glomeruli [119]. This increased deformability of glomeruli could directly contribute to disease by permitting increased distension with haemodynamic force or represent a mechanically inhospitable environment for glomerular cells.

In summary, hypertension is a major risk factor for stroke, heart attacks, heart failure, aneurysms of the arteries (e.g. aortic aneurysm), peripheral arterial disease, and is a cause of chronic kidney disease. Hypertension-related nephrosclerosis is a top cause of progressive renal damage and failure. Hypertension-caused injury of podocytes; loss of nephron mass, proteinuria and progressive loss of kidney function are an accompanying modulation of different gene expression and signaling activation. Understanding the molecular mechanism of hypertension and

podocyte injury has high potential to identify novel target(s) for pharmacologic consideration and to search for new therapeutic strategies for patients with hypertension and renal disease.

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## REFERENCES

- Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *J Clin Invest.* 2012; 122: 4306-4313.
- Chiu JJ, Chien S. Effects of disturbed flow on vascular endothelium: pathophysiological basis and clinical perspectives. *Physiol Rev.* 2011; 91: 327-387.
- Urban D, Ewen S, Ukena C, Linz D, Böhm M, Mahfoud F. Treating resistant hypertension: role of renal denervation. *Integr Blood Press Control.* 2013; 6: 119-128.
- Endlich N, Endlich K. The challenge and response of podocytes to glomerular hypertension. *Semin Nephrol.* 2012; 32: 327-341.
- Hsu CY, McCulloch CE, Darbinian J, Go AS, Iribarren C. Elevated blood pressure and risk of end-stage renal disease in subjects without baseline kidney disease. *Arch Intern Med.* 2005; 165: 923-928.
- Kobori H, Nangaku M, Navar LG, Nishiyama A. The intrarenal renin-angiotensin system: from physiology to the pathobiology of hypertension and kidney disease. *Pharmacol Rev.* 2007; 59: 251-287.
- Bakris GL, Ritz E. The message for World Kidney Day 2009: hypertension and kidney disease: a marriage that should be prevented. *Kidney Int.* 2009; 75: 449-52.
- Brantsma AH, Bakker SJ, de Zeeuw D, de Jong PE, Gansevoort RT. Urinary albumin excretion as a predictor of the development of hypertension in the general population. *J Am Soc Nephrol.* 2006; 17: 331-335.
- Anwar MA, Shalhoub J, Lim CS, Gohel MS, Davies AH. The effect of pressure-induced mechanical stretch on vascular wall differential gene expression. *J Vasc Res.* 2012; 49: 463-478.
- Lemarié CA, Tharaux PL, Lehoux S. Extracellular matrix alterations in hypertensive vascular remodeling. *J Mol Cell Cardiol.* 2010; 48: 433-439.
- Ando J, Yamamoto K. Vascular mechanobiology: endothelial cell responses to fluid shear stress. *Circ J.* 2009; 73: 1983-1992.
- Hoefer IE, den Adel B, Daemen MJ. Biomechanical factors as triggers of vascular growth. *Cardiovasc Res.* 2013; 99: 276-283.
- Mulvany MJ, Baumbach GL, Aalkjaer C, Heagerty AM, Korsgaard N, Schiffrin EL, et al. Vascular remodeling. *Hypertension.* 1996; 28: 505-506.
- Chiquet M, Gelman L, Lutz R, Maier S. From mechanotransduction to extracellular matrix gene expression in fibroblasts. *Biochim Biophys Acta.* 2009; 1793: 911-920.
- Pavenstädt H, Kriz W, Kretzler M. Cell biology of the glomerular podocyte. *Physiol Rev.* 2003; 83: 253-307.
- Haraldsson B, Nyström J, Deen WM. Properties of the glomerular barrier and mechanisms of proteinuria. *Physiol Rev.* 2008; 88: 451-487.
- Comper WD, Russo LM. The glomerular filter: an imperfect barrier is required for perfect renal function. *Curr Opin Nephrol Hypertens.* 2009; 18: 336-342.

18. Endlich N, Kress KR, Reiser J, Uttenweiler D, Kriz W, Mundel P, et al. Podocytes respond to mechanical stress in vitro. *J Am Soc Nephrol*. 2001; 12: 413-422.
19. Zhang H, Labouesse M. Signalling through mechanical inputs: a coordinated process. *J Cell Sci*. 2012; 125: 3039-3049.
20. Goehring NW, Grill SW. Cell polarity: mechanochemical patterning. *Trends Cell Biol*. 2013; 23: 72-80.
21. Alenghat FJ1, Ingber DE. Mechanotransduction: all signals point to cytoskeleton, matrix, and integrins. *Sci STKE*. 2002; 2002: pe6.
22. Shyy JY, Chien S. Role of integrins in endothelial mechanosensing of shear stress. *Circ Res*. 2002; 91: 769-775.
23. Cattaruzza M, Dimigen C, Ehrenreich H, Hecker M. Stretch-induced endothelin B receptor-mediated apoptosis in vascular smooth muscle cells. *FASEB J*. 2000; 14: 991-998.
24. Durvasula RV, Petermann AT, Hiromura K, Blonski M, Pippin J, Mundel P, et al. Activation of a local tissue angiotensin system in podocytes by mechanical strain. *Kidney Int*. 2004; 65: 30-39.
25. Humar R, Zimmerli L, Battegay E. Angiogenesis and hypertension: an update. *J Hum Hypertens*. 2009; 23: 773-782.
26. Brinks HL, Eckhart AD. Regulation of GPCR signaling in hypertension. *Biochim Biophys Acta*. 2010; 1802: 1268-1275.
27. Dardik A, Yamashita A, Aziz F, Asada H, Sumpio BE. Shear stress-stimulated endothelial cells induce smooth muscle cell chemotaxis via platelet-derived growth factor-BB and interleukin-1alpha. *J Vasc Surg*. 2005; 41: 321-31.
28. Malek AM, Gibbons GH, Dzau VJ, Izumo S. Fluid shear stress differentially modulates expression of genes encoding basic fibroblast growth factor and platelet-derived growth factor B chain in vascular endothelium. *J Clin Invest*. 1993; 92: 2013-21.
29. Lee SI, Park KH, Kim SJ, Kang YG, Lee YM, Kim EC. Mechanical stress-activated immune response genes via Sirtuin 1 expression in human periodontal ligament cells. *Clin Exp Immunol*. 2012; 168: 113-124.
30. Wong VW, Rustad KC, Akaishi S, Sorkin M, Glotzbach JP, Januszyn M, et al. Focal adhesion kinase links mechanical force to skin fibrosis via inflammatory signaling. *Nat Med*. 2011; 18: 148-152.
31. Wong VW, Longaker MT, Gurtner GC. Soft tissue mechanotransduction in wound healing and fibrosis. *Semin Cell Dev Biol*. 2012; 23: 981-986.
32. Chao JT, Davis MJ. The roles of integrins in mediating the effects of mechanical force and growth factors on blood vessels in hypertension. *Curr Hypertens Rep*. 2011; 13: 421-429.
33. Kresh JY, Chopra A. Intercellular and extracellular mechanotransduction in cardiac myocytes. *Pflugers Arch*. 2011; 462: 75-87.
34. Wickström SA, Fässler R. Regulation of membrane traffic by integrin signaling. *Trends Cell Biol*. 2011; 21: 266-273.
35. Ross TD, Coon BG, Yun S, Baeyens N, Tanaka K, Ouyang M, et al. Integrins in mechanotransduction. *Curr Opin Cell Biol*. 2013; 25: 613-618.
36. Kong F, Li Z, Parks WM, Dumbauld DW, García AJ, Mould AP, et al. Cyclic mechanical reinforcement of integrin-ligand interactions. *Mol Cell*. 2013; 49: 1060-1068.
37. Wu D, Schaffler MB, Weinbaum S, Spray DC. Matrix-dependent adhesion mediates network responses to physiological stimulation of the osteocyte cell process. *Proc Natl Acad Sci U S A*. 2013; 110: 12096-12101.
38. Valencik ML, Zhang D, Punske B, Hu P, McDonald JA, Litwin SE. Integrin activation in the heart: a link between electrical and contractile dysfunction? *Circ Res*. 2006; 99: 1403-1410.
39. Storch U, Mederos y Schnitzler M, Gudermann T. G protein-mediated stretch reception. *Am J Physiol Heart Circ Physiol*. 2012; 302: H1241-1249.
40. Haswell ES, Phillips R, Rees DC. Mechanosensitive channels: what can they do and how do they do it? *Structure*. 2011; 19: 1356-1369.
41. Martinac B. Mechanosensitive ion channels: molecules of mechanotransduction. *J Cell Sci*. 2004; 117: 2449-2460.
42. Honoré E, Patel AJ, Chemin J, Suchyna T, Sachs F. Desensitization of mechano-gated K2P channels. *Proc Natl Acad Sci U S A*. 2006; 103: 6859-6864.
43. Lesage F, Lazdunski M. Molecular and functional properties of two-pore-domain potassium channels. *Am J Physiol Renal Physiol*. 2000; 279: F793-801.
44. Carattino MD, Sheng S, Kleyman TR. Epithelial Na<sup>+</sup> channels are activated by laminar shear stress. *J Biol Chem*. 2004; 279: 4120-4126.
45. Satlin LM, Sheng S, Woda CB, Kleyman TR. Epithelial Na<sup>(+)</sup> channels are regulated by flow. *Am J Physiol Renal Physiol*. 2001; 280: F1010-1018.
46. Butterworth MB, Edinger RS, Frizzell RA, Johnson JP. Regulation of the epithelial sodium channel by membrane trafficking. *Am J Physiol Renal Physiol*. 2009; 296: F10-24.
47. Loffing J, Korbmacher C. Regulated sodium transport in the renal connecting tubule (CNT) via the epithelial sodium channel (ENaC). *Pflugers Arch*. 2009; 458: 111-135.
48. Moran MM, McAlexander MA, Bíró T, Szallasi A. Transient receptor potential channels as therapeutic targets. *Nat Rev Drug Discov*. 2011; 10: 601-620.
49. Eijkelkamp N, Quick K, Wood JN. Transient receptor potential channels and mechanosensation. *Annu Rev Neurosci*. 2013; 36: 519-546.
50. Friedrich O, Wagner S, Battle AR, Schürmann S, Martinac B. Mechano-regulation of the beating heart at the cellular level--mechanosensitive channels in normal and diseased heart. *Prog Biophys Mol Biol*. 2012; 110: 226-238.
51. Hsu YJ, Hoenderop JG, Bindels RJ. TRP channels in kidney disease. *Biochim Biophys Acta*. 2007; 1772: 928-936.
52. Woudenberg-Vrenken TE, Bindels RJ, Hoenderop JG. The role of transient receptor potential channels in kidney disease. *Nat Rev Nephrol*. 2009; 5: 441-449.
53. Zou Y, Akazawa H, Qin Y, Sano M, Takano H, Minamino T, et al. Mechanical stress activates angiotensin II type 1 receptor without the involvement of angiotensin II. *Nat Cell Biol*. 2004; 6: 499-506.
54. Toda M, Yamamoto K, Shimizu N, Obi S, Kumagaya S, Igarashi T, et al. Differential gene responses in endothelial cells exposed to a combination of shear stress and cyclic stretch. *J Biotechnol*. 2008; 133: 239-244.
55. Dorn GW 2nd, Brown JH. Gq signaling in cardiac adaptation and maladaptation. *Trends Cardiovasc Med*. 1999; 9: 26-34.
56. Adams JW, Sakata Y, Davis MG, Sah VP, Wang Y, Liggett SB, et al. Enhanced Galphaq signaling: a common pathway mediates cardiac hypertrophy and apoptotic heart failure. *Proc Natl Acad Sci U S A*. 1998; 95: 10140-10145.
57. Mende U, Kagen A, Cohen A, Aramburu J, Schoen FJ, Neer EJ. Transient cardiac expression of constitutively active Galphaq leads to hypertrophy and dilated cardiomyopathy by calcineurin-dependent and independent pathways. *Proc Natl Acad Sci U S A*. 1998; 95: 13893-13898.

58. Mishra S, Ling H, Grimm M, Zhang T, Bers DM, Brown JH. Cardiac hypertrophy and heart failure development through Gq and CaM kinase II signaling. *J Cardiovasc Pharmacol*. 2010; 56: 598-603.
59. Zhang P, Mende U. Regulators of G-protein signaling in the heart and their potential as therapeutic targets. *Circ Res*. 2011; 109: 320-333.
60. Heximer SP, Knutsen RH, Sun X, Kaltenbronn KM, Rhee MH, Peng N, et al. Hypertension and prolonged vasoconstrictor signaling in RGS2-deficient mice. *J Clin Invest*. 2003; 111: 445-52.
61. Calò LA, Pagnin E, Davis PA, Sartori M, Ceolotto G, Pessina AC, et al. Increased expression of regulator of G protein signaling-2 (RGS-2) in Bartter's/Gitelman's syndrome. A role in the control of vascular tone and implication for hypertension. *J Clin Endocrinol Metab*. 2004; 89: 4153-4157.
62. Matozaki T, Nakanishi H, Takai Y. Small G-protein networks: their crosstalk and signal cascades. *Cell Signal*. 2000; 12: 515-524.
63. Tzima E, del Pozo MA, Shattil SJ, Chien S, Schwartz MA. Activation of integrins in endothelial cells by fluid shear stress mediates Rho-dependent cytoskeletal alignment. *EMBO J*. 2001; 20: 4639-4647.
64. Wojciak-Stothard B, Ridley AJ. Shear stress-induced endothelial cell polarization is mediated by Rho and Rac but not Cdc42 or PI 3-kinases. *J Cell Biol*. 2003; 161: 429-439.
65. Shiu YT, Li S, Marganski WA, Usami S, Schwartz MA, Wang YL, et al. Rho mediates the shear-enhancement of endothelial cell migration and traction force generation. *Biophys J*. 2004; 86: 2558-2565.
66. Ziembicki J, Tandon R, Schelling JR, Sedor JR, Miller RT, Huang C. Mechanical force-activated phospholipase D is mediated by Galpha12/13-Rho and calmodulin-dependent kinase in renal epithelial cells. *Am J Physiol Renal Physiol*. 2005; 289: F826-834.
67. Tomar A, Schlaepfer DD. Focal adhesion kinase: switching between GAPs and GEFs in the regulation of cell motility. *Curr Opin Cell Biol*. 2009; 21: 676-683.
68. Peng F, Zhang B, Ingram AJ, Gao B, Zhang Y, Krepinsky JC. Mechanical stretch-induced RhoA activation is mediated by the RhoGEF Vav2 in mesangial cells. *Cell Signal*. 2010; 22: 34-40.
69. Guilluy C, Swaminathan V, Garcia-Mata R, O'Brien ET, Superfine R, Burridge K. The Rho GEFs LARG and GEF-H1 regulate the mechanical response to force on integrins. *Nat Cell Biol*. 2011; 13: 722-727.
70. Yang B, Radcliff C, Hughes D, Kelemen S, Rizzo V. p190 RhoGTPase-activating protein links the  $\beta 1$  integrin/caveolin-1 mechanosignaling complex to RhoA and actin remodeling. *Arterioscler Thromb Vasc Biol*. 2011; 31: 376-383.
71. Hatton JP1, Pooran M, Li CF, Luzzio C, Hughes-Fulford M. A short pulse of mechanical force induces gene expression and growth in MC3T3-E1 osteoblasts via an ERK 1/2 pathway. *J Bone Miner Res*. 2003; 18: 58-66.
72. Yan YX, Gong YW, Guo Y, Lv Q, Guo C, Zhuang Y, et al. Mechanical strain regulates osteoblast proliferation through integrin-mediated ERK activation. *PLoS One*. 2012; 7: e35709.
73. Lien SC, Chang SF, Lee PL, Wei SY, Chang MD, Chang JY, et al. Mechanical regulation of cancer cell apoptosis and autophagy: Roles of bone morphogenetic protein receptor, Smad1/5, and p38 MAPK. *Biochim Biophys Acta*. 2013; 1833: 3124-3133.
74. Jo H, Sipes K, Go YM, Law R, Rong J, McDonald JM. Differential effect of shear stress on extracellular signal-regulated kinase and N-terminal Jun kinase in endothelial cells. Gi2- and Gbeta/gamma-dependent signaling pathways. *J Biol Chem*. 1997; 272: 1395-1401.
75. Scott JE, Yang SY, Stanik E, Anderson JE. Influence of strain on [3H] thymidine incorporation, surfactant-related phospholipid synthesis, and cAMP levels in fetal type II alveolar cells. *Am J Respir Cell Mol Biol*. 1993; 8: 258-65.
76. Russo LA, Rannels SR, Laslow KS, Rannels DE. Stretch-related changes in lung cAMP after partial pneumonectomy. *Am J Physiol*. 1989; 257: E261-268.
77. Cheng J, Du J. Mechanical stretch simulates proliferation of venous smooth muscle cells through activation of the insulin-like growth factor-1 receptor. *Arterioscler Thromb Vasc Biol*. 2007; 27: 1744-51.
78. Peng XQ, Damarla M, Skirball J, Nonas S, Wang XY, Han EJ, et al. Protective role of PI3-kinase/Akt/eNOS signaling in mechanical stress through inhibition of p38 mitogen-activated protein kinase in mouse lung. *Acta Pharmacol Sin*. 2010; 31: 175-183.
79. Mitra S, Sammani S, Wang T, Boone DL, Meyer NJ, Dudek SM, et al. Role of growth arrest and DNA damage-inducible  $\alpha$  in Akt phosphorylation and ubiquitination after mechanical stress-induced vascular injury. *Am J Respir Crit Care Med*. 2011; 184: 1030-1040.
80. Churchill E, Budas G, Vallentin A, Koyanagi T, Mochly-Rosen D. PKC isozymes in chronic cardiac disease: possible therapeutic targets? *Annu Rev Pharmacol Toxicol*. 2008; 48: 569-599.
81. Kulik TJ, Bialecki RA, Colucci WS, Rothman A, Glennon ET, Underwood RH. Stretch increases inositol trisphosphate and inositol tetrakisphosphate in cultured pulmonary vascular smooth muscle cells. *Biochem Biophys Res Commun*. 1991; 180: 982-7.
82. Strait JB 3rd, Martin JL, Bayer A, Mestral R, Eble DM, Samarel AM. Role of protein kinase C-epsilon in hypertrophy of cultured neonatal rat ventricular myocytes. *Am J Physiol Heart Circ Physiol*. 2001; 280: H756-766.
83. Heidkamp MC, Bayer AL, Scully BT, Eble DM, Samarel AM. Activation of focal adhesion kinase by protein kinase C epsilon in neonatal rat ventricular myocytes. *Am J Physiol Heart Circ Physiol*. 2003; 285: H1684-1696.
84. Hubbard SR, Till JH. Protein tyrosine kinase structure and function. *Annu Rev Biochem*. 2000; 69: 373-398.
85. Hubbard SR. Structural analysis of receptor tyrosine kinases. *Prog Biophys Mol Biol*. 1999; 71: 343-358.
86. Haga JH, Li YS, Chien S. Molecular basis of the effects of mechanical stretch on vascular smooth muscle cells. *J Biomech*. 2007; 40: 947-960.
87. Boudreault F, Tschumperlin DJ. Stretch-induced mitogen-activated protein kinase activation in lung fibroblasts is independent of receptor tyrosine kinases. *Am J Respir Cell Mol Biol*. 2010; 43: 64-73.
88. Desai LP, White SR, Waters CM. Mechanical stretch decreases FAK phosphorylation and reduces cell migration through loss of JIP3-induced JNK phosphorylation in airway epithelial cells. *Am J Physiol Lung Cell Mol Physiol*. 2009; 297: L520-529.
89. Huang C, Bruggeman LA, Hydo LM, Miller RT. Shear stress induces cell apoptosis via a c-Src-phospholipase D-mTOR signaling pathway in cultured podocytes. *Exp Cell Res*. 2012; 318: 1075-1085.
90. Davis ME, Cai H, Drummond GR, Harrison DG. Shear stress regulates endothelial nitric oxide synthase expression through c-Src by divergent signaling pathways. *Circ Res*. 2001; 89: 1073-1080.
91. Meliton AY, Muñoz NM, Meliton LN, Birukova AA, Leff AR, Birukov KG. Mechanical induction of group V phospholipase A(2) causes lung inflammation and acute lung injury. *Am J Physiol Lung Cell Mol Physiol*. 2013; 304: L689-700.
92. Liu M, Xu J, Liu J, Kraw ME, Tanswell AK, Post M. Mechanical strain-enhanced fetal lung cell proliferation is mediated by phospholipase C and D and protein kinase C. *Am J Physiol*. 1995; 268: L729-738.



93. Ruwhof C, van Wamel JT, Noordzij LA, Aydin S, Harper JC, van der Laarse A. Mechanical stress stimulates phospholipase C activity and intracellular calcium ion levels in neonatal rat cardiomyocytes. *Cell Calcium*. 2001; 29: 73-83.
94. Hornberger TA, Chu WK, Mak YW, Hsiung JW, Huang SA, Chien S. The role of phospholipase D and phosphatidic acid in the mechanical activation of mTOR signaling in skeletal muscle. *Proc Natl Acad Sci U S A*. 2006; 103: 4741-4746.
95. Bhagyalakshmi A, Berthiaume F, Reich KM, Frangos JA. Fluid shear stress stimulates membrane phospholipid metabolism in cultured human endothelial cells. *J Vasc Res*. 1992; 29: 443-449.
96. Lamb RG, Harper CC, McKinney JS, Rzigalinski BA, Ellis EF. Alterations in phosphatidylcholine metabolism of stretch-injured cultured rat astrocytes. *J Neurochem*. 1997; 68: 1904-1910.
97. Heisenberg CP, Bellaïche Y. Forces in tissue morphogenesis and patterning. *Cell*. 2013; 153: 948-962.
98. Chien S, Li S, Shyy YJ. Effects of mechanical forces on signal transduction and gene expression in endothelial cells. *Hypertension*. 1998; 31: 162-169.
99. Brooks AR, Lelkes PI, Rubanyi GM. Gene expression profiling of vascular endothelial cells exposed to fluid mechanical forces: relevance for focal susceptibility to atherosclerosis. *Endothelium*. 2004; 11: 45-57.
100. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell*. 2004; 116: 281-297.
101. Turczynska KM, Sadegh MK, Hellstrand P, Swärd K, Albinsson S. MicroRNAs are essential for stretch-induced vascular smooth muscle contractile differentiation via microRNA (miR)-145-dependent expression of L-type calcium channels. *J Biol Chem*. 2012; 287: 19199-19206.
102. Huang Y, Crawford M, Higuaita-Castro N, Nana-Sinkam P, Ghadiali SN. miR-146a regulates mechanotransduction and pressure-induced inflammation in small airway epithelium. *FASEB J*. 2012; 26: 3351-3364.
103. Turczyńska KM, Bhattachariya A, Säll J, Göransson O, Swärd K, Hellstrand P, et al. Stretch-Sensitive Down-Regulation of the miR-144/451 Cluster in Vascular Smooth Muscle and Its Role in AMP-Activated Protein Kinase Signaling. *PLoS One*. 2013; 8:e65135.
104. Watts JA, Marchick MR, Kline JA. Right ventricular heart failure from pulmonary embolism: key distinctions from chronic pulmonary hypertension. *J Card Fail*. 2010; 16: 250-259.
105. Greka A, Mundel P. Cell biology and pathology of podocytes. *Annu Rev Physiol*. 2012; 74: 299-323.
106. Petermann AT, Pippin J, Durvasula R, Pichler R, Hiromura K, Monkawa T, et al. Mechanical stretch induces podocyte hypertrophy in vitro. *Kidney Int*. 2005; 67: 157-166.
107. Stieger N, Worthmann K, Schiffer M. The role of metabolic and haemodynamic factors in podocyte injury in diabetes. *Diabetes Metab Res Rev*. 2011; 27: 207-215.
108. Patrakka J, Tryggvason K. New insights into the role of podocytes in proteinuria. *Nat Rev Nephrol*. 2009; 5: 463-468.
109. Wiggins RC. The spectrum of podocytopathies: a unifying view of glomerular diseases. *Kidney Int*. 2007; 71: 1205-1214.
110. Shankland SJ. The podocyte's response to injury: role in proteinuria and glomerulosclerosis. *Kidney Int*. 2006; 69: 2131-2147.
111. Kriz W. Podocyte is the major culprit accounting for the progression of chronic renal disease. *Microsc Res Tech*. 2002; 57: 189-195.
112. Jefferson JA, Shankland SJ, Pichler RH. Proteinuria in diabetic kidney disease: a mechanistic viewpoint. *Kidney Int*. 2008; 74: 22-36.
113. Reddy GR, Kotlyarevska K, Ransom RF, Menon RK. The podocyte and diabetes mellitus: is the podocyte the key to the origins of diabetic nephropathy? *Curr Opin Nephrol Hypertens*. 2008; 17: 32-36.
114. Forbes JM, Fukami K, Cooper ME. Diabetic nephropathy: where hemodynamics meets metabolism. *Exp Clin Endocrinol Diabetes*. 2007; 115: 69-84.
115. Steffes MW, Schmidt D, McCreery R, Basgen JM; International Diabetic Nephropathy Study Group. Glomerular cell number in normal subjects and in type 1 diabetic patients. *Kidney Int*. 2001; 59: 2104-2113.
116. White KE, Bilous RW, Marshall SM, El Nahas M, Remuzzi G, Piras G, et al. Podocyte number in normotensive type 1 diabetic patients with albuminuria. *Diabetes*. 2002; 51: 3083-3089.
117. Pagtalunan ME, Miller PL, Jumping-Eagle S, Nelson RG, Myers BD, Rennke HG, et al. Podocyte loss and progressive glomerular injury in type II diabetes. *J Clin Invest*. 1997; 99: 342-348.
118. Janmey PA, Miller RT. Mechanisms of mechanical signaling in development and disease. *J Cell Sci*. 2011; 124: 9-18.
119. Wyss HM, Henderson JM, Byfield FJ, Bruggeman LA, Ding Y, Huang C, et al. Biophysical properties of normal and diseased renal glomeruli. *Am J Physiol Cell Physiol*. 2011; 300: C397-405.

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