

Short Note

Anticancer Evaluation of S-Glycosylated Quinazolines

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According to the world health organization (WHO), cancer is an important health problem that claims the level of more than 7 million people worldwide on an annual basis [1,2]. Because of the limitation of surgery and radiotherapy in effecting a cure for cancer, chemotherapy has been increasingly important [1,2]. Therefore, identification of novel potent, selective, and less toxic anticancer agents remains one of the most pressing health problems. In the vast cancer chemotherapeutic space, glycosides have played a very important role as established cancer chemotherapeutic agents, either in their nature, semi-synthetically, or synthetically forms [3-62]. As cited above, among the natural glycosides based antitumor the antibiotic doxorubicin, anthracycline O- glycoside, ranks among the most effective anticancer drug for acute myelocytic leukaemia [5-7]. Furthermore, many sugar modified nucleoside analogues are clinically useful chemotherapeutics [3]. For example, capecitabine [14], N-nucleoside and C-nucleoside, are applied in the treatment of metastatic breast cancer and hairy cell leukaemia, respectively. Recently, several S-glycosides, a new non-classical class of nucleosides, have been proved to be potential anticancer agents against many cell Lines [17-22]. Khodair et al., described the synthesis of a series of heterocyclic S-glycosides, thiohydantoin [31-47], rhodanines [48], thioquinazolines [49,50], thiopyridines (51-53), and thiopyrimidine [54], S-glycosides and revealed their potential antitumor activities.

Breast cancer is the most frequent malignancy in females. Due to its major impact on the population, this disease represents a critical public health problem that requires further research at the molecular level to define its prognosis and specific treatment. Basic research is required to accomplish this task and this involves cell lines as they can be widely used in many aspects of laboratory research and, particularly, as *in vitro* models in cancer research. MCF-7 is a commonly used breast cancer cell line, that has been promoted for more than 40 years by multiple research groups but its characteristics have never been gathered in a consistent review article. The current paper provides a broad description of the MCF-7 cell line, including the molecular profile, proliferation, migration, invasion, spheroid formation, its involvement in angiogenesis and lymphangiogenesis and its interaction with the mesenchymal stem cells [63].

Breast cancer is a commonly diagnosed cancer and a leading cause of cancer-related death in women worldwide [64]. It remains an area of active research both clinically and

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experimentally. Recent advances in metabolomics show that metabolic profiling can be useful for the identification of biomarkers in breast cancer. Metabolic profiles of human breast cancer show differences among breast cancer subtypes and offer a way to identify and develop strategies for precise prevention and treatment [65-67]. Obesity is a risk factor for breast cancer; its occurrence is positively associated with the risk of breast cancer [68,69]. Obesity is a modern disorder that has resulted, not just from changes in energy balance, but from changes in lifestyle that alter meal times and eating patterns [70,71]. These changes, as environmental factors, disrupt biological rhythms and contribute to metabolic dysfunction [72,73]. Laboratory studies have shown that the feeding timing modifies obesogenic in rodents. For example, mice fed a high-fat diet (HFD) during the light phase (rest phase for nocturnal animals) gain more weight than mice fed during the dark phase (active phase for nocturnal animals) [74]. Mice fed an HFD during both light and dark phases exhibit altered daily pattern of energy expenditure and gain body fat [75]. Time-restricted feeding (TRF) is an effective tool in obesity research in rodents. It reinforces the circadian rhythms of energy metabolism by temporal regulation of the feeding/fasting pattern to a fixed time during the dark phase of the day. Available studies have shown that TRF restores the diurnal rhythms of energy metabolism [73], and circadian gene expression [76], improves insulin sensitivity, and reduces body adiposity and inflammation in mice fed an HFD [75-77].

Our research interest focused on design and synthesize new small heterocyclic nucleosides targeting cancer especially MCF-7 and HepG2 cell lines. The elaboration of quinazoline derivatives linked with ethoxy ethyl and glycopyranose sugars (Figure 1), to form the target nucleosides was our task [49,50]. The *in vitro* cytotoxic activity against MCF-7 and HepG2 cell lines showed effective anti-proliferative activity of the analyzed derivatives with lower IC₅₀ values especially **9a** with IC₅₀ = 2.09 and 2.08 μM against MCF-7 and HepG2, respectively, and their treatments were safe against the normal cell line Gingival mesenchymal stem cells (GMSC). Moreover, RT-PCR reaction investigated the apoptotic pathway for the compound **9a**, which activated the P53 genes and its related genes. So, further work is recommended for developing it as a chemotherapeutic drug. We found that anticancer activity of the promising derivatives **5**, **8a,b** and **9a,b** was tested against breast (MCF-7), liver (HepG2) cell lines

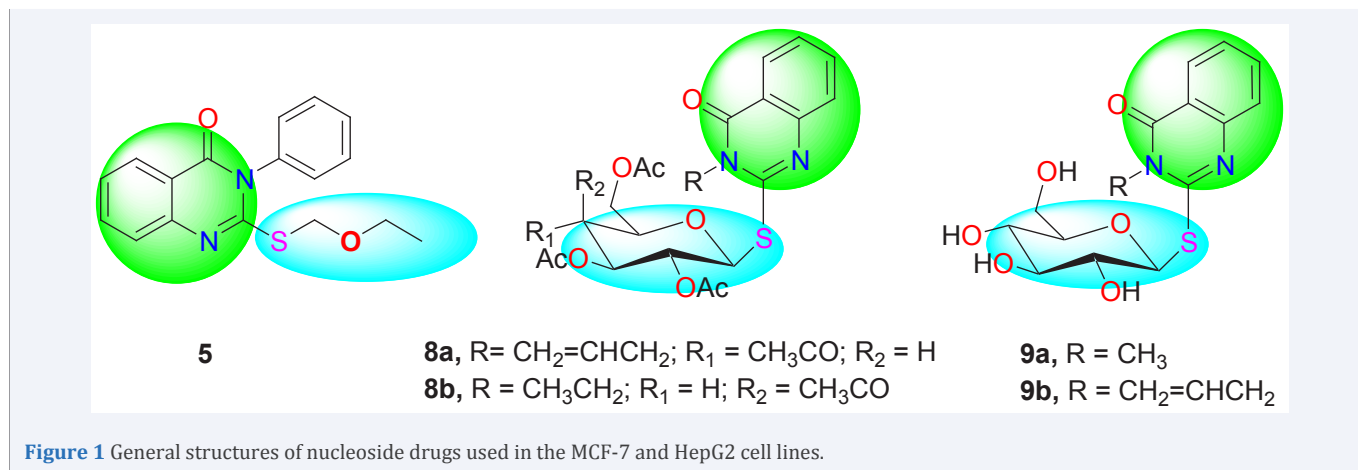


Figure 1 General structures of nucleoside drugs used in the MCF-7 and HepG2 cell lines.

Table 1: Summarized IC₅₀ for the activity of the analyzed compounds against the MCF-7 and HepG2 cell lines.

	IC ₅₀ (μM)		
	MCF-7	HepG2	GMSC
5-FU	4.23	4.43	> 50
5	8.96	1.52	> 50
8a	5.93	3.79	> 50
8b	2.42	1.17	ND
9a	2.09	2.08	> 50
9b	2.04	2.09	> 50

ND= Not Determined

by measuring the percentage of cell survival against their serial dilutions (0.01, 0.1, 1, 10, and 100 μM) [50]. Moreover, they were screened against the GMSC as normal cell line to test their safety [50]. We conclude the incorporation of sugar portion to the nucleus, enhanced the cytotoxic activity against the MCF-7 and HepG2 cell lines by having lower IC₅₀ values, as shown in **Table 1**. Although both compounds **9a** and **9b** have near IC₅₀ values (2.09 and 2.04 μM, respectively) against HepG2 cells, **9a** was considered as the lead compound in our study according to the molecular docking results. It has a higher binding affinity towards the EGFR tyrosine kinase receptor because it forms a larger number of hydrogen bonds with the key amino acid residue **Met 769** compared to other derivatives, so it was selected for further testing as the molecular mode of action. An attempt to study the structure-activity relationship using the molecular docking tool for elucidation the binding interactions of the nucleosides which might justify their higher potency [50]. Glycosides of structurally similar heterocyclic systems have been reported before [31-62].

The nucleoside bases 3-substituted 2-thioxo-2,3-dihydro-1H-quinazolin-4-ones and 3-substituted 2-thioxo-2,3-dihydro-1H-benzo[g]quinazolin-4-ones can be utilized as starting materials for the synthesis of other carbohydrate derivatives as deoxy, amino and azido nucleosides.

REFERENCES

- Gupta SP. Quantitative structure-activity relationship studies on anticancer drugs. *Chemical Review*. 1994; 94: 1507-1551.
- Keri G, Toth I. In "molecular path mechanisms and new trends in drug research", London, New York, Taylor and Francis 1st edition. 2003; 227.
- Kren V, Martinkova L. Glycosides in medicine: The role of glycosidic residue in biological activity. *Curr Med Chem*. 2001; 8: 1303-1328.
- Buchanan JG, Edgar AR, Hutchison RJ, Stobie A, Wightman RH. A new synthesis of formycin via nitropyrazole derivatives. *J Chem Soc Chem Commun*. 1980; 5:237-238.
- Monneret C. Recent developments in the field of antitumour anthracyclines. *Eur J Med Chem*. 2001; 36: 483-493.
- Minotti G, Menna P, Salvatorelli E, Cairo G, Gianni L. Anthracyclines: molecular advances and pharmacologic developments in antitumor activity and cardiotoxicity. *Pharmacol Rev*. 2004; 56: 185-229.
- Krohn KE. Topics in current chemistry: Anthracycline chemistry and biology. 2008; 282.
- Grdadolnik SG, Pristovsek P, Mierke DF. Vancomycin: conformational consequences of the sugar substituent. *J Med Chem*. 1998; 41: 2090-2099.
- Zhang H, Qian DZ, Tai YS, Lee K, Gao P, et al. Digoxin and other cardiac glycosides inhibit HIF-1? synthesis and block tumour growth. *Proc Natl Acad Sci USA*. 2008; 105: 19579.
- Peterson LB, Blagg BS. Click chemistry to probe Hsp90: Synthesis and evaluation of a series of triazole-containing novobiocin analogues. *Bioorg Med Chem Lett*. 2010; 20: 3957-3960.
- Moyer JD, Oliver JT, Handschumacher RE. Salvage of circulating pyrimidine nucleosides in the rat. *Cancer Res*. 1981; 41: 3010-3017.
- Cadman E, Benz C. Uridine and cytidine metabolism following

- inhibition of de novo pyrimidine synthesis by pyrazofurin. *Biochim Biophys Acta*. 1980; 609: 372-382.
13. Saran A. Correlation between the conformation of nucleoside antibiotics and their biological activity. *Int J Quantum Chem*. 1989; 35: 193.
 14. Tiwari KN, Shortnacy-Fowler AT, Parker WB, Waud WR, Secrist JA 3rd. Synthesis and anticancer evaluation of 4'-C-methyl-2'-fluoro arabino nucleosides. *Nucleosides Nucleotides Nucleic Acids*. 2009; 28: 657-677.
 15. Pomeisl K, Votruba I, Holy A, Pohl R. Synthesis of pyrimidine acyclic nucleosides phosphonates as potent inhibitors of thymidine phosphorylase from sd-lymphoma (BD-ECGF). *Nucleosides Nucleotides Nucleic acids*. 2007; 26: 1025.
 16. Elgemeie GH, El-Enany, Ismail MM, Ahmed EK. Nucleic acid components and their analogues: a novel and efficient method for the synthesis of a new class of bipyridyl and biheterocyclic-nitrogen thioglycosides from pyridine2(1H)-thiones. *Nucleosides Nucleotide Nucleic Acids*. 2002; 21: 477.
 17. Rashad AE, Mahmoud AE, Ali MM. Synthesis and anti-cancer effects of some novel pyrazolo[3,4-d] pyrimidine derivatives by generating reactive oxygen species in human breast adenocarcinoma cells. *Eur J Med Chem*. 2011; 46: 1019.
 18. Saad HA, Moustafa AH. Synthesis and anticancer activity of some new S-glycosyl and S-alkyl,2,4-triazinone derivatives. *Molecules*. 2011; 16: 5682-5700.
 19. Al-Mutairi MS, Al-Abdullah ES, Haiba ME, Khedr MA, Zaghary WA. Synthesis and molecular docking and preliminary in vitro cytotoxic evaluation of some substituted tetrahydroxy naphthalene (2',3',4',6'-tetra-O-acetyl-D-glucosyl and/or galactopyranosyl) derivatives. *Molecules*. 2012; 17: 4717.
 20. Scala S, Akhmed N, Rao US, Paul K, Lan L, et al. P-glycoprotein substrates and antagonists cluster into two distinct groups. *Molecular Pharmacol*. 1997; 51: 1024.
 21. Abu-Zaied MZ, Nawwar GA, Swellem RH, El-Sayed SH. Synthesis and screening of new 5- substituted,3,4-oxadiazole-2-thioglycosides as potent anticancer. *Pharmacol pharmacy*. 2012; 3: 254.
 22. Akihino I, Yuichi M, Yukishige. Synergistic solvent effect in, 1,2-*cis*-glycosides formation. *Tetrahedron*. 2008; 64: 92.
 23. Larsen JS, Zahran MA, Pedersen EB, Nielsen C. Synthesis of triazenopyrazole derivatives as potential inhibitors of HIV-1. *Monatsch Chem*. 1999; 130: 1167.
 24. Storer R, Ashton CJ, Baxter AD, Hann MM, Marr CL, et al. The synthesis and antiviral activity of 4-fluoro-1-beta-D-ribofuranosyl-1H-pyrazole3-carboxamide. *Nucleosides Nucleotides Nucleic acids*. 1999; 18: 203.
 25. Manfredini S, Baraldi PG, Bazzanini R, Durini E, Vertuani S, et al. Pyrazole related nucleosides 5. Synthesis and biological activity of 2'-deoxy2',3'-dideoxy- and acyclo-analogues of 4-iodo-1-beta-D-ribofuranosyl-3carboxymethyl pyrazole (IPCAR). *Nucleosides Nucleotides Nucleic acids*. 2000; 19: 705.
 26. Hafez HN, El-Gazzar AR, Nawwar GA. Synthesis, biological and medicinal significance of S-glycosido-thieno[2,3-d]-pyrimidines as new anti-inflammatory and analgesic agents. *Eur J Med Chem*. 2010; 45: 1485-1493.
 27. Schimdtt RR. New Methods of synthesis of glycosides and oligosaccharides. *Angew Chem Int Ed Engl*. 1986; 25: 212.
 28. Elgemeie GH, Zaghary WA, Amin KM, Nasr TM. First synthesis of thiophene thioglycosides. *J Carbohydr Chem*. 2009; 28: 161.
 29. Cristescu C, Czobor F. As-triazine derivatives with potential therapeutic action. XXVI. Syntheses of 5-substituted-6-azauracil acyclonucleosides. *Nucleosides Nucleotides*. 1998; 17: 1319-1324.
 30. Abu-Zaied MA, El-Telbani EM, Elgemeie GH, Nawwar GA. Synthesis and in vitro anti-tumour activity of new oxadiazole thioglycosides. *Eur J Med Chem*. 2011; 46: 229-235.
 31. Khodair AI, Gesson JP. a new approach for the N-galactosylation and S-galactosylation of 5-arylidene-2-thioxo-4-thiazolidinediones. *Carbohydr Res*. 2011; 364: 2831-2837.
 32. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. S-glycosylated hydantoins as new antiviral agents. *J Med Chem*. 1994; 37: 73-77.
 33. Khodair AI, EL-Subagh HI, El-Emam AA. Synthesis of certain 5-substituted 2-thiohydantoin derivatives as potential cytotoxic and antiviral agents. *Boll. Chim. Farm.* 1997; 136: 561-567.
 34. Al-Obaid AM, EL-Subagh HI, Khodair AI, Elmazar MMA. 5-substituted-2-thiohydantoin analogues as a novel class of antitumor agents, *Anti-Cancer Drugs*. 1996; 7: 873-880.
 35. Khodair AI. Glycosylation of 2-thiohydantoin derivatives, synthesis of some novel S-alkylated and S-glycosylated hydantoins, *Carbohydr. Res*. 2001; 331: 445-453.
 36. Khodair AI. Synthesis of 2-thiohydantoins and their S-glycosylated derivatives as potential antiviral and antitumor agents, *Nucleosides, Nucleotide & Nucleic Acids*. 2001; 20: 1735-1750.
 37. Khodair AI, Ibrahim EE. Synthesis of hydantoin nucleosides with naphthyl methylene substituents in the 5-position, *Nucleosides and Nucleotides*. 1996; 15: 1927-1943.
 38. Khodair AI, El-Subbagh HI, El-Emam AA. Synthesis of certain 5-substituted 2-thiohydantoin derivatives as potential cytotoxic and antiviral agents. *Boll Chim Farm*. 1997; 136: 561-567.
 39. Khodair AI. A convenient synthesis of glycosylated hydantoins as potential antiviral agents, *Phosphorus, Sulfur and Silicon & the Related Elements*. 1997; 122: 9-26.
 40. Khodair AI. Synthesis and evaluation of the activity of arylidene hydrazono- and glycopyranosyl hydrazine sulfonyl benzylidene-2,4-imidazolidindiones, *Carbohydrate Research*. 1998; 306: 567-573.
 41. Khodair AI, Jean-Pierre Gesson. Sulfur glycosylation reactions involving 3-allyl-2-thiohydantoin nucleoside bases as potential antiviral and antitumor agents, *Phosphorus, Sulfur and Silicon & the Related Elements*. 1998; 142: 167-190
 42. Khodair AI. Glycosylation of 2-thiohydantoin derivatives. synthesis of some novel S-alkylated and S-glycosylated hydantoins, *Carbohydrate Res*. 2001; 331: 445-453.
 43. Khodair AI. Synthesis of 2-thiohydantoins and their S-glycosylated derivatives as potential antiviral and antitumor agents, *Nucleosides, Nucleotides & Nucleic Acids*. 2001; 20: 1735-1750.
 44. Khodair AI, El-Barbary AA, Abbas YA, Imam DR. Synthesis, reactions and conformational analysis of 5-arylidene-2-thiohydantoins as potential antiviral agents, *Phosphorus, Sulfur and Silicon & the Related Elements*. 2001; 170: 261-278.
 45. Al-Masoudi IA, Khodair AI, Al-Soud YA, Al-Masoudi NA. Synthesis of N-substituted 1-amino-2,3-dihydro-1H-imidazole-2-thione-N-nucleosides and S-glycosylated derivatives, *Nucleosides, Nucleotides & Nucleic Acids*. 2003; 3: 299-307.
 46. Al-Masoudi NA, Al-Soud YA, Khodair AI. Some 2'-modified 4'-thionucleosides via sulfur participation and synthesis of thio-AZT from 4'-thiofuranoid 1,2-glycol, *Phosphorus, Sulfur and Silicon & the Related Elements*. 2003; 178: 1199-1209.

47. Khodair AI, El-Ashry EH, Al-Masoudi NAL. Thiohydantoin nucleosides synthesis approaches, *Monch. für Chemi.* 2004; 135: 1061-1079.
48. Khodair AI, Awad MK, Gesson J-P, Elshaier YAMM. New N-ribosides and N-mannosides of rhodanine derivatives with anticancer activity on leukaemia cell line: Design, synthesis, DFT and molecular modelling studies, *Carbohydrate Research.* 2020; 487: 107894.
49. Khodair AI, Elsafi MA, Al-Essa SA. Al-Essa. Simple and efficient synthesis of novel 3-substituted-2-thioxo-2,3-dihydro-1H-benzo[g]quinazolin-4-ones and their reactions with alkyl halides and α -glycopyranosyl bromides. *J Heterocyclic Chem.* 2019; 56: 2358-2368.
50. Khodair AI, Alsafi MA, Nafie MS. Synthesis, molecular modelling and anti-cancer evaluation of a series of quinazoline derivatives, *Carbohydrate Res.* 2019; 486: 107832.
51. Khodair AI, Al-Masoudi NA, Gesson J-P. A modified synthesis of benzothiazole, benzoxazole and pyridine nucleosides and evaluation of antitumor activity, *Nucleosides, Nucleotides & Nucleic Acids.* 2003; 22: 2061-2076.
52. Khodair AI, Attia AM, Gendy EA, Elshaier YAMM, El-Magd MA. Design, synthesis and cytotoxicity evaluation of some novel of S-glycoside of 2-thioxopyridine and N-glycoside of 2-oxopyridine derivatives as anti-breast cancer. *J Heterocyclic Chem.* 2019; 56: 1733-1747.
53. Attia AM, Khodair AI, Gendy EA, El-Magd MA, Elshaier YAMM. New 2-oxopyridine/2-thiopyridine derivatives tethered to a benzotriazole with cytotoxicity on MCF7 cell lines and with antiviral activities. *Letters in Drug Design & Discovery.* 2019; 16: 1-14.
54. Khodair AI, Ibrahim EE, El-Ashry EH. Glycosylation of 2-thiouracil derivatives. a synthetic approach to 3-glycosyl-2,4-dioxypyrimidines, *Nucleosides & Nucleotides.* 1997; 16: 433-444.
55. Khodair AI. Convenient synthesis of 2-arylidene-5H-thiazolo[2,3-b]quinazoline-3,5[2H]-diones and their benzoquinazoline derivatives. *J Heterocycl Chem.* 2002; 39: 1153-1160.
56. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Synthesis of uridine with 5-methylene-2-thiohydantoin substituent in the 5-position. *Liebigs Ann Chem.* 1994; 619-621.
57. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Synthesis and evaluation of activity of 2-deoxyuridines with 5-methylene-2-thiohydantoin substituents in the 5-position. *Monatsh Chem.* 1994; 125: 593-598.
58. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Synthesis of uridine with 5-methylene-2-thiohydantoin substituents in the 5-position. *Liebigs Ann Chem.* 1994; 619-621.
59. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Synthesis of 3-amino and 5-amino hydantoin 2-deoxynucleosides, *Nucleosides & Nucleotides.* 1994; 13: 707-717.
60. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Synthesis and antiviral evaluation of hydantoin analogues of AZT, *Arch. Pharm. (Weinheim).* 1994; 327: 633-655.
61. El-Barbary AA, Khodair AI, Pedersen EB, Nielsen C. Convergent synthesis of 2,3-dideoxy-3-mercapto nucleosides potential anti-HIV agents. *Monatsh. Chem.* 1994; 125: 1017-1025.
62. El-Bary HA, El-Barbary AA, Khodair AI, Abdel Magied AE, Pedersen EB, Nielsen C. Synthesis of hydantoin analogues of 3'-fluoro-3'-deoxythymidine (ft). *Bull Soc Chim Fr.* 1995; 132: 149-155.
63. Comşa S, Cimpean AM, Raica M. The story of MCF-7 breast cancer cell line: 40 years of experience in research. *Anticancer Res.* 2015; 35: 3147-3154.
64. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics. *Globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries.* *CA Cancer J Clin.* 2018; 68: 394-424.
65. Giskeodegard GF, Grinde MT, Sitter B, Axelson DE, Lundgren S, Fjosne HF, et al. Multivariate modelling and prediction of breast cancer prognostic factors using MR metabolomics. *J Proteome Res.* 2010; 9: 972-979.
66. Sitter B, Bathen TF, Singstad TE, Fjosne HE, Lundgren S, Halgunset J, et al. Quantification of metabolites in breast cancer patients with different clinical prognosis using HR-MAS MR spectroscopy. *NMR Biomed.* 2010; 23: 424-431.
67. Tang X, Lin CC, Spasojevic I, Iversen ES, Chi JT, Marks JR. A joint analysis of metabolomics and genetics of breast cancer. *Breast Cancer Res.* 2014; 16: 415.
68. Picon-Ruiz M, Morata-Tarifa C, Valle-Goffin JJ, Friedman ER, Slingerland JM. Obesity and adverse breast cancer risk and outcome: Mechanistic insights and strategies for intervention. *CA Cancer J Clin.* 2017; 67: 378-397.
69. Rohan TE, Heo M, Choi L, Datta M, Freudenheim JL, Kamensky V, et al. Body fat and breast cancer risk in postmenopausal women: a longitudinal study. *J Cancer.* 2013; 754815.
70. Bae SA, Fang MZ, Rustgi V, Zarbl H, Androulakis IP. At the interface of lifestyle, behaviour, and circadian rhythms: metabolic implications. *Front Nutr.* 2019; 6: 132.
71. Maury E, Brichard SM. Adipokine dysregulation. *Adipose tissue inflammation and metabolic syndrome.* *Mol Cel Endocrinol.* 2010; 314: 1-16.
72. Branecky KL, Niswender KD, Pendergast JS. Disruption of daily rhythms by high-fat diet is reversible, *PLOS One.* 2015; 10: 137970.
73. Zarrinpar A, Chaix A, Panda S. Daily eating patterns and their impact on health and disease, *Trends Endocrinol Metab.* 2016; 27: 69-83.
74. Arble DM, Bass J, Laposky AD, Vitaterna MH, Turek FW. Circadian timing of food intake contributes to weight gain. *Obesity.* 2009; 17: 2100-2102.
75. Sundaram S, Yan L. Time-restricted feeding reduces adiposity in mice fed a high-fat diet. *Nutr Res.* 2016; 36: 603-611.
76. Hatori M, Vollmers C, Zarrinpar A, Dittacchio L, Bushong EA, Gill S, et al. Time-restricted feeding without reducing caloric intake prevents metabolic diseases in mice fed a high-fat diet. *Cell Metab.* 2012; 15: 848-860.
77. Chaix A, Lin T, Le HD, Chang MW, Panda S. Time-restricted feeding prevents obesity and metabolic syndrome in mice lacking a circadian clock. *Cell Metab.* 2019; 29: 303-319.

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