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Perspective

The Bioartificial Pancreas — How should We Address the Issue of Oxygen Delivery?

Emmanuel C Opara* and Benjamin S Harrison

Wake Forest Institute for Regenerative Medicine and Virginia Tech-Wake Forest School of Biomedical Engineering and Sciences (SBES), Wake Forest School of Medicine, USA

INTRODUCTION

Diabetes mellitus represents a growing burden both on health-care expenditures and the quality of life of the afflicted individuals. Current estimates for the prevalence of diabetes indicate a global prevalence of about 285 million people [1]. Type 1 diabetes is a significant cause of morbidity and mortality in young adults. Secondary diabetic complications include a quadrupled risk of heart attack and stroke and a significant decrease in life expectancy. The economic impact of diabetes is tremendous across the world, with a projected impact of over \$200 billion in direct annual costs in North America in 2010 and an estimated 25% of U.S Medicare annual in-patient care expenditures being attributed to the treatment of diabetes and its associated complications [2].

The current standard treatment for Type 1 diabetes is daily injections of exogenous insulin to control blood sugar. An alternative treatment modality for Type 1 diabetes is the replacement of the missing β -cells through transplantation of whole pancreas, which in contrast to insulin administration, is capable of achieving normoglycemia along with the prevention and even reversal of certain secondary diabetic complications, such as nephropathy and atherosclerosis [3]. The advantageous effects of β -cell replacement therapy on diabetic complications compared to insulin treatment may be attributed to the role played by the byproduct of pro-insulin cleavage, named C-peptide, during insulin processing in the β -cell [4-7]. However, the benefits of the β -cell replacement may be masked by collateral risks associated with the use of immunosuppressive drugs to prevent transplant rejection in transplant recipients [3].

While whole pancreas transplantations have been performed, it is a complex surgical procedure that is fraught with significant morbidities and challenging technical issues including the drainage of exocrine secretions from the transplanted pancreas [3]. Successful islet transplantation in diabetic patients remained elusive [8] until the introduction of the glucocorticoid-free immunosuppressive regimen by the Edmonton group, and this protocol has successfully led to insulin independence in a limited number of diabetic patients transplanted with isolated human islets [9,10]. In a few cases, insulin independence has been achieved for several years [11,12], thus showing islet

*Corresponding author

Dr. Emmanuel C Opara, Wake Forest Institute for Regenerative Medicine, Wake Forest School of Medicine, Medical Center Blvd., Winston-Salem, NC 27157, USA, Tel: 336-713-1297; Fax: 336-713-7290; Email: eopara@wakehealth.edu

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transplantation to be a viable therapeutic option for patients with Type1 diabetes. Still, the need to use immunosuppressive drugs to prevent graft rejection and the severe shortage of human islets remain two major barriers to clinical islet transplantation [12-16].

An attractive strategy to overcome these two obstacles to routine use of islet transplantation is the technique of immunoisolation by microencapsulation of islets prior to transplantation, as it could potentially solve the problem of islet shortage by opening up the possibility of using islets from non-human sources while obviating the need for longterm immunosuppression of transplant recipients [13,16-24]. However, there are a number of issues that need to be resolved before microencapsulated islet transplantation can become a clinical reality. A major obstacle of this approach is the death of large proportions of the encapsulated islet grafts owing to severe hypoxia, resulting in the need for large quantities of islets to achieve normoglycemia in experimental diabetic animals.

Why is Oxygen So Important?

Although islets constitute approximately 1% of the pancreas, they receive about 6-10% of the blood flow to this gland [25,26], indicating a disproportionate level of perfusion in which islets receive and consume oxygen. The unusually high oxygen requirement of islets is interrupted during the process of islet isolation and processing for transplantation, and studies have shown that hypoxia has significant deleterious effects on the survival and function of islets [27-29]. In the immediate post-transplant period, isolated islet transplants are forced to depend upon diffusion of oxygen and nutrients through peripheral perfusion from the surrounding tissue within the site of transplantation [30], until the islet transplants are revascularized by angiogenesis, a process that requires 7 - 10 days [28]. As a result most studies with encapsulated islets have used extraordinarily high doses of these cells to achieve variable effects on blood glucose levels in large animals and human subjects [20,31-35]. Using more cells and hoping enough survive is an inefficient and counterproductive approach because more cells mean more oxygen demand for a limited supply. This may inadvertantly select for islets with the lowest metabolism in the struggle for surival possibly leading to a less effective therapy.

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To achieve the goal of maintaining normal blood glucose levels through islet transplantation, a key factor for producing high quality of islets is the prevention of oxidative stress during islet preparation [36,37] and a sufficient oxygen supply during the immediate post-transplant period [30]. How do we get oxygen to islets and how do we overcome the challenges associated with its delivery?

Significant cell death can occur during the process of isolating islets. One should consider the oxygen tensions during the islet isolation process. Culture and transport of islets could be performed in gas-permeable devices such as silicone rubbers or hollow fibers bioreactors [38]. A factor to consider in using these processes would be the need to avoid aggregation or accumulation of islets to ensure optimal oxygenation of the cells.

Once implantation occurs, oxygen delivery becomes an even more significant requirement during the time window between implantation and establishment of the support vascularity. In general, while there are mechanical means to deliver oxygen (e.g. perfusion pumps with oxygen carrier solutions), one must also consider that after implantation having to perform any second procedures to remove an oxygen-delivery system would be highly undesirable. There is therefore a crucial need for a more elegant solution for oxygen delivery in the immediate posttransplantation period. One approach could involve incorporation of oxygen delivery systems into the microencapsulation process that would be exhausted in due time without adverse events in the body.

Having the ability to co-encapsulate islets with a source of oxygen rich materials may fulfill this goal. Investigators have microencapsulated islets in barium-alginate with perfluorocarbon (PFC) emulsion. After low oxygen culture for 2 days, islets in control alginate capsules without PFC lost substantial viable tissue and displayed necrotic cores, whereas most of the original oxygen consumption rate was recovered with the oxygen-supplying PFC in the microcapsules [39]. The PFC can carry, through adsorption, oxygen at much greater concerntrations than water. However, other investigators have noted that reformulation of the PFC emulsion is required to reduce toxicity to the islets, and it has also been shown that PFC emulsions may have little or no benefit to encapsulated β -cells in culture [40].

An alternative to perfluorocarbons that involves taking advantage of chemical reactions which allows for greater densities of oxygen to be stored and delivered subsequently may work. Co-encapsulation of islets with micro- or nano- particulate oxygen generators that can chemically generate oxygen to help bridge to revascularization seems to be a particularly attractive option [41]. Solid peroxides, such as sodium percarbonate or calcium peroxide, can potentially deliver 100 times the amount of oxygen that can be stored in an equivalent amount of water. Of course, too much of a good thing can be detrimental and so the oxygen delivery systems need to be designed to provide oxygen at a therapeutic dosage. Yet such an approach should be practical as our own bodies have developed strategies to keep oxygen and reactive oxygen species under control.

SUMMARY AND CONCLUSION

With the impact of diabetes mellitus continuing to grow, there is an urgent need for creating effective strategies to treat this disease. For those patients who could most benefit from islet transplantation, developing approaches which enhance islet cell survival during the periods of isolation, encapsulation, and implantation, as well as during the period of integration to systemic circulation is critical. Optimal oxygen delivery during these processes is critically important. From the brief review above there are several potential approaches which may provide the needed boost to enhance the delivery of oxygenin the development of the bioartificial pancreas. Further development and evaluation of the efficacy of these approaches are needed for successful use of the bioartificial pancreas in diabetic animals and humans.

REFERENCES

- 1. Hu FB. Globalization of diabetes: the role of diet, lifestyle, and genes. Diabetes Care. 2011; 34: 1249-1257.
- Opara EC, Mirmalek-Sani SH, Khanna O, Moya ML, Brey EM. Design of a bioartificial pancreas(+). J Investig Med. 2010; 58: 831-837.
- 3. White SA, Shaw JA, Sutherland DE. Pancreas transplantation. Lancet. 2009; 373: 1808-1817.
- Ido Y, Vindigni A, Chang K, Stramm L, Chance R, Heath WF, et al. Prevention of vascular and neural dysfunction in diabetic rats by C-peptide. Science. 1997; 277: 563-566.
- 5. Hansen A, Johansson BL, Wahren J, von Bibra H. C-peptide exerts beneficial effects on myocardial blood flow and function in patients with type 1 diabetes. Diabetes. 2002; 51: 3077-3082.
- Ekberg K, Brismar T, Johansson BL, Jonsson B, Lindström P, Wahren J. Amelioration of sensory nerve dysfunction by C-Peptide in patients with type 1 diabetes. Diabetes. 2003; 52: 536-541.
- Wahren J, Sima AA. C-Peptide is Relevant in Type 1 Diabetes and its Complications: Summary and Conclusions to the Special Issue. Rev Diabet Stud. 2009; 6: 223-224.
- 8. Alejandro R, Lehmann R, Ricordi C, Kenyon NS, Angelico MC, Burke G, et al. Long-term function (6 years) of islet allografts in type 1 diabetes. Diabetes. 1997; 46: 1983-1989.
- Shapiro AM, Lakey JR, Ryan EA, Korbutt GS, Toth E, Warnock GL, et al. Islet transplantation in seven patients with type1 diabetes mellitus using a glucocorticoid-free immunosuppressive regimen. N Engl J Med. 2000; 343: 230-238.
- 10. Ryan EA, Lakey JR, Rajotte RV, Korbutt GS, Kin T, Imes S, et al. Clinical outcomes and insulin secretion after islet transplantation with the Edmonton protocol. Diabetes. 2001; 50: 710-719.
- Rother KI, Harlan DM. Challenges facing islet transplantation for the treatment of type 1 diabetes mellitus. J Clin Invest. 2004; 114: 877-883.
- 12. Ryan EA, Paty BW, Senior PA, Bigam D, Alfadhli E, Kneteman NM, et al. Five-year follow-up after clinical islet transplantation. Diabetes. 2005; 54: 2060-2069.
- 13.Weir GC, Bonner-Weir S. Scientific and political impediments to successful islet transplantation. Diabetes. 1997; 46: 1247-1256.
- 14. Robertson RP. Successful islet transplantation for patients with diabetes--fact or fantasy? N Engl J Med. 2000; 343: 289-290.
- 15. Hogan A, Pileggi A, Ricordi C. Transplantation: current developments

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and future directions; the future of clinical islet transplantation as a cure for diabetes. Front Biosci. 2008; 13: 1192-1205.

- 16. Kizilel S, Garfinkel M, Opara E. The bioartificial pancreas: progress and challenges. Diabetes Technol Ther. 2005; 7: 968-985.
- 17.de Vos P, Marchetti P. Encapsulation of pancreatic islets for transplantation in diabetes: the untouchable islets. Trends Mol Med. 2002; 8: 363-366.
- Schrezenmeir J, Kirchgessner J, Gerö L, Kunz LA, Beyer J, Mueller-Klieser W. Effect of microencapsulation on oxygen distribution in islets organs. Transplantation. 1994; 57: 1308-1314.
- 19. De Vos P, Van Straaten JF, Nieuwenhuizen AG, de Groot M, Ploeg RJ, De Haan BJ, et al. Why do microencapsulated islet grafts fail in the absence of fibrotic overgrowth? Diabetes. 1999; 48: 1381-1388.
- 20.Sun Y, Ma X, Zhou D, Vacek I, Sun AM. Normalization of diabetes in spontaneously diabetic cynomologus monkeys by xenografts of microencapsulated porcine islets without immunosuppression. J Clin Invest. 1996; 98: 1417-1422.
- 21.Garfinkel MR, Harland RC, Opara EC. Optimization of the microencapsulated islet for transplantation. J Surg Res. 1998; 76: 7-10.
- 22. de Groot M, Schuurs TA, van Schilfgaarde R. Causes of limited survival of microencapsulated pancreatic islet grafts. J Surg Res. 2004; 121: 141-150.
- 23.Kendall WF Jr, Collins BH, Opara EC. Islet cell transplantation for the treatment of diabetes mellitus. Expert Opin Biol Ther. 2001; 1: 109-119.
- 24.Hill RS, Cruise GM, Hager SR, Lamberti FV, Yu X, Garufis C, et al. Immunoisolation of adult porcine islets for the treatment of diabetes mellitus. Use of photopolymerizable polyethylene glycol in the conformal coating of mass-isolated porcine islets. Ann NY Acad Sci. 1997; 831: 332- 343.
- 25.Lifson N, Lassa CV, Dixit PK. Relation between blood flow and morphology in islet organ of rat pancreas. Am J Physiol. 1985; 249: E43-48.
- 26. Jansson L, Hellerström C. Stimulation by glucose of the blood flow to the pancreatic islets of the rat. Diabetologia. 1983; 25: 45-50.
- 27.Dionne KE, Colton CK, Yarmush ML. Effect of hypoxia on insulin secretion by isolated rat and canine islets of Langerhans. Diabetes. 1993; 42: 12-21.
- 28. Menger MD, Jaeger S, Walter P, Feifel G, Hammersen F, Messmer K. Angiogenesis and hemodynamics of microvasculature of transplanted islets of Langerhans. Diabetes. 1989; 38 Suppl 1: 199-201.
- 29. Mendoza V, Klein D, Ichii H, Ribeiro MM, Ricordi C, Hankeln T, et al. Protection of islets in culture by delivery of oxygen binding neuroglobin via protein transduction. Transplant Proc. 2005; 37: 237-240.

- 30. Davalli AM, Scaglia L, Zangen DH, Hollister J, Bonner-Weir S, Weir GC. Vulnerability of islets in the immediate posttransplantation period. Dynamic changes in structure and function. Diabetes. 1996; 45: 1161-1167.
- 31.Soon-Shiong P, Heintz RE, Merideth N, Yao QX, Yao Z, Zheng T, et al. Insulin independence in a type 1 diabetic patient after encapsulated islet transplantation. Lancet. 1994; 343: 950-951.
- 32.Dufrane D, Goebbels RM, Saliez A, Guiot Y, Gianello P. Six-month survival of microencapsulated pig islets and alginate biocompatibility in primates: proof of concept. Transplantation. 2006; 81: 1345-1353.
- 33.Calafiore R, Basta G, Luca G, Lemmi A, Montanucci MP, Calabrese G, et al. Microencapsulated pancreatic islet allografts into nonimmunosuppressed patients with type 1 diabetes: first two cases. Diabetes Care. 2006; 29: 137-138.
- 34. Elliott RB, Escobar L, Tan PL, Muzina M, Zwain S, Buchanan C. Live encapsulated porcine islets from a type 1 diabetic patient 9.5 yr after xenotransplantation. Xenotransplantation. 2007; 14: 157-161.
- 35.Wang T, Adcock J, Kühtreiber W, Qiang D, Salleng KJ, Trenary I, et al. Successful allotransplantation of encapsulated islets in pancreatectomized canines for diabetic management without the use of immunosuppression. Transplantation. 2008; 85: 331-337.
- 36.Wang T, Adcock J, Kühtreiber W, Qiang D, Salleng KJ, Trenary I, et al. Successful allotransplantation of encapsulated islets in pancreatectomized canines for diabetic management without the use of immunosuppression. Transplantation. 2008; 85: 331-337.
- 37.Bottino R, Balamurugan AN, Bertera S, Pietropaolo M, Trucco M, Piganelli JD. Preservation of human islet cell functional mass by antioxidative action of a novel SOD mimic compound. Diabetes. 2002; 51: 2561-2567.
- 38.Bottino R, Balamurugan AN, Tse H, Thirunavukkarasu C, Ge X, Profozich J, et al. Response of human islets to isolation stress and the effect of antioxidant treatment. Diabetes. 2004; 53: 2559-2568.
- 39. Papas KK, Avgoustiniatos ES, Tempelman LA, Weir GC, Colton CK, Pisania A, et al. High-density culture of human islets on top of silicone rubber membranes. Transplant Proc. 2005; 37: 3412-3414.
- 40. Johnson AS, O'Sullivan E, D'Aoust LN, Omer A, Bonner-Weir S, Fisher RJ, et al. Quantitative assessment of islets of Langerhans encapsulated in alginate. Tissue Eng Part C Methods. 2011; 17: 435-449.
- 41.Goh F, Gross JD, Simpson NE, Sambanis A. Limited beneficial effects of perfluorocarbon emulsions on encapsulated cells in culture: experimental and modeling studies. J Biotechnol. 2010; 150: 232-239.
- 42. Ward CL, Corona BT, Yoo JJ, Harrison BS, Christ GJ. Oxygen Generating Biomaterials Preserve Skeletal Muscle Homeostasis under Hypoxic and Ischemic Conditions. PLoS One. 2013; 8: e72485.

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