

Special Issue on von Hippel Lindau Disease

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Review Article

Nervous System Manifestations of von Hippel-Lindau Disease

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Abstract

von Hippel-Lindau (VHL) disease is a highly penetrant multiple organ heritable cancer syndrome with a spectrum of benign and malignant tumors. VHL results from a germline mutation of the *VHL tumor suppressor gene* (short arm of chromosome 3). VHL affects 1 in 36,000 to 39,000 livebirths as an autosomal dominant trait. Individuals harboring a mutation in *VHL* can develop benign and malignant tumors, both of the central nervous system (CNS) and systemically. VHL-associated lesions of the CNS include retinal and craniospinal hemangioblastomas, as well as endolymphatic sac tumors (ELSTs). While CNS hemangioblastomas (most common tumor in VHL) and ELSTs are benign tumors, they are associated with significant neurologic morbidity and mortality based on their location and multiplicity. Because of the management complexities of this disease, multidisciplinary screening and treatment, as well as a deep understanding of the natural course of the disease are needed.

INTRODUCTION

von Hippel-Lindau disease (VHL) (OMIM 193300) is an autosomal dominant heritable neoplastic syndrome typified by the frequent development of benign and malignant, vascular tumors in highly conserved topographic distribution [1]. Affected individuals can develop central nervous system (CNS) lesions including retinal and craniospinal hemangioblastomas, as well as endolymphatic sac tumors (ELSTs) of the temporal bone. Visceral VHL-associated lesions frequently include renal cell carcinomas, renal cysts, pheochromocytomas, extra-adrenal paragangliomas, pancreatic microcystic adenomas, pancreatic cysts, pancreatic neuroendocrine tumors, as well as cystadenomas of the epididymis and broad ligament (Figure 1) [2-12].

The disease eponym is derived from the German ophthalmologist, Eugen von Hippel, who described the retinal angioma ("angiomatosis retinae", which are currently referred to as retinal hemangioblastomas) [13] and the Swedish pathologist, Arvid Lindau, who associated hemangioblastomas of the retina and cerebellum with other VHL-associated visceral lesions [14]. VHL has an estimated incidence of 1 in 36,000 to 39,000 live births [15,16] and a penetrance of 90% by 65 years of age [17]. VHL is caused by an underlying germline mutation of *VHL tumor suppressor gene*. Tumor development occurs after inactivation of the wild-type allele [2,18-20]. Judicious multidisciplinary treatment of VHL-associated lesions has increased median life expectancy to 52.5 years [21].

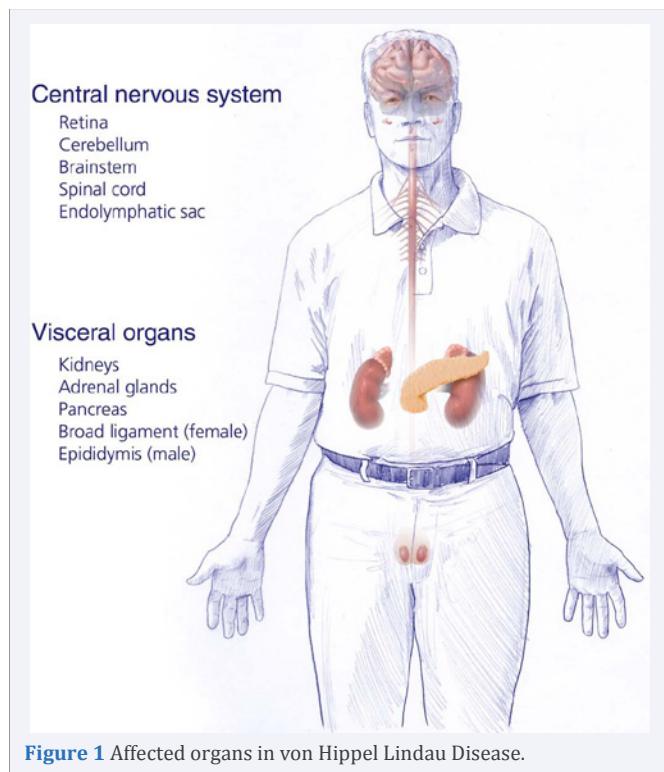


Figure 1 Affected organs in von Hippel Lindau Disease.

MOLECULAR GENETICS

The *VHL* gene is on the short arm of chromosome 3 (3p25-26) [22]. Germline mutations of *VHL* account for more than 95% of the patients affected by VHL [23,24]. Individuals inherit a *VHL* germline mutation from the carrier parent and a normal (wild type) gene from the non-affected parent [25]. Tumorigenesis

occurs when the wild type *VHL* allele is inactivated (loss of heterozygosity) in defined susceptible target organs (Figure 1). Fifty percent of patients have somatic inactivation of the *VHL* gene in sporadically occurring hemangioblastomas [26-28] and renal cell carcinomas [29-32].

VHL is composed of 3 exons that encode for the 213 amino-acid protein (pVHL), which has a molecular weight of 30 kDa [22]. pVHL subcellular shuttling between the cytoplasm and nucleus may be due to posttranslational modification necessary for its oxygen sensing properties [33]. pVHL contains 2 functional domains, α and β . The α domain binds to elongin C recruiting elongin B and Cullin 2 (CUL2) to form the VCB-Cul2, an E3 ubiquitin-ligase complex, that targets substrates for proteasomal degradation (Figure 2) [34-37]. The substrate-recognition site for targeting proteins β domain [34-37].

VHL is critical for proteasomal degradation of hypoxia inducible transcription factor (HIF). HIF is basic helix-loop helix transcription factor that transcriptionally activates genes that participate in homeostatic responses to oxygen changes and, thereby, controls angiogenesis/vascularization, cell proliferation, apoptosis, cell differentiation and glucose metabolism. HIF1 is a heterodimer protein consisting of HIF1 α and HIF1 β . Under states of normoxia HIF1 α is hydroxylated by prolyl hydroxylase 2 (PHD2), facilitating the binding of VCB-CUL2 complex tagging HIF α for ubiquitination and targeted degradation.

Under hypoxic conditions, the HIF1 α -subunit is stabilized, accumulates and dimerizes with HIF1 β . The dimer translocates to the nucleus and co-activates transcription of genes that contain hypoxia responsive elements (HREs). Currently, over than 200 genes have been identified to be target genes of HIF that contain the HRE sequence, including platelet derivative

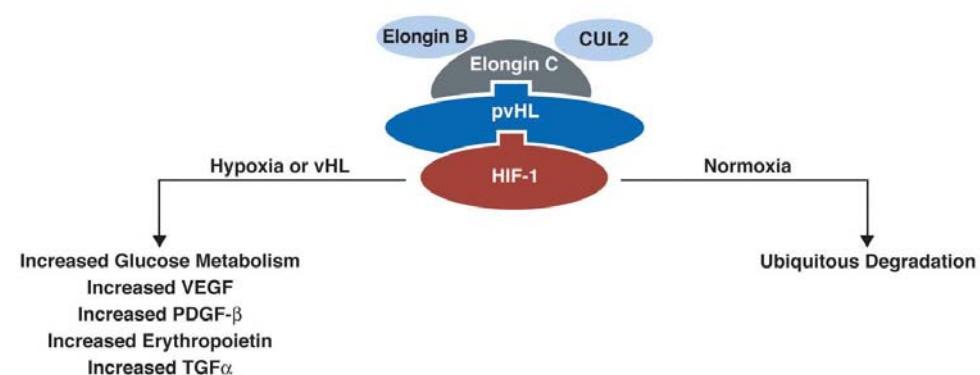


Figure 2 The VCB-CUL2 complex and the functions of the complex under normoxia and hypoxia. (Adapted from Lonser et al., Lancet 2003) [1].

Table 1: VHL subtypes based on family linkage analysis and genotype-phenotype classifications.

	Clinical characteristics
Type 1	Retinal hemangioblastomas, CNS hemangioblastomas, Renal cell carcinoma, Pancreatic neoplasms and cysts
Type 2A	Pheochromocytomas, Retinal hemangioblastomas, CNS hemangioblastomas
Type 2B	Pheochromocytomas, Retinal hemangioblastomas, CNS hemangioblastomas, Renal cell carcinomas, Pancreatic neoplasms and cysts
Type 2C	Pheochromocytoma only

growth factor β (PDGF β), transforming growth factor α (TGF α), vascular endothelial growth factor (VEGF), erythropoietin (EPO) and glucose transporter-1 (Glut-1) [38-41]. If VHL protein is absent or abnormal, HIF initiates transcription of genes favoring tumor development including those listed above, as well as those involved in extracellular matrix formation, vasculogenesis, chemotaxis, pH regulation and cell cycle regulation [20,42].

DIAGNOSIS

VHL can be diagnosed based on clinical criteria and/or genetic testing. In individuals with a family history of VHL, a crani spinal hemangioblastoma, retinal hemangioblastoma, renal clear cell carcinoma or pheochromocytoma are diagnostic. In individuals with no VHL family history, the diagnosis of VHL is made if they have 2 or more CNS hemangioblastomas, or 1 CNS hemangioblastoma and a visceral tumor (with the exception of renal and epididymal cysts) [5,43]. Genetic (germline) testing in VHL families using Southern blotting and DNA-sequence analysis is 100% [44]. However, in a study of 181 VHL kindreds, it was determined that postzygotic mosaicism was the mechanism in 2 (4.8%) of 42 parents. Consequently, mosaicism can be an infrequent cause for failure of a VHL molecular diagnosis [45].

Specific correlations have emerged between genotype and phenotype by linkage analysis, which has aided in providing screening and counseling of affected family members, and those with similar genetic aberrations (Table 1) [19,20,46-50].

CRANIOSPINAL HEMANGIOBLASTOMAS

General

Hemangioblastomas are highly vascular CNS tumors. They are a defining feature in VHL. They affect 80% or more of VHL patients and it is the most common tumor (CNS or visceral) associated with VHL [1,17, 43,51]. Mean age at symptom formation due to VHL-associated CNS hemangioblastomas is approximately 30 years and hemangioblastoma(s) are the presenting feature in half of VHL patients [7,52,53]. Although these tumors are benign, they are a leading cause of VHL-associated mortality and neurologic morbidity.

Natural history

Large scale VHL patient studies have shown that hemangioblastomas have a sporadic growth pattern with periods of growth followed by growth arrest ("saltatory growth pattern") [1,6-8,54]. Patterns of growth vary and are categorized as saltatory (72% of growing tumors), linear (6%), or exponential (22%). Many tumors will remain the same size for several years [7]. In recent studies [54], VHL patients were found to have a mean of 8.5 tumors/patient (range, 1 to 33 tumors/patients) at initial evaluation. Mean tumor development was 0.4 new tumors/year and was correlated with age (more frequent development in younger patients).

Hemangioblastomas are associated with edema and peritumoral cysts (approximately 10% of all CNS hemangioblastomas). Depending on location, the peritumoral cyst mass effect often has a greater capacity to cause signs/

Table 2: Symptoms based on anatomic location of tumors.*

Location	Frequency	Percentage
Brainstem		
Headache	10	83.3%
Singultus	8	66.6%
Nausea/vomiting	6	50.0%
Dysphagia	5	41.7%
Cough	3	25.0%
Paresthesia	3	25.0%
Cauda equina		
Urinary/bowel abnormalities	5	100.0%
Pain	5	100.0%
Paresthesia	4	80.0%
Headache	2	40.0%
Cerebellum		
Headache	77	77.0%
Gait ataxia	57	57.0%
Nausea/vomiting	19	19.0%
Vertigo	18	18.0%
Speech difficulties	15	15.0%
Dysmetria	11	11.0%
Nerve root		
Cruciate paralysis	1	100.0%
Spine		
Paresthesia	28	75.7%
Pain	24	64.9%
Gait ataxia	13	35.1%
Dysesthesias	9	24.3%
Urinary/bowel abnormalities	7	18.9%
Supratentorial		
Vision disturbance/loss	2	50.0%
Weakness	2	50.0%

*. The symptoms listed here are the major symptoms recorded from patients that underwent resection of symptomatic tumors.

symptoms than the tumor itself (because of more rapid growth of the cyst compared to the tumor itself). Edema formation is the result of increased hemangioblastoma vascular permeability associated, in turn with increased interstitial pressure in the tumor. This results in plasma distribution by bulk flow into the adjacent tissue [55]. Once the capacity of the surrounding tissue to absorb the interstitial fluid is exceeded, cyst formation occurs. The walls of peritumoral cysts (when associated with hemangioblastomas) are lined by chronic astrocytosis with abundant Rosenthal fibers.

Clinical features

Signs and symptoms vary based on the location of the hemangioblastoma, edema and/or cyst (tumor volume, edema and/or cyst size) (Table 2). On average, tumors that become symptomatic and require resection grow faster than tumors that

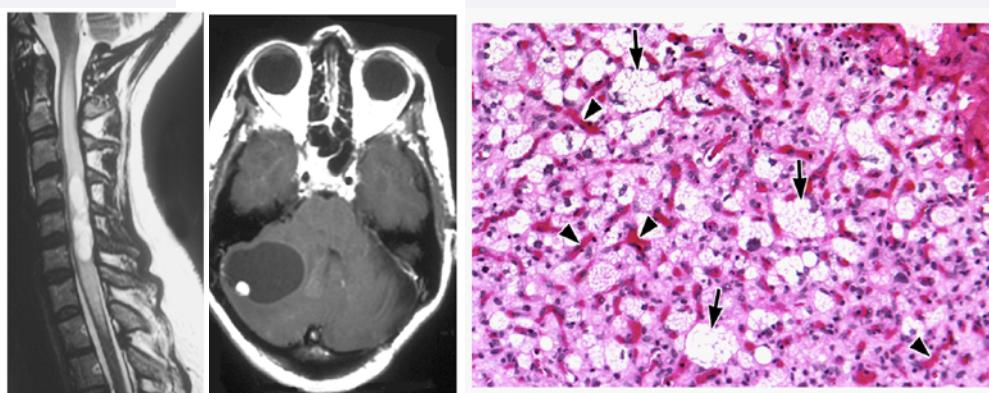


Figure 3 Diagnostic radiologic and histologic images of von Hippel-Lindau disease (VHL)-associated hemangioblastomas of the craniospinal axis. (Left panel) Sagittal T2-weighted magnetic resonance (MR)-imaging of an intramedullary homogenously enhancing hemangioblastoma with associated spinal cord edema. (Middle panel) Axial T1-weighted MR-image of a right cerebellar hemisphere hemangioblastoma associated with a peritumoral cyst. (Right panel) Hematoxylin and eosin staining demonstrating classic vascular elements (arrowheads) along with stromal vacuolated cells (arrows). Original magnification X 20. (Adapted from Lonser et al. Ann Neurol, 2005) [55].

are asymptomatic. Similarly, associated peritumoral cysts that become symptomatic and require resection grow much faster (3.2-fold faster versus asymptomatic lesions at 1.7-fold) than the tumors they are associated with [7].

Radiologic findings

Hemangioblastomas are best visualized by contrast-enhanced T1-weighted MR-imaging. T2-weighted MR-imaging allows excellent quantification of edema and peritumoral cysts. Arteriography can be used to highlight the arteriovenous shunting and early draining veins associated with these tumors prior to resection (Figure 3).

Origin and histology

Hemangioblastomas are characterized by "stromal" lipid laden neoplastic cells within an abundant irregular reticular capillary network [56]. Similar to the embryologic hemangioblast, hemangioblastoma stromal cells express brachyury, Scl (stem cell leukemia) and Flk-1 (VEGF-R 2) [57]. Scl expression is developmentally regulated in the observed distribution of CNS hemangioblastomas in VHL patients. These findings indicate that CNS hemangioblastomas may originate from embryologic arrested-hemangioblasts. Grossly, the tumor nodule is orange-red in color (Figure 3).

Treatment

Using well-defined microsurgical techniques, the vast majority of hemangioblastomas in the CNS can be excised safely [6,8]. Most VHL patients will develop numerous hemangioblastomas growing at different rates at several locations. Because of the irregular growth pattern (including saltatory growth) and the inability to accurately predict which tumors will become symptomatic, surgical resection should be reserved until the first onset of signs/symptoms that correlate to the location of the hemangioblastoma. This surgical management paradigm avoids unnecessary surgery and can be used to maintain neurologic function in most patients [6-8].

Arteriography is sometimes used as a vascular map to

better define feeding vessels before the resection of large CNS hemangioblastomas. Some surgeons will use preoperative arteriography and embolization before surgical removal of hemangioblastomas in an effort to control and reduce the tumors vascular supply [6,8]. However, we and others have found that embolization of hemangioblastomas has limited benefit and does not justify the corresponding risks associated with it [8,58,59]. Careful complete microsurgical resection can be performed with minimal blood loss without embolization [1,6,8,60].

Radiation therapy has been used for treatment of CNS hemangioblastomas in VHL. A recent study by Asthagiri and colleagues [61] demonstrated that stereotactic radiosurgery had an acceptable risk for adverse radiation effects, but had diminishing tumor control over long-term follow-up. Consequently, stereotactic radiosurgery typically used for the treating hemangioblastomas that are not surgically resectable. The use of infratentorial craniospinal radiation therapy (ICSR) has also been investigated in VHL patients (7 patients with 84 hemangioblastomas). With long term follow-up (mean, 73.8 months), complete radiographic resolution was achieved in 18% of lesions. Over the study duration, 4 surgeries were required for treatment of symptomatic lesions after ICSR [62].

Pharmacological treatments have been used for management of CNS hemangioblastomas. Due to the highly vascular nature of hemangioblastomas, VEGF has been a major chemotherapy target, as well as other anti-angiogenic agents. Observations of Interferon α (IFN α) in inhibiting angiogenesis have made it an intriguing target for hemangioblastoma treatment. A small scale study demonstrated that treatment with INF α did not prevent growth of visceral cysts and hemangioblastomas. Semaxanib, is an inhibitor of the Flk-1/KDR receptor tyrosine kinase (VEGFR-receptor), and vatalanib, an inhibitor of VEGF-receptor tyrosine kinase, have shown promise but these agents require additional study to determine effectiveness [63]. Recent data suggest that hemangioblastomas that have a missense germline mutation may be treated with proteasome inhibitors [64].

RETINA HEMANGIOBLASTOMA

General

Retinal hemangioblastomas can occur in an estimated 60% of VHL patients and are the first symptomatic manifestation in 33% of patients. Mean age for diagnosis of retinal hemangioblastomas is 25 years, but retinal hemangioblastomas can occur as early as infancy. In fact, 5% of retinal hemangioblastomas can be found in VHL patients less than 10 years of age [1,9,10].

Natural history

Retinal hemangioblastomas are often multiple and bilateral (50%). They vary in size from less than one optic disc to several optic discs in diameter. They may arise in the peripheral retina or the juxtapapillary retina. Fifteen percent of hemangioblastomas are located at the optic disc [9,65]. Despite exhibiting slow and/or saltatory growth, they are capable of causing significant visual morbidity. Dollfus and colleagues found that the average number of hemangioblastomas observed per gene carrier with an ocular manifestation was 1.4 at initial evaluation (range, 1 to 20 hemangioblastomas) and nearly 3 (range, 1 to 20 hemangioblastomas) at final evaluation. Similar to craniospinal hemangioblastomas in VHL patients, the mean number of new hemangioblastomas per patient (of the group with ocular manifestation) per year was 0.4 [9].

Clinical features

Retinal hemangioblastomas are often asymptomatic in the initial stages. Nevertheless, they can ultimately lead to partial or total loss of vision. Symptoms correlate with the hemangioblastoma's location on the retina/optic disc. Generally, symptoms are caused by tumor exudate or tractional effects.

Ophthalmologic findings

Indirect fundoscopy and fluorescein angiography are used for diagnosis. Macular function associated with peripheral and optic nerve lesions is assessed by fluorescein angiography. Grossly, retinal hemangioblastomas are round, circumscribed, orange-red vascular associated with a feeding artery leading from the optic disc and a draining vein.

Histology and origin

Similar to other CNS hemangioblastomas, retinal hemangioblastomas are composed of vacuolated foamy cells that contain large intracytoplasmic lipid inclusions and fenestrated channels [65].

Treatment

Early diagnosis and treatment is crucial to prevent retinal detachment, hemorrhage, glaucoma, and cataracts. These complications can result in blindness in up to 25% of the affected eyes of VHL patients [9]. Treatments include laser photocoagulation, cryotherapy, photodynamic therapy, radiation or surgical excision. Location of the tumor affects treatment efficacy and applicability. Treatments for peripheral retinal hemangioblastomas cannot always safely be used for juxtapapillary hemangioblastomas. Many peripheral retinal tumors are effectively treated by laser cryotherapy or photocoagulation [66]. In cases of severe lesions with a large fibrovascular component, vitroretinal surgery has been used effectively to improve or prolong visual function [67]. Tumors located on the optic disc may be monitored, as some treatments can cause visual damage. Enucleation may be needed in cases of irreversible glaucoma (with severe pain) with end-stage ocular angiogenesis. Finally, intravitreal injections of anti-VEGF therapeutics have been reported to reduce retinal thickening and retinal hard exudates associated with retinal hemangioblastomas that are not treatable by conventional methods (Table 3) [68].

ELSTS

General

ELSTs are vascular low-grade papillary adenocarcinomas that were identified as part of the VHL spectrum in 1997 [69]. They affect up to 11% of the individuals with VHL. Mean age of diagnosis of 22 years (range, 12 to 50 years). Bilateral ELSTs (30%) can be found in VHL [69]. ELST frequency in VHL is likely higher because many VHL patients may have an infraradiologic ELST [70].

Natural history

In one study, 31 VHL patients with ELSTs (15 males, 16 females) underwent surgical resection of their ELST at a mean age of 38.2 ± 10.2 years (range, 12-67 years). The majority of the patients (29 patients, 31 ears, 94% of ears) had associated audiovestibular symptoms, including sensorineural hearing loss (84% of ears), tinnitus (73%), and vertigo (68%) that did not correlate with tumor size [71]. Based on data from these patients and prior studies, 3 distinct mechanisms were found to underlie audivestibular findings associated with ELSTs [72]. The mechanisms include intralabyrinthine hemorrhage, endolymphatic hydrops and direct invasion of the otic capsule by tumor [73-75].

Table 3: Recommended screening and intervals of test for at-risk individuals.

Test	Start age (frequency)
Ophthalmoscopy	Infancy (yearly)
Plasma or 24 h urinary catecholamines and metanephrines	2 years of age (yearly and when blood pressure is raised)
MRI of crani spinal axis	11 years of age (yearly)
CT and MRI of internal auditory canals	Onset of symptoms (hearing loss, tinnitus, vertigo, or unexplained, difficulties of balance)
Ultrasound of abdomen	8 years of age (yearly; MRI as clinically indicated)
CT of abdomen	18 years of age or earlier if clinically indicated (yearly)
Audiological function tests	When clinically indicated



Figure 4 Imaging and histological findings from a left-sided endolymphatic sac tumor (ELST) within the vestibular aqueduct. (Left panel) Axial, T1-weighted, enhanced magnetic resonance (MR)-imaging of an ELST within the proximal vestibular aqueduct. (Middle panel) Corresponding axial, unenhanced CT imaging demonstrated tumor-associae erosion in the vestibular aqueduct. (Right panel) Hematoxylin and eosin staining demonstrates a papillary-cystic ELST. Original magnificationX20. (Adapted from Lonser et. al. J Neurosurg, 2008) [75].

Clinical features

Because ELST patients most frequently present with a triad of findings, consisting of hearing loss, tinnitus, and vertigo, they can be misdiagnosed with Meniere's disease. Aural fullness, aural pain and facial nerve weakness are less common signs and symptoms [69]. Sudden hearing loss (43%) has been correlated with intralabyrinthine hemorrhage. Gradual hearing loss (47%), however, may be related to endolymphatic hydrops [72,74].

Radiologic findings

MR-imaging of larger ELSTs can show a homogeneous or variable pattern of patchy enhancement post-contrast. Recently efforts have been made to focus on symptomatic ELSTs below the detectable size of that seen on traditional CT or MR-imaging. Nonenhanced T1-weighted MRI has been used to reliably detect ELST-associated intralabyrinthine hemorrhage within the vestibule, cochlea, or semicircular canals distinct from the site of tumor; this may be detected prior to the tumor mass being identified on imaging. Contrast-enhanced delayed FLAIR MR-imaging has been shown to detect ELST-associated hydrops (Figure 4) [76]. Imaging studies should be supplemented with audiograms to document the extent of hearing loss in order to detect any lesions that may not be detectable by radiographical studies.

Histology and origin

ELSTs arise within the vestibular aqueduct portion of the endolymphatic duct/sac system [75]. Histologically, ELSTs are highly vascular, bright or dark red soft tissue masses that may often preferentially erode the immediately adjacent temporal bone. They often have proteinaceous papillary cystic regions [69]. ELST are immunoreactive to anti-NSE, anti-MAK6, and anti-AE1/AE3, as well as EMA and S100 in a subset of cases [Figure 4].

Treatment

Regular screening of VHL patients for ELSTs is recommended, with surgical intervention in selected patients before morbidity develops. Surgery is curative for completely excised tumors. Kim and colleagues found that hearing was stabilized postoperatively in 90% of patients after ELST resection [71]. Current indications for ELST resection in VHL include imaging evidence of an ELST with serviceable hearing (and/or audiovestibular signs/symptoms), evidence of ELST-associated intralabyrinthine

hemorrhage, ELST-associated hydrops or mass effect by the ELST [74]. The role of adjuvant therapy, including chemotherapy, fractionated radiotherapy or gamma knife radiosurgery is not established.

REFERENCES

1. Lonser RR, Glenn GM, Walther M, Chew EY, Libutti SK, Linehan WM, et al. von Hippel-Lindau disease. Lancet. 2003; 361: 2059-2067.
2. Maher ER, Kaelin WG Jr. von Hippel-Lindau disease. Medicine (Baltimore). 1997; 76: 381-391.
3. Maddock IR, Moran A, Maher ER, Teare MD, Norman A, Payne SJ, et al. A genetic register for von Hippel-Lindau disease. J Med Genet. 1996; 33: 120-127.
4. Richard S, Campello C, Taillandier L, Parker F, Resche F. Haemangioblastoma of the central nervous system in von Hippel-Lindau disease. French VHL Study Group. J Intern Med. 1998; 243: 547-553.
5. Lamiell JM, Salazar FG, Hsia YE. von Hippel-Lindau disease affecting 43 members of a single kindred. Medicine (Baltimore). 1989; 68: 1-29.
6. Lonser RR, Weil RJ, Wanebo JE, DeVroom HL, Oldfield EH. Surgical management of spinal cord hemangioblastomas in patients with von Hippel-Lindau disease. J Neurosurg. 2003; 98: 106-116.
7. Wanebo JE, Lonser RR, Glenn GM, Oldfield EH. The natural history of hemangioblastomas of the central nervous system in patients with von Hippel-Lindau disease. J Neurosurg. 2003; 98: 82-94.
8. Weil RJ, Lonser RR, DeVroom HL, Wanebo JE, Oldfield EH. Surgical management of brainstem hemangioblastomas in patients with von Hippel-Lindau disease. J Neurosurg. 2003; 98: 95-105.
9. Dollfus H, Massin P, Taupin P, Nemeth C, Amara S, Giraud S, et al. Retinal hemangioblastoma in von Hippel-Lindau disease: a clinical and molecular study. Invest Ophthalmol Vis Sci. 2002; 43: 3067-3074.
10. Webster AR, Maher ER, Moore AT. Clinical characteristics of ocular angiomyomatosis in von Hippel-Lindau disease and correlation with germline mutation. Arch Ophthalmol. 1999; 117: 371-378.
11. Hammel PR, Vilgrain V, Terris B, Penifornis A, Sauvanet A, Correas JM, et al. Pancreatic involvement in von Hippel-Lindau disease. The Groupe Francophone d'Etude de la Maladie de von Hippel-Lindau. Gastroenterology. 2000; 119: 1087-1095.
12. Friedrich CA. Von Hippel-Lindau syndrome. A pleomorphic condition. Cancer. 1999; 86: 2478-2482.
13. Hippel EV. Die anatomische Grund lage der von mir beschriebenen 'sehr seltenen Erkrankung der Netzhaut'. Graefes Arch Ophthalmol. 1911; 79: 350-77.

14. Lindau A. Studien über Kleinhirnssystem: Bau, Pathogenese und Beziehungen zur Angiomatosis Retinae. *Acta Pathol Microbiol Scand.* 1926; Suppl I, 1.
15. Maher ER, Iselius L, Yates JR, Littler M, Benjamin C, Harris R, et al. Von Hippel-Lindau disease: a genetic study. *J Med Genet.* 1991; 28: 443-447.
16. Neumann HP, Wiestler OD. Clustering of features of von Hippel-Lindau syndrome: evidence for a complex genetic locus. *Lancet.* 1991; 337: 1052-1054.
17. Maher ER, Yates JR, Harries R, Benjamin C, Harris R, Moore AT, et al. Clinical features and natural history of von Hippel-Lindau disease. *Q J Med.* 1990; 77: 1151-1163.
18. Linehan WM, Lerman MI, Zbar B. Identification of the von Hippel-Lindau (VHL) gene. Its role in renal cancer. *JAMA.* 1995; 273: 564-570.
19. Clifford SC, Maher ER. Von Hippel-Lindau disease: clinical and molecular perspectives. *Adv Cancer Res.* 2001; 82: 85-105.
20. Kaelin WG Jr. Molecular basis of the VHL hereditary cancer syndrome. *Nat Rev Cancer.* 2002; 2: 673-682.
21. Wilding A, Ingham SL, Laloo F, Clancy T, Huson SM, Moran A, et al. Life expectancy in hereditary cancer predisposing diseases: an observational study. *J Med Genet.* 2012; 49: 264-269.
22. Latif F, Tory K, Gnarra J, Yao M, Duh FM, Orcutt ML, et al. Identification of the von Hippel-Lindau disease tumor suppressor gene. *Science.* 1993; 260: 1317-1320.
23. Maher ER, Neumann HP, Richard S. von Hippel-Lindau disease: a clinical and scientific review. *Eur J Hum Genet.* 2011; 19: 617-623.
24. Nordstrom-O'Brien M, van der Luijt RB, van Rooijen E, van den Ouwehand AM, Majoor-Krakauer DF, Lolkema MP, et al. Genetic analysis of von Hippel-Lindau disease. *Hum Mutat.* 2010; 31: 521-537.
25. Knudson AG Jr, Strong LC. Mutation and cancer: neuroblastoma and pheochromocytoma. *Am J Hum Genet.* 1972; 24: 514-532.
26. Kanno H, Kondo K, Ito S, Yamamoto I, Fujii S, Torigoe S, et al. Somatic mutations of the von Hippel-Lindau tumor suppressor gene in sporadic central nervous system hemangioblastomas. *Cancer Res.* 1994; 54: 4845-4847.
27. Lee JY, Dong SM, Park WS, Yoo NJ, Kim CS, Jang JJ, et al. Loss of heterozygosity and somatic mutations of the VHL tumor suppressor gene in sporadic cerebellar hemangioblastomas. *Cancer Res.* 1998; 58: 504-508.
28. Tse JY, Wong JH, Lo KW, Poon WS, Huang DP, Ng HK. Molecular genetic analysis of the von Hippel-Lindau disease tumor suppressor gene in familial and sporadic cerebellar hemangioblastomas. *Am J Clin Pathol.* 1997; 107: 459-466.
29. Gnarra JR, Tory K, Weng Y, Schmidt L, Wei MH, Li H, et al. Mutations of the VHL tumour suppressor gene in renal carcinoma. *Nat Genet.* 1994; 7: 85-90.
30. Brieger J, Weidt EJ, Schirmacher P, Störkel S, Huber C, Decker HJ. Inverse regulation of vascular endothelial growth factor and VHL tumor suppressor gene in sporadic renal cell carcinomas is correlated with vascular growth: an in vivo study on 29 tumors. *J Mol Med (Berl).* 1999; 77: 505-510.
31. Gallou C, Joly D, Méjean A, Staroz F, Martin N, Tarlet G, et al. Mutations of the VHL gene in sporadic renal cell carcinoma: definition of a risk factor for VHL patients to develop an RCC. *Hum Mutat.* 1999; 13: 464-475.
32. Kondo K, Yao M, Yoshida M, Kishida T, Shuin T, Miura T, et al. Comprehensive mutational analysis of the VHL gene in sporadic renal cell carcinoma: relationship to clinicopathological parameters. *Genes Chromosomes Cancer.* 2002; 34: 58-68.
33. Cai Q, Robertson ES. Ubiquitin/SUMO modification regulates VHL protein stability and nucleocytoplasmic localization. *PLoS One.* 2010; 5.
34. Duan DR, Pause A, Burgess WH, Aso T, Chen DY, Garrett KP, et al. Inhibition of transcription elongation by the VHL tumor suppressor protein. *Science.* 1995; 269: 1402-1406.
35. Pause A, Lee S, Worrell RA, Chen DY, Burgess WH, Linehan WM, et al. The von Hippel-Lindau tumor-suppressor gene product forms a stable complex with human CUL-2, a member of the Cdc53 family of proteins. *Proc Natl Acad Sci U S A.* 1997; 94: 2156-2161.
36. Maxwell PH, Wiesener MS, Chang GW, Clifford SC, Vaux EC, Cockman ME, et al. The tumour suppressor protein VHL targets hypoxia-inducible factors for oxygen-dependent proteolysis. *Nature.* 1999; 399: 271-275.
37. Stebbins CE, Kaelin WG Jr, Pavletich NP. Structure of the VHL-ElonginC-ElonginB complex: implications for VHL tumor suppressor function. *Science.* 1999; 284: 455-461.
38. Ohh M, Park CW, Ivan M, Hoffman MA, Kim TY, Huang LE, et al. Ubiquitination of hypoxia-inducible factor requires direct binding to the beta-domain of the von Hippel-Lindau protein. *Nat Cell Biol.* 2000; 2: 423-427.
39. Ivan M, Kondo K, Yang H, Kim W, Valiando J, Ohh M, et al. HIFalpha targeted for VHL-mediated destruction by proline hydroxylation: implications for O2 sensing. *Science.* 2001; 292: 464-468.
40. Jaakkola P, Mole DR, Tian YM, Wilson MI, Gielbert J, Gaskell SJ, et al. Targeting of HIF-alpha to the von Hippel-Lindau ubiquitylation complex by O2-regulated prolyl hydroxylation. *Science.* 2001; 292: 468-472.
41. Carmeliet P, Dor Y, Herbert JM, Fukumura D, Brusselmans K, Dewerchin M, et al. Role of HIF-1alpha in hypoxia-mediated apoptosis, cell proliferation and tumour angiogenesis. *Nature.* 1998; 394: 485-490.
42. Kuhnert F, Tam BY, Sennino B, Gray JT, Yuan J, Jocson A, et al. Soluble receptor-mediated selective inhibition of VEGFR and PDGFRbeta signaling during physiologic and tumor angiogenesis. *Proc Natl Acad Sci U S A.* 2008; 105: 10185-10190.
43. Melmon Kl, Rosen Sw. Lindau's Disease. Review Of The Literature And Study Of A Large Kindred. *Am J Med.* 1964; 36: 595-617.
44. Stolle C, Glenn G, Zbar B, Humphrey JS, Choyke P, Walther M, et al. Improved detection of germline mutations in the von Hippel-Lindau disease tumor suppressor gene. *Hum Mutat.* 1998; 12: 417-423.
45. Sgambati MT, Stolle C, Choyke PL, Walther MM, Zbar B, Linehan WM, et al. Mosaicism in von Hippel-Lindau disease: lessons from kindreds with germline mutations identified in offspring with mosaic parents. *Am J Hum Genet.* 2000; 66: 84-91.
46. Zbar B, Kishida T, Chen F, Schmidt L, Maher ER, Richards FM, et al. Germline mutations in the Von Hippel-Lindau disease (VHL) gene in families from North America, Europe, and Japan. *Hum Mutat.* 1996; 8: 348-357.
47. Chen F, Kishida T, Yao M, Hustad T, Glavac D, Dean M, et al. Germline mutations in the von Hippel-Lindau disease tumor suppressor gene: correlations with phenotype. *Hum Mutat.* 1995; 5: 66-75.
48. Brauch H, Kishida T, Glavac D, Chen F, Pausch F, Höfler H, et al. Von Hippel-Lindau (VHL) disease with pheochromocytoma in the Black Forest region of Germany: evidence for a founder effect. *Hum Genet.* 1995; 95: 551-556.

49. Cybulski C, Krzystolik K, Murgia A, Górska B, Debniak T, Jakubowska A, et al. Germline mutations in the von Hippel-Lindau (VHL) gene in patients from Poland: disease presentation in patients with deletions of the entire VHL gene. *J Med Genet.* 2002; 39: E38.
50. Hes F, Zewald R, Peeters T, Sijmons R, Links T, Verheij J, et al. Genotype-phenotype correlations in families with deletions in the von Hippel-Lindau (VHL) gene. *Hum Genet.* 2000; 106: 425-431.
51. Choyke PL, Glenn GM, Walther MM, Patronas NJ, Linehan WM, Zbar B. von Hippel-Lindau disease: genetic, clinical, and imaging features. *Radiology.* 1995; 194: 629-642.
52. Neumann HP, Eggert HR, Scheremet R, Schumacher M, Mohadjer M, Wakhloo AK, et al. Central nervous system lesions in von Hippel-Lindau syndrome. *J Neurol Neurosurg Psychiatry.* 1992; 55: 898-901.
53. Richard S, David P, Marsot-Dupuch K, Giraud S, Béroud C, Resche F. Central nervous system hemangioblastomas, endolymphatic sac tumors, and von Hippel-Lindau disease. *Neurosurg Rev.* 2000; 23: 1-22.
54. Lonser RR, Butman JA, Huntoon K, Asthagiri A, Wu T, Bakhtian K, et al. Prospective natural history of central nervous system hemangioblastomas in von Hippel-Lindau disease. *Journal of neurosurgery.* 2014;in press.
55. Lonser RR, Vortmeyer AO, Butman JA, Glasker S, Finn MA, Ammerman JM, et al. Edema is a precursor to central nervous system peritumoral cyst formation. *Ann Neurol.* 2005; 58: 392-399.
56. Vortmeyer AO, Gnarra JR, Emmert-Buck MR, Katz D, Linehan WM, Oldfield EH, et al. von Hippel-Lindau gene deletion detected in the stromal cell component of a cerebellar hemangioblastoma associated with von Hippel-Lindau disease. *Hum Pathol.* 1997; 28: 540-543.
57. Park DM, Zhuang Z, Chen L, Szerlip N, Maric I, Li J, et al. von Hippel-Lindau disease-associated hemangioblastomas are derived from embryologic multipotent cells. *PLoS Med.* 2007; 4: e60.
58. Krishnan KG, Schackert G. Outcomes of surgical resection of large solitary hemangioblastomas of the craniocervical junction with limitations in preoperative angiographic intervention: report of three cases. *Zentralbl Neurochir.* 2006; 67: 137-143.
59. Wan JQ, Cui H, Wang Y. Surgical management of large solid hemangioblastomas of the posterior fossa. *J Clin Neurosci.* 2011; 18: 39-42.
60. Jagannathan J, Lonser RR, Smith R, DeVroom HL, Oldfield EH. Surgical management of cerebellar hemangioblastomas in patients with von Hippel-Lindau disease. *J Neurosurg.* 2008; 108: 210-222.
61. Asthagiri AR, Mehta GU, Zach L, Li X, Butman JA, Camphausen KA, et al. Prospective evaluation of radiosurgery for hemangioblastomas in von Hippel-Lindau disease. *Neuro Oncol.* 2010; 12: 80-86.
62. Simone CB 2nd, Lonser RR, Ondos J, Oldfield EH, Camphausen K, Simone NL. Infratentorial craniospinal irradiation for von Hippel-Lindau: a retrospective study supporting a new treatment for patients with CNS hemangioblastomas. *Neuro Oncol.* 2011; 13: 1030-6.
63. Capitanio JF, Mazza E, Motta M, Mortini P, Reni M. Mechanisms, indications and results of salvage systemic therapy for sporadic and von Hippel-Lindau related hemangioblastomas of the central nervous system. *Crit Rev Oncol Hematol.* 2013; 86: 69-84.
64. Yang C, Huntoon K, Ksendzovsky A, Zhuang Z, Lonser RR. Proteostasis modulators prolong missense VHL protein activity and halt tumor progression. *Cell Rep.* 2013; 3: 52-59.
65. Chan CC, Collins AB, Chew EY. Molecular pathology of eyes with von Hippel-Lindau (VHL) Disease: a review. *Retina.* 2007; 27: 1-7.
66. Wong WT, Chew EY. Ocular von Hippel-Lindau disease: clinical update and emerging treatments. *Curr Opin Ophthalmol.* 2008; 19: 213-217.
67. Gaudric A, Krivacic V, Duguid G, Massin P, Giraud S, Richard S. Vitreoretinal surgery for severe retinal capillary hemangiomas in von Hippel-Lindau disease. *Ophthalmology.* 2011; 118: 142-149.
68. Dahr SS, Cusick M, Rodriguez-Coleman H, Srivastava SK, Thompson DJ, Linehan WM, et al. Intravitreal anti-vascular endothelial growth factor therapy with pegaptanib for advanced von Hippel-Lindau disease of the retina. *Retina.* 2007; 27: 150-158.
69. Manski TJ, Heffner DK, Glenn GM, Patronas NJ, Pikus AT, Katz D, et al. Endolymphatic sac tumors. A source of morbid hearing loss in von Hippel-Lindau disease. *JAMA.* 1997; 277: 1461-1466.
70. Choo D, Shotland L, Mastroianni M, Glenn G, van Waes C, Linehan WM, et al. Endolymphatic sac tumors in von Hippel-Lindau disease. *J Neurosurg.* 2004; 100: 480-487.
71. Kim HJ, Hagan M, Butman JA, Baggenstos M, Brewer C, Zalewski C, et al. Surgical resection of endolymphatic sac tumors in von Hippel-Lindau disease: findings, results, and indications. *Laryngoscope.* 2013; 123: 477-483.
72. Lonser RR, Kim HJ, Butman JA, Vortmeyer AO, Choo DI, Oldfield EH. Tumors of the endolymphatic sac in von Hippel-Lindau disease. *N Engl J Med.* 2004; 350: 2481-2486.
73. Kim HJ, Butman JA, Brewer C, Zalewski C, Vortmeyer AO, Glenn G, et al. Tumors of the endolymphatic sac in patients with von Hippel-Lindau disease: implications for their natural history, diagnosis, and treatment. *J Neurosurg.* 2005; 102: 503-512.
74. Butman JA, Kim HJ, Baggenstos M, Ammerman JM, Dambrosia J, Patsalides A, et al. Mechanisms of morbid hearing loss associated with tumors of the endolymphatic sac in von Hippel-Lindau disease. *JAMA.* 2007; 298: 41-48.
75. Lonser RR, Baggenstos M, Kim HJ, Butman JA, Vortmeyer AO. The vestibular aqueduct: site of origin of endolymphatic sac tumors. *J Neurosurg.* 2008; 108: 751-756.
76. Butman JA, Nduom E, Kim HJ, Lonser RR. Imaging detection of endolymphatic sac tumor-associated hydrops. *J Neurosurg.* 2013; 119: 406-411.

Cite this article

Huntoon K, Lonser RR (2014) Nervous System Manifestations of von Hippel-Lindau Disease. *J Transl Med Epidemiol* 2(1): 1015.