

Research Article

Epidemiology of Gastrointestinal Parasites of Sheep in Three Agroecological Zones in West Shewa Zone, Oromia State, Central Ethiopia

Waktole Terfa*, Bersisa Kumsa, Dinka Ayana, Hora Bekele, Anna Maurizio, Cinzia Tessarin, and Rudi Cassini

Department of Veterinary Science, Mamo Mezemir campus Ambo University, Ethiopia

*Corresponding author

Waktole Terfa, Department of Veterinary Science, School of Veterinary Medicine, Mamo Mezemir campus Ambo university, Ethiopia, Tel: +251924483645, +251-943811659

Submitted: 03 May 2023

Accepted: 29 June 2023

Published: 30 June 2023

ISSN: 2379-948X

Copyright

© 2023 Terfa W, et al.

OPEN ACCESS

Keywords

- Epidemiology
- Ethiopia
- Gastrointestinal Parasites
- Infection intensity
- Prevalence
- West Shewa

Abstract

Background: Gastrointestinal parasites are one of the globally occurring most important diseases that include the groups of cestodes, trematodes, the coccidian and gastrointestinal nematodes, particularly the strongyles are the most pathogenic parasites and limit the production of sheep, goats, cattle, equine.

Methods: Across sectional study was carried out in West Shewa Zone of the Oromia regional state from August 2019 to November 2021. Flotation and McMaster techniques were used to recover and quantify Nematode, Monezia eggs and Eimeria oocysts from 659 rectally collected faeces. The parasite causing the infection was identified by the floating method, and the infection intensity was calculated by the modified McMaster method. The severity of infection was classified as mild (50–799 EPG), moderate (800–1200 EPG), or severe (>1200 EPG). Coproculture was conducted to identify Strongyle Species.

Results and discussion: Out of 659 Overall prevalence of gastrointestinal parasites in sheep was 546(82.7%) (95% CI=79.8-85.6). Among the gastrointestinal parasites genera found in the study, Strongyle spp 354 (53.72%), Eimeria sp.375 (56.90%), Monezia 120(18.21%), Strongyloides sp.113 (17.15%) and Trichuris spp 76(11.53%) were identified. The EPG/OPG count showed 2080.96, 1424.78, 240.64, 107.91and 17.45 burdens of Eimeria, Strongyle spp, Monezia, strongyloides and Trichuris spp respectively in decreasing order. Among the positive fecal samples, 204(30.96%) had single infection and while 342(51.89%) samples had mixed infections. Among sheep population (28.07%) had showed a light infection, followed by (21.85%) heavy infection and (6.98%) moderate infection intensity with gastrointestinal nematodes. The most frequently occurring nematodes were Trichostrongylus spp.33.4%), followed by Haemonchus spp. 581(30.7%), Bunostomum spp. 387(20.5%), Osephagostomum spp. 158(8.4%) and Teladorsagia 84(4.4%), whereas, Chabertia spp. 49(2.6%) was the least identified gastrointestinal nematode. Among the potential risk factors considered in the study, body condition, season, age and sex were associated with the occurrence of Nematode species; season, age, study areas and body condition with the occurrence of Eimeria species whereas only body condition and agroecology were associated with the occurrence of Monezia infections ($p < 0.05$). In general, the occurrences of high prevalence of gastrointestinal parasites in the study area suggest that they are major constraints for production and productivity of sheep.

Conclusion: The present study indicated that gastrointestinal parasitic Strongyle nematodes, Monezia and Eimeria are highly prevalent in sheep in the study areas. This study identified that season, study area, age and body condition of the sheep as risk factors for GIT parasites in sheep were significantly associated with the prevalence of git Strongyle Nematode, Eimeria and Monezia species infections of sheep.

INTRODUCTION

Sheep production in Ethiopia, with 42.9 million heads [1], and a diverse genetic resource including 15 local and two exotic breeds estimated at 99.6% and 0.3% for local breeds and hybrids, respectively, is a major livestock enterprise contributing to the country's economic development, food and nutrition security, and poverty reduction [1]. Sheep in particular are a major source of livelihoods for smallholder farmers in the sub-moist

highlands and for pastoralists. In spite of huge population and importance of small ruminants, Ethiopia has benefited little from this enormous resource owing to a multitude of problems like poor nutrition, poor animal production systems, reproductive inefficiency, management constraints, lack of veterinary care, and disease being the most important [2-5].

One of the most important diseases that limit the production of ruminants globally are the groups of the gastrointestinal

parasites(GIP) that include cestodes, trematodes, the coccidian and gastrointestinal nematodes(GIN), particularly the strongyles are the most pathogenic parasites that cause high mortality rates, as well as losses resulting from subclinical infections that are reflected in low productivity [6-8], but their impact is greater in sub-Saharan Africa in general and Ethiopia in particular due to the availability of a wide range of agro-ecological factors suitable for diversified hosts and parasite species [3,9,10].

Among the GIPs, gastrointestinal nematodes, mainly the strongyles are the most pathogenic and economically-important species affecting small ruminants worldwide [6]. Losses due to infection by helminth parasites occur through mortalities, reduced production due to sub-clinical parasitism, and direct cost associated with control measures [11,2] In Ethiopia, helminthiasis is responsible for 25% mortality and 3.8% weight loss in highland sheep [12].

GIP infections can be associated with several factors such as animal's age, breed, parasite species involved, and degree of parasitic infection [13]. There are many associated risk factors such as host age, physiological status, breed, parasite species involved, and the epidemiological patterns (husbandry practices and climate variables) determine the degree of infection [13,14] More importantly, environmental conditions such as temperature, rainfall and humidity are conducive to the development of nematode eggs [15] and free-living stages [11].

For the rational and sustainable control of gastrointestinal parasitism in sheep, a thorough knowledge of the parasites' epidemiology and their interaction with the host in a specific environment is required [16]. Therefore, knowledge of the parasite species found in a specific region, their prevalence, degree of infection, characteristics of the local climate, average flock size, and local management practices are considered essential information [17].

There have been numerous studies on internal parasites of sheep in different regions of Ethiopia [48,9,4,18-28]. These studies had shown that the predominant GI parasites in sheep were *Hemonchus* spp, *oesophagostomum* spp, *strongloides* spp, *trichostrongylus* spp, *Bunostomum* spp, *Chabertia* and *Trichuris* spp nematodes ; *Fasciola*, *Paraphistomum*spp , *Monezia* spp trematodes and *Eimeria* spp protozoan with a prevalence ranged from 2.3-100%. Gastrointestinal (GI) nematode infection rates are high in Ethiopia with an average prevalence of 75.8% [29].

Although, many studies have been conducted to estimate the prevalence of gastrointestinal tract (GIT) parasites of sheep in Ethiopia, most of them were conducted using simple fecal egg examination rather than larval identification to identify the nematodes and gastrointestinal parasites burden estimation. Moreover, there is limited information or report about the infection prevalence of coccidia in sheep in the country and most of the epidemiological studies conducted on the GI parasites in the country tended to be in the east, south, and northern regions of Ethiopia and majorly focused on the GIH and very little is known about the epidemiology of GI coccidia infecting sheep

in West Shewa Zone, Oromia, Ethiopia. Thus, the aim of the present study was to estimate the prevalence of gastrointestinal nematodes, cestodes and coccidia infections of sheep in selected areas of West Shewa zone, and to identify potential risk factors associated with the occurrence of the parasites and to estimate the burden of parasitic infections.

MATERIALS AND METHODS

Study area

The study was conducted in three selected districts (one district each from highland, midland and lowland agroecology) of West Shewa Zone of the Oromia regional state from august 2019 to November, 2021 G.C. West Shewa Zone is located in the central part of Ethiopia at 9°08'22.1"N (9.1394800°) Latitude (width), 37°51'00.6"E (37.8501600°) Longitude (length) and has larger livestock population which was estimated at 2,294,593 cattle; 1,074,939 sheep; 264,931 goats; 263,558 horses; 11,210 mules; 265,736 donkeys [1]. The predominant production system is mixed crop-livestock farming in the area.

The three districts were located west of Addis Ababa and along the highway from Addis Ababa to Nekemete, they were selected purposively from a list of all accessible districts of the zone in representation of three different agroecological areas, namely a highland area (Ejersa lafo district) is located around 47 km west of Addis Ababa with an altitude of about 2300 meters above sea level (masl). The average annual temperature 21° degrees and there is about 1350 mm of rain in a year with an average humidity of 64 % [30]. The district lies between 98° 43' 0" N to 100 03' 0" N latitudes and 410 51' 0" E to 420 80' 0" E longitude. In the district the livestock populations found are 94,955 cattle, 30,678 goats, 47,481 sheep, 10,945 horses, 6,513 donkeys, 574 mules and 37,305 poultry [31]. The 2nd study district Toke kutaye is with midland agroecology which is located about 128 km west of Addis Ababa and 12 km west direction of Ambo town along the highway from Addis Ababa to Nekemete. Geographically, the district lies between 08° 52' 30" N to 90 3' 0" N latitudes and 370 31' 30" E to 370 42' 0" E longitude and the altitude and rainfall of the area ranges from 1600-3194m above sea level and 800- 1100mm respectively and has temperature range of 10-29°C, it has 185,596 heads of cattle, 27,349 sheep, 24,782 goats, 84,530 chickens, 10,850 horses, 2,371 mules, and 1,398 donkeys according to [32]. A lowland area representative Ilu Gelan district, geographically, lies between 08° 49' 0" N to 90 5' 30" N latitudes and 370 9' 30" E to 370 25' 0" E longitude. It is located around 200 km west of Addis Ababa. As most parts of the district are found in the low land, the mean annual temperature of the area is relatively high. The highest mean annual rainfall was 1351 mm and recorded in July whereas the lowest mean annual rainfall was 11.2 mm and recorded in February. The average temperature of the district is 27.3°C with average altitude of 1665 - 1790 m.a.s.l. Total livestock population of the district was 108,732 (cattle), 37780 (sheep) and 38980 (goats) [33-36].

Similarly, a list of Peasant Associations (PAs) was prepared

based on the following inclusion criteria: i) accessibility for vehicle all year round, ii) availability of veterinary clinics and animal health assistants from the Ministry of Agriculture, iii) willingness of peasant farmers' representatives to participate in the study, iv) availability of small ruminants in the study sites. Accordingly, a total of six PAs (two per each selected district) were purposively selected.

Study animal and sample size determination

The study animals were 659 sheep of both sexes (316 male and 343 female), under traditional management production systems. In this study the risk factors such as animal's species, age, sex, body condition, and peasant association were included. Those animals with the age of between three months and less than one year were considered as young while those greater than or equal to one year were included as adults as described by [37]. The age of the animal was estimated by looking at the dentition pattern of the animals according to [38] and also by owners' response. Body condition score was done according to [34,37], and recorded as poor, medium or good. Simple random sampling technique was employed to select the study animals. The sample size was determined by the formula described by [39], at 95% confidence level and 5% precision, and considering 50% estimated prevalence as there was no previous such combined study at the current study area. However, to increase the precision of the study sample size was increased to 659

$N = [1.962 \text{ Pexp} (1 - \text{Pexp})] / d^2$ Where, N = sample size; Pexp = expected prevalence; D = desired absolute precision.

Ethical Clearance

All the participants in this study were treated according to the ethical standards of Addis Ababa University, and the protocol was assessed and approved with Certificate Reference number: VM/ERC/17/05/13/2021 by the Addis Ababa University Research Ethics Review Committee. The participants were informed about the purpose and the methods of the study. Oral consent was obtained from each owner and participant before commencement of the study.

Study design

A cross sectional study was carried out from August 2019 to November 2021 to collect data on events associated with gastrointestinal parasites of sheep and identify nematode parasites, cestodes and coccidia. The study animals in each selected districts and peasant association are categorized into two age groups, that is, less than 1 year as young and above 1 year as adult.

For all sampled animals, simple floatation and modified McMaster egg counting technique was carried out to determine the prevalence and the number of eggs (oocysts) per gram (EPG/OPG) burdens in the faecal samples. The degree of infections was determined by counting the eggs per gram faeces through modified McMaster technique and classified as light infection

level as (50-800 FEC), medium (801-1,200 FEC) and heavy (>1,200 FEC) according to [34] given for a mixed infection in small ruminants fecal culture of pooled gastrointestinal strongyles positive samples and L3 identification was also carried out according to [35,36].

Study methodology

Faecal sample collection and examination: About 5-10g faecal samples were collected from each animal directly from the rectum using disposal gloves and coded with the animal identification number by permanent marker. After collection, fecal samples were transported to the Veterinary laboratory of college of agriculture and veterinary sciences, Mamo Mezamir Guder Campus of Ambo University and processed for qualitative and quantitative coprological examination and coproculture on the day of collection or stored in refrigeration at +4°C to be processed on the next day. In the laboratory, the samples were processed by both centrifugal floatation and McMaster technique to detect the presence of GIP and determine faecal egg count (EPG)/ faecal oocyst count (OPG) following the procedure described by [34].

During sampling, data with regard to age, sex, species, body condition and other relevant factors were collected for each sampled animal. The age of the animals was determined based on the farmers' response and teeth inspection. Body condition scoring (BCS) were made according to [40] and recorded as 1 (poor), 2 (average or medium) or 3 (good). Faecal samples were analyzed for helminthic eggs, and coccidial oocysts using a modified McMaster technique [41]. Identification of characteristic helminthic eggs of Strongyle type egg, Trichuris, Strongyloides, Monezia species, and Eimeria oocysts were done at genus level only and were based on the morphological features as described by [6].

The McMaster egg counting technique was carried out for each faecal sample in order to determine the number of eggs per gram of faeces as described by [42]. Briefly, 3g of the faeces were mixed in 42 ml of saturated NaCl and Sugar solution (by the ratio of 400g of NaCl + 1 liter of tap water + 500g of Sugar = 1.28 specific gravity) with a sensitivity of 50 EPG of faeces [34].

Faecal culture and L3 identification: At present, the only practical method available for routine laboratory estimation of the proportions of GIT worm genera present in the living animal is to identify the larvae that develop in faecal cultures [36]. Faecal samples positive for strongyle type eggs were subjected to coproculture and the infective 3rd stage larvae (L3) were identified to genus level. Faecal samples from nematode eggs positive animals were pooled and cultured for harvesting of third stage larvae and identification of genera of nematode eggs. Pooled faecal samples were then finely broken using stirring device, kept moist and brittle; the mixtures were transferred to Petri dishes and placed at 27°C in an incubator for 7 to 10 days. The culture was kept moist by adding water every 2 days. During this period, the larvae was hatched from the eggs and developed into L3. Finally, larvae were recovered using the modified

Baermann technique [34]. The presence of larvae was assessed by using a stereomicroscope, when present; two drops of larval suspension were mixed with drop of lugols iodine on glass slide, and examined at low magnification power for identification. From each culture, the third-stage larvae (L3) were morphologically differentiated and identified according to [35,36]. Conventional characteristics for identification (total length, esophagus length, tail sheath length, filament length and the number of intestinal cells) of infective larvae from gastrointestinal nematode genera/species were microscopically examined.

Data analysis

All collected data were entered into a Microsoft Excel spreadsheet, edited and coded; and then summarized by descriptive statistics like mean and proportion. Multivariate logistic regression analysis was used to see the association of the potential risk factors considered for the study with infection by nematodes, cestodes and Eimeria species infection. Finally, the model fitness was assessed by the Hosmer–Lemeshow goodness-of-fit test [43]. For the data analysis STATA 14.2 software was used. The study considered a 95% level of confidence and 5% desired level of precision.

RESULTS

Prevalence and parasite burden of gastrointestinal Nematodes, cestodes and Eimeria

From the fecal examination results, it can be seen that the GIP infection rate of Sheep in west Shewa zone was very high. Of the 659 Sheep examined, 82.7 % (n = 546; 95% CI=79.8-85.6) were found to harbor one or more parasite species. The gastrointestinal parasites infecting sheep in the sampled areas in this study were Emeriasis, strongyliasis, Moneziasis, strongloidiasis and trichuriasis (Table 1). Table 1 shows the gastrointestinal parasites genera found in the study, are Strongyle spp 354 (53.72%), Eimeria sp. 375 (56.90%), Monezia 120 (18.21%), Strongyloides sp. 113 (17.15%) and Trichuris spp 76 (11.53%) the prevalent infections. Out of the ovine infected by gastrointestinal parasites 204 (30.96%) had single infection and 342 (51.89%) ovine had mixed infections (Table 1). The present study revealed that most of the samples were found with multiple infections than single infection with gastrointestinal nematodes and Eimeria coinfection being high. The Table 1 further more indicated the mean burdens EPG/OPG count and prevalence rates of 2080.96 (56.90%), 1424.78 (53.72%), 240.64 (18.21%), 107.91 (17.15%) and 7.45 (11.53%) Eimeria, Strongyle spp, Monezia, strongyloides and Trichuris spp respectively in decreasing order (Table 1).

Table 2 shows the prevalence and abundance of GIP species infection varied across agroecologies, age category, Sex, Season and bodycondition. The overall mean faecal egg/oocyst count for gastrointestinal nematode, Eimeria spp. and Monezia spp. positive animals combined was generally in the highland having a highest mean faecal egg/oocyst count than the midland and lowland. The prevalence of GIS and Eimeria in High land and mid lands were higher than in lowland agroecology. Young sheep

had overall mean faecal egg count of (2977.3) Eimeria, (2505.8) Nematode spp. and (319.1) Monezia spp. which is significantly higher than that of adults. The overall mean faecal egg/oocyst count of gastrointestinal parasites species infections for positive animals was high for categories of risk factors like highland, poor body conditions, wet season and when animals are young (Table 2).

The coproculture of infected faecal samples revealed that, the about third of most identified type of GIS was Trichostrongylus spp. followed by Haemonchus spp., Bunostomum spp., Osephagostomum spp. and Teladorsagia, whereas, Chabertia spp. was the least observed GIS (Table 3).

The intensity of infection with GIN in Sheep was graded into four categories and presented in (Table 4), out of the infected Sheep (28.07%) showed a light infection, followed by (21.85%) heavy infection and (6.98%) moderate infection intensity with GIN; whereas 43.10% were with no infection.

Logistic regression analysis of potential risk factors for GIP infection

The Multivariable logistic regression analysis revealed that among the potential risk factors considered in the study, body condition, season and agroecology were associated with the occurrence of Strongyle nematode species (Table 5); season, age, study areas and body condition were associated with the occurrence of Eimeria species in sheep (Table 7) whereas only body condition and agroecology are associated with the occurrence of Monezia infections (Table 6). The odds of occurrence of gastrointestinal Strongyle nematodes infection in sheep reduces about 5.6 times when the agroecology is lowland (Ilugalan) than being highland (Ejersalafo) (P=0.000). Similarly, the odds of gastrointestinal Strongyle nematodes in sheep reduces about 16.2 times and 5.8 times when the body condition is good or medium respectively than in sheep with poor body condition (P=0.000) (Table 5). The odds of acquiring gastrointestinal Strongyle nematodes infection in sheep increases about 4.0 times when it's wet season than being in dry season (P=0.000) (Table 5).

The odds of acquiring Monezia infection by sheep in Ejersalafo were 1.63 times higher as compared to Ilugalan (Table 6). The odds of acquiring Eimeria infection by sheep with good body conditions is 3.49 times lower as compared to sheep with poor body conditions and the odds of acquiring Eimeria infection by young sheep was 2.55 times higher as compared to adult sheep (Table 7). The odds of acquiring Eimeria infection was 4.368604 and 2.09 times higher in Ilugalan and Tokecutaye respectively as compared to sheep in Highlands (Ejersalafo); the odds of acquiring Eimeria infection by sheep in mutullu village was 3.77 times high as compared to Birbirsa village (Table 7).

DISCUSSION

The gastrointestinal parasites of sheep are one of the major important parasitic diseases that reduce the productivity of sheep

Table 1: Prevalence and burden of ovine gastrointestinal parasites in west Shewa zone

| Parasite genera/species | Frequency | % | Std. Err. | [95% Conf. Interval] | Mean | Std. Err. | 95% Conf. Interval |
|-------------------------|-----------|-------|-----------|----------------------|---------|-----------|--------------------|
| Strongyle spp | 354 | 53.72 | 1.94 | 49.9-57.5 | 1424.78 | 121.9 | 1185.4-1664.1 |
| Trichuris spp | 76 | 11.53 | 1.25 | 9.1-14 | 17.45 | 2.5 | 12.51-22.4 |
| strongloides | 113 | 17.15 | 1.47 | 14.3-20.0 | 107.91 | 18.7 | 71.3-144.5 |
| ginematodes | 379 | 57.51 | 1.93 | 53.7-61.3 | 1529.15 | 129.1 | 1275.6-1782.7 |
| Monezia | 120 | 18.21 | 1.50 | 15.3-21.2 | 240.64 | 50.6 | 141.3-340 |
| Eimeria | 375 | 56.90 | 1.93 | 53.1- 60.7 | 2080.96 | 220.3 | 1648.3-2513.6 |
| Overall gitparasites | 545 | 82.70 | 1.47 | 79.8-85.6 | | | |

Table 2: Prevalence and abundance (mean output) of selected parasitic taxa in the subgroups identified by the individual (BCS, age and sex) and environmental (season and agroecology) factors in sheep

| Risk Factors | Variables Category | No examined | GIS spp. | | | Monezia spp. | | | Eimeria Spp. | | |
|----------------|--------------------|-------------|----------------|-------------------|----------|----------------|-------------------|-----------|----------------|-------------------|-----------|
| | | | No(%) positive | Mean output (EPG) | Min- Max | No(%) positive | Mean output (EPG) | Min - Max | No(%) positive | Mean output (OPG) | Min - Max |
| Study area | Ilugeln | 244 | 83(34.0) | 350.2 | 0-14000 | 36(14.8) | 224.0 | 0-14000 | 105(42.2) | 3055.1 | 0- 52000 |
| | Tokekutaye | 166 | 102(61.4) | 830.5 | 0-12500 | 27(16.3) | 128.9 | 0-4000 | 115(69.3) | 1432.5 | 0- 10000 |
| | Ejersalafo | 249 | 169 (67.9) | 2874.0 | 0-16200 | 57(22.9) | 331.4 | 0-22000 | 155(63.5) | 1558.6 | 0- 80000 |
| Body condition | Poor | 242 | 199 (81.2) | 2921.2 | 0-16200 | 52(21.2) | 456.5 | 0- 22000 | 155(63.3) | 2467.8 | 0-52000 |
| | Medium | 253 | 116(46.4) | 748.5 | 0-12500 | 47(188) | 140.6 | 0- 4000 | 154(61.6) | 2236.8 | 0- 80000 |
| | Good | 164 | 39(23.8) | 259.8 | 0-6200 | 21(12.8) | 76.5 | 0-3000 | 66(40.2) | 1269.8 | 0- 50000 |
| Age category | Young | 216 | 130 (58.0) | 2337.9 | 0- 16200 | 49 (21.9) | 319.1 | 0- 22000 | 142(63.4) | 2977.3 | 0- 80000 |
| | Adult | 443 | 224 (51.5) | 979.5 | 0-15800 | 71(16.3) | 202.4 | 0- 14000 | 233(53.6) | 1643.9 | 0- 50000 |
| Sex | Female | 343 | 209 (60.9) | 1764.1 | 0-15800 | 67(19.5) | 311.8 | 0- 22000 | 174(50.7) | 1332.9 | 0- 80000 |
| | male | 316 | 145 (45.9) | 1056.5 | 0-16200 | 53(16.8) | 163.4 | 0- 8000 | 201(63.6) | 2892.9 | 0- 52000 |
| Season | dry | 335 | 137 (40.9) | 368.209 | 0-8800 | 56(16.7) | 193.9 | 0- 22000 | 178(53.1) | 878.7 | 0- 10000 |
| | Wet | 324 | 217 (67.0) | 2517.22 | 0-16200 | 64(19.8) | 289 | 0- 14000 | 197(60.8) | 3324.1 | 0- 80000 |

Table 3: Number and relative percentage of third-stage larvae of the different genera of GIS identified in the whole study area (n=1,890). Larvae ascribable to Strongyloides were not considered here

| | Haemonchus | Trichostrongylus | Bunostomum | Oesophagostomum | Teladorsagia | Chabertia |
|------------|------------|------------------|------------|-----------------|--------------|-----------|
| n larvae | 581 | 631 | 387 | 158 | 84 | 49 |
| relative % | 30.7% | 33.4% | 20.5% | 8.4% | 4.4% | 2.6% |

Table 4: Estimation of infection degrees of Ovine gastrointestinal nematodes

| Infection Degrees | Freq. | Percent (Proportion) | Std. Err. | [95% Conf. Interval] |
|-------------------|-------|----------------------|-----------|----------------------|
| No infection | 284 | 43.10 | 1.93053 | 39.30486 46.88634 |
| Light | 185 | 28.07 | 1.75177 | 24.63311 31.51257 |
| Medium | 46 | 6.98 | 0.99337 | 5.02972 8.93083 |
| Heavy | 144 | 21.85 | 1.61097 | 18.68804 25.01454 |
| Total | 659 | 100.00 | | |

Table 5: Multivariate Logistic regression analysis of ovine strongyle nematodes prevalence

| | strongyle | Odds Ratio | Std. Err. | z | P> z | 95% Conf. Interval] | |
|---------------|------------|------------|-----------|--------|-------|---------------------|----------|
| | | | | | | Lowe | upper |
| season | dry | reference | | | | | |
| | wet | 4.04713 | 0.814344 | 6.95 | 0.000 | 2.728176 | 6.003739 |
| bodycondition | poor | reference | | | | | |
| | medium | 5.812142 | 0.03975 | -7.62 | 0.000 | 3.695554 | 9.140985 |
| | good | 16.23266 | 0.016788 | -10.23 | 0.000 | 9.515418 | 27.69186 |
| agroecology | ejersalafo | reference | | | | | |
| | tokekutaye | 1.538172 | 0.161206 | -1.74 | 0.082 | 0.946104 | 2.500752 |
| | ilugelan | 5.630523 | 0.041601 | -7.38 | 0.000 | 3.557673 | 8.911099 |
| cons | | 5.212705 | 1.208576 | 7.12 | 0.000 | 3.309104 | 8.211375 |

Table 6: Multivariate Logistic regression analysis of Monezia of ovine

| Monezia | Odds Ratio | Std. Err. | z | P> z | [95% Conf. Interval] |
|---------------|------------|-----------|-------|-------|----------------------|
| agroecology | | | | | |
| Tokekutaye | 1.4961305 | 0.1736746 | -1.55 | 0.121 | 0.89907 2.489691 |
| llugelan | 1.6289492 | 0.1455466 | -2.06 | 0.040 | 1.023524 2.59249 |
| bodycondition | | | | | |
| midium | 1.1502417 | 0.1966667 | -0.62 | 0.536 | 0.738304 1.79202 |
| good | 1.7296959 | 0.163856 | -1.93 | 0.053 | 0.992479 3.014521 |
| cons | 0.3450955 | 0.0655557 | -5.6 | 0.000 | 0.237816 0.500768 |

Table 7: Multivariate Logistic regression analysis of ovine Eimeria

| Eimeria | Odds Ratio | Std. Err. | z | P> z | [95% Conf. Interval] |
|---------------|------------|-----------|-------|-------|----------------------|
| agroecology | | | | | |
| Tokekutaye | 2.08504 | 0.657641 | 2.33 | 0.02 | 1.123666 3.868935 |
| llugelan | 4.368604 | 1.462641 | 4.4 | 0.000 | 2.266491 8.420371 |
| Wetseason | 1.142272 | 0.256914 | 0.59 | 0.554 | 0.735058 1.775078 |
| Adultage | 2.549556 | 0.078805 | -4.66 | 0.000 | 1.719663 3.779949 |
| bodycondition | | | | | |
| midium | 1.103247 | 0.180672 | -0.49 | 0.622 | 0.74646 1.63057 |
| good | 3.493298 | 0.065407 | -5.47 | 0.000 | 2.232281 5.466668 |
| kebele | | | | | |
| olankomiBobe | 1.228982 | 0.226401 | -0.74 | 0.459 | 0.712369 2.120245 |
| mutullu | 3.773676 | 1.390298 | 3.6 | 0.000 | 1.83301 7.768988 |
| birbirsa | 1(omitted) | | | | |
| llaalaa | 1.373184 | 0.263301 | -0.88 | 0.38 | 0.676033 2.789263 |
| Allewarailu | 1(omitted) | | | | |
| cons | 1.523978 | 0.376923 | 1.7 | 0.088 | 0.93854 2.474598 |

raised by farmers using traditional husbandry management system in west Shewa zone. GIH and Eimeria coccidia infections in sheep are globally present however with differences in the prevalence rate and infection intensity attributed to variations in sheep’s immunity, climate, and feeding and management conditions and remains a major problem to sheep keepers. The present study demonstrated the overall prevalence of gastrointestinal parasitic infection was 82.7 %, and an average infection intensity and prevalence rates of GIP Eimeria, Strongyle spp, Monezia, Strongyloides and Trichuris spp were 2080.96 (56.90%),1424.78 (53.72%), 240.64 (18.21%),107.91 (17.15%) and 17.45 (11.53%), respectively in decreasing order. Our results on prevalence of GIPs were in line with the reports of a previous study in [9]; [44,21-24,45] who also found high prevalence of GIP that range 71-87% in different places in Ethiopia. Similarly, [46] reported overall prevalence 74.76% GI parasitic infection in sheep in northern-Nigeria; [26] in Kenya the GI parasite prevalence in the sheep populations reached as high as (75.7%); [47] reported (72%) prevalence of GIP of Sheep in and around Rawalpindi and Islamabad, Pakistan.

However, our result is less than the report of [20,48,49] in eastern and southern part of Ethiopia who found the overall prevalence of 90-97.8 %; elsewhere [50] of Djallonke sheep in Ayeduase, Kumasi, Ghana a prevalence rate of 98.2% with the total infection rate of GIT nematodes and coccidia oocysts were 94.5% and 51.8%, respectively; [51] where the GIN and coccidial infection rates and an average infection intensity in Kazakh sheep, China was (96.96%). The overall higher prevalence

of GIP infections of sheep in the surveyed areas could be attributed to lower immunity of hosts as a result of malnutrition, dependence on grazing in deteriorated grazing lands, use of the same contaminated pasture for grazing throughout the year by different mixed species of animals, grazing of young and adult animals together and existence of good agro-climatic conditions that might provide ideal conditions for development and transmission of the endoparasites.

The present finding in sheep was higher than the prevalence rates reported in other areas of Ethiopia [28]in sheep in Hawasa town, southern Ethiopia that showed a GIN infection rate of (51.3%); [52] in Adami Tulu Jiddo Kombolcha district, East Shoa zone of Oromia region, Ethiopia 117(60.9%);Southern Ethiopia an infection rate of 43.5% by [53,54] the infection rate of (63.33%) in ovine in East and Western Hararghe, Ethiopia; [18] who reported (24.7%) prevalence of gastrointestinal nematodes in sheep in West Oromia, Ethiopia; [15] in Wayu Tuka and Diga district west Ethiopia(44.0%) ; [55] in sheep in kurmuk woreda, Assosa Zone of Benishangul Gumuz Regional State, Western Ethiopia (26%); [55] prevalence rate of 185 (59.11%) Monezia (2.5%), Eimeria spp (0.260%)in sheep in and around Gondar town, Northwest, Ethiopia; [10] infection rate of (63.16%) gastrointestinal parasites were detected in sheep in Minna Abattoir, Niger Sate, Nigeria; [56] in Bangladesh revealed (67.9%) sheep were infected with various types of GI helminths; [57] in Brazil the overall prevalence rates of nematodes, coccidia, and cestodes were 53.9%, 46.5%, and 4.3% in the Maseru district, respectively and for the Quthing district he also indicated the prevalence rates

of 65.0%, 38.2%, and 0.9% for nematodes, coccidia, and cestodes, respectively; The observed differences in prevalence could be attributed to factors such as different management practices, breeds, and the difference of health care activities, sample size and the variation in geographical and climatic conditions and season under which the various studies were carried out. Among the factors influencing strongyle infections, geographical conditions, temperature, climate, rainfall, humidity, soil conditions and farm management are considered the most important. This could be related to the country's extremes temperature and rainfall which is the most important factor that influence the development, distribution and survival of nematode parasites [2].

The present study revealed that most of the samples were found with multiple infections 342(51.89%) than single infection with 204(30.96%) gastrointestinal nematodes and *Eimeria* coinfection being high. The findings of present study are in close agreement with the reports of [58] in Brazil where 68.61% mixed infection prevalence rate of gastrointestinal parasites was detected in sheep herds, between helminths and protozoans [57]. The overall prevalence rates of nematodes, coccidia, and cestodes were 53.9%, 46.5%, and 4.3% in the Maseru district, respectively.

Our present infection intensity record of *Eimeria* Strongyle spp *Monezia*, *Strongyloides* and *Trichuris* spp were 2080.96, 1424.78, 240.64, 107.91 and 17.45, respectively in sheep from west Shewa is in general agreement with the average infection intensity reported by [51]; [58] in Kazakh sheep and sheep herds from the Brazilian Pampa biome, Rio Grande do Sul state, Brazil who found similar findings. Though the GIN and coccidial infection rate were higher than our result the average infection intensity was in general agreement.

Our results to classify the severity of infection based on the level of EPG as proposed by [34] revealed that 28.07% of the infected sheep had light nematode faecal egg counts per gram (50 to 800), 21.85% of the animals had heavy (>1200) EPG counts and 6.98% showed moderate (800 to 1200) degree of infection, are on the same line with [9] west Ethiopia; [22] from Southern Ethiopia and [50] in Kumasi, Ghana who also reported sheep were lightly, highly, and moderately infested, respectively; but it disagrees with reports of [48,22-24,52] in sheep from different parts of Ethiopia. The difference in degrees of infections could be due to the variations in animal managements, breed differences, immunity, season, agroecology, feed etc.

The coproculture of infected faecal samples revealed that, the most identified type of GIS was *Trichostrongylus* spp. 631(33.4%), followed by *Haemonchus* spp. 581(30.7%), *Bunostomum* spp. 387(20.5%), *Osephagostomum* spp. 158(8.4%) and *Teladorsagia* 84(4.4%), whereas, *Chabertia* spp. 49(2.6%) was the less frequently observed GIS. A predominance of *Haemonchus* spp. and *Trichostrongylus* spp. nematodes was observed in this study. The considerably high prevalence of *Trichostrongylus* and *Haemonchus* in west Shewa was consistent with the reports presented in studies conducted by [11,48,20] in Ethiopia who have also reported similar findings. Whereas higher proportion

of *Haemonchus* followed by *Trichostrongylus* in sheep were reported by [29,14,53,18,49,27] in Ethiopia and [10] in Nigeria; and [58] in Brazil. The variation in these reported findings could be attributed to differences in management system, topography, de-worming practices, and climatic condition that favor the survival of infective stage of the parasite.

In our findings *Eimeria* spp was the most frequently 56.90% recovered GIP followed by 53.72% *Strongyles* spp., *Monezia* 18.21%, *Strongyloides* spp. 17.15% and *Trichuris* spp 11.53%. Among the gastrointestinal parasites observed in this study, *Coccidia* have the highest prevalence. This is in conformity with the findings of [46,50,57,58], who also reported a high prevalence of *Coccidia* and gastrointestinal strongyle from different countries.

In our findings *Eimeria* spp was the most frequently 56.90% recovered gastrointestinal parasites and this prevalence of infection by *Eimeria* species is in agreement with previous reports of [50] who found infection rate of coccidia oocysts (51.8%), in Ayeduase, Kumasi, Ghana; [47] in Sheep in and around Rawalpindi and Islamabad, Pakistan *Coccidia* (51.61%) and [57] in Brazil who recorded the overall prevalence rates of coccidia was 46.5% in the Maseru district. The high prevalence of *Coccidia* obtained in this study could be as a result of the management system operated by most small ruminants' owners especially during the rainy season when animals are confined to avoid damage to crops. Consequently, such animals are with the pens not properly cleaned. These factors with the high humidity of the rainy season predispose them to the parasitic infections.

However, our result of *Eimeria* spp. prevalence disagrees with the very high records of (89.32%) [46] In northern-Nigeria; (90.89%) of coccidia infection rate by [51] of China; (70.55%) by [58] in Brazil; and with the previous *Eimeria* prevalence reports in sheep which were lower than our results in [9] 26.7% reported in sheep in west Oromia; [55] prevalence rate of (12.5%) *Eimeria* spp parasites of sheep and goat in and around Gondar town, northwest, Ethiopia. The observed difference could be attributed to the difference in the sample size, age, host management situations like feeding, housing and watering and hygiene status of animals, difference in environmental factors, like season, humidity and temperature of study areas.

The cestode observed in the sheep in the present study was *Monezia* with prevalence of 18.21%, the occurrence of these parasites is low as compared to *Eimeria* and strongyles parasites. The present report of *Moniezia* infection rate was in conformity with reports of in Jeldu district, west Showa Zone of Oromia Regional State, Ethiopia 20.2%; [58] *Moniezia expansa* (20.39%) from sheep herds in Brazil. However, our result is less than the report of [48] in sheep in eastern part of Ethiopia during the dry season 27.65%, and greater than the prevalence reported in [48] in Dire Dawa, East Ethiopia (13.67%); [21] in the Afar, Ethiopia (8.53%) *Monezia* infection; [55] (0.26%) *Monezia* of sheep in and around Gondar town, northwest; [57] in Brazil the overall prevalence rates of cestodes was 4.3% in the Maseru district;

[10] 8 (6.8%) detected *Moniezia* spp. in sheep in Minna Abattoir, Niger State, Nigeria; [46] Specific prevalence of *Moniezia* species infections (4.85%) in sheep at slaughter in Kano central abattoir, northern-Nigeria and [56] *Moniezia* sp. (3.8%) of sheep in Sherpur, Bangladesh was reported. This might be due to the fact that the climatic condition is not equally suitable for survival of the intermediate host (oribatid mite) and transmission of *Moniezia* in different study areas.

The Multivariable logistic regression analysis revealed that among the potential risk factors considered in the study, body condition, season and agroecology were associated with the occurrences of GIS nematode and *Eimeria* species; whereas only agroecology is associated with the occurrence of *Moniezia* infections. The odds of occurrence of gastrointestinal Strongyle nematodes infection in sheep reduces about 5.6 times when the agroecology is lowland (Ilugalan) than being highland (Ejersalafo) ($P=0.000$). Similarly, the odds of gastrointestinal Strongyle nematodes in sheep reduces about 16.2 times and 5.8 times when the body condition is good or medium respectively than in sheep with poor body condition ($P=0.000$). The odds of acquiring gastrointestinal Strongyle nematodes infection in sheep increases about 4.0 times when it's wet season than being in dry season ($P=0.000$).

The odds of occurrence of gastrointestinal strongyle nematodes infection in sheep reduces about 16.233 times and 5.812 times when the body condition is good or medium respectively than in sheep with poor body condition. The odds of acquiring gastrointestinal nematodes infection in sheep increases about 4.05 times when it's wet season than being in dry season. Sheep in good and medium body condition were more likely to be free from GIN infection by ($P = 0.000$) about 16.373 and 5.81 times, respectively than in sheep with poor body condition.

The odds of occurrence of gastrointestinal strongyle nematodes infection in sheep showed no significant associations ($p > 0.05$) in prevalence between different sex male and female sheep this finding conforms to [53,21,22] who found in south, Afar and West Hararge of Ethiopia respectively no significant difference ($p > 0.05$) in prevalence between different sex; [10] in Minna Abattoir, Niger State, Nigeria reported that there was no significant difference ($p > 0.05$) on the infection rate in relation to gender. Our findings disagree with [54,23,55,45] who reported high occurrence of gastrointestinal nematodes infection in female than in male sheep at significant difference ($P < 0.05$), and with [28] whose findings revealed the prevalence was high in male than in female and The differences may be related with sample size variation, physiologic conditions like pregnancies, lactating state in female genders in the study.

The multivariable logistic regression analysis revealed that among the potential risk factors considered in this study, agroecology was associated with the occurrence of all GIP (*Strongyle*, *Moniezia* and *Eimeria*) species in sheep. In this study there was significant Strongyle nematode variation among PAs and districts ($p > 0.05$) of the study areas in West Shewa zone. The odds of occurrence of gastrointestinal Strongyle nematodes infection in sheep reduces about 5.6 times when the agroecology is lowland

(Ilugalan) than being highland (Ejersalafo) ($P=0.000$). The odds of acquiring gastrointestinal Strongyle nematodes infection in sheep increases about 4.0 times when it's wet season than being in dry season ($P=0.000$).

The odd of acquiring *Moniezia* infection by sheep in Ejersalafo was 1.63 times higher as compared to Ilugalan. The odds of acquiring *Eimeria* infection by sheep in mutullu village was 3.77 times high as compared to Birbirsa village, the odds of acquiring *Eimeria* infection was 4.368604 and 2.09 times higher in Ilugalan and Tokekutaye respectively as compared to sheep in Highlands (Ejersalafo). This findings conforms with [9] who reported association between prevalence rates and EPG with agro-ecology where higher values were recorded for lowland areas followed by mid altitude areas with lowest values in highland areas in west Ethiopia; [54] who reported significant nematode variation among Pas ($p > 0.05$) of the four districts of the two zones, west Hararghe (Chiro and Tullo districts) and east Hararghe (Meta and Haramaya districts) East, Ethiopia and [21] who reported significant variation in overall parasite prevalence was observed between the two study districts ($OR=0.169$, $p=0.000$) of Afar. These findings may be attributed to the fact that areas receiving high annual rainfall provide optimum conditions of humidity and temperature required for development and dissemination of infective stages but disagrees with [53] who reported that there was no significant association between study areas and GIP infection occurrence in southern Ethiopia. The difference between this and ours could be that the sampling was from similar rainfall, ecology and environmental factors in their survey.

The multivariable logistic regression analysis revealed that among the potential risk factors considered in this study, body conditions was associated with the occurrence of gastrointestinal Strongyle nematodes and *Eimeria* species in sheep. In this study, a significant difference ($P < 0.05$) was observed in GIT nematode infection in relation to body condition where a higher prevalence of Strongyle nematodes was recorded in sheep with poor body condition and odds of occurrence of gastrointestinal strongyle nematodes in poor body condition is about 16.2 times and 5.8 times higher than animals with the good or medium body condition respectively. Similarly, the odds of gastrointestinal Strongyle nematodes in sheep reduces about 16.2 times and 5.8 times when the body condition is good or medium respectively than in sheep with poor body condition ($P=0.000$). The odds of acquiring *Eimeria* infection by sheep with good body conditions is 3.49 times lower as compared to sheep with poor body conditions and the odds of acquiring *Eimeria* infection by young sheep was 2.55 times higher as compared to adult sheep. This poor body condition might be due to malnutrition, other concurrent disease or the current parasitic infection which lead to poor immunological response to infective stage of the parasite. This finding agrees with [54,21, 22,55,25,46,28] in Ethiopia and [56] in Bangladesh who revealed also helminth infection was significantly ($P < 0.05$) higher in poor body conditioned sheep as contrasted to good body conditioned sheep but contradicts the findings of [9,23]; who reported that body condition of the animal did not show significant association with prevalence of

the parasite. This could be explained by the fact that loss of body condition in the study animals could be due to other factors, such as seasonal change of forageable feedstuff and the presence of other concurrent disease conditions.

Results of the present study in sheep showed seasonal variation in the prevalence and burden of gastrointestinal nematodes, and *Eimeria* with the prevalence and level of burden of infection being higher during the rainy season than the dry season. The prevalence and burden of gastrointestinal strongyle nematodes and *Eimeria* parasites 2517.2 (67.0%), 3324.1 (60.8%), were higher during wet season than the dry season 368.2 (40.9%), 878.7 (53.1%) respectively with occurrence of strong association OR 4.047 (P=0.000, CI=2.7- 6) for gastrointestinal strongyle nematodes. This agrees with [11,9,55]. During the rainy season, high humidity and moderate temperature are factors that facilitate the survival and sporulation of the oocysts [59]. Climatic conditions, particularly rainfall, frequently are considered to account for differences in the prevalence of GI-parasitic infection, since infective stages such as eggs, cysts and oocysts are known to survive longer in cool, moist conditions [60].

The high prevalence and burden of gastrointestinal strongyle nematodes, *Monezia* and *Eimeria* parasites 2337.9 (58.0%), 319.1 (21.9%), 2977.3 of GIS, in young sheep were observed than in adult sheep 979.5 (51.5%), 202.4 (16.3%), 1643.9 (53.6%) respectively, though no significant association occurred. The odds of occurrence *Eimeria* infections in sheep increase about 2.549556 times in young sheep than when the age category is adult. This agrees with [9,50,54,55,58] which showed that the susceptibility and pathogenicity *Eimeria* infections were greater in young animals than in mature animals; but disagrees with [21] in the Afar, Ethiopia; [22] in and around Hirna, western Hararghe, Ethiopia; [23] in and around Yabello, south Ethiopia ; [53,45] in and around Alage Ethiopia had no statistical association ($p>0.05$) for parasitic infection between different age groups; In Bangladesh [56] Parasitic counts in lambs, young and adult showed no significant variations (P=0.511) from one other; [10] in Minna Abattoir, Niger State, Nigeria reported that there was no significant difference ($p>0.05$) on the infection rate in relation to age. These observed associations could be related to the immunological immaturity, age-specific susceptibility in lambs, or might be due to a limited previous exposure and immaturity of the immune system that resulted in higher development of the parasite and adult animals may acquire immunity to the parasites through frequent challenge and expel the ingested parasite before they establish infection. The overall higher incidence of nematodes infestation in the areas surveyed could be attributed to lower immunity of hosts as a result of malnutrition.

CONCLUSION AND RECOMMENDATIONS

In conclusion, our study has confirmed gastrointestinal parasitic infections are highly prevalent in West Shewa. The most widespread GIS in this zone were *Trichostrongylus* spp. followed by *Haemonchus* spp., *Bunostomum* spp., *Osephagostomum* spp. and *Teladorsagia*, whereas, *Chabertia* spp. was the least

observed. This study identified that season, study area, age and body condition of the sheep as risk factors for GIT parasites in sheep were significantly associated with the prevalence of git Strongyle Nematode, *Eimeria* and *Monezia* species infections of sheep. We therefore recommend selective treatment of sick animals during wet season, regular monitoring and management of infections in traditional sheep herds as well as the further parasitological study using the molecular techniques and monthly dynamic *Trichostrongylus* spp. followed by *Haemonchus* spp., *Bunostomum* spp., *Osephagostomum* spp. and *Teladorsagia*, whereas, *Chabertia* spp. was the least observed GIS s distribution study to provide more epidemiological information.

REFERENCES

1. Central Statistical Agency of Federal Democratic Republic of Ethiopia (2021): Livestock and livestock characteristics, agricultural sample survey. Addis Ababa, Ethiopia. Statistical Bulletin. 2021; 2:13-24.
2. Biffa D, Jobre Y, Chakka H. Ovine helminthosis, a major health constraint to productivity of sheep in Ethiopia. *Anim Health Res Rev.* 2006; 7: 107-118.
3. Kumsa B, Wossene A. Abomasal nematodes of small ruminants of Ogaden region, Eastern Ethiopia prevalence, worm burden and species composition. *Revue de Médecine Vétérinaire.* 2007; 157: 27-32.
4. Management Entity. Ethiopia's Livestock Systems: Overview and Areas of Inquiry. Gainesville, FL, USA: Feed the Future Innovation Lab for Livestock Systems. 2021.
5. Menkir S. Helminth Parasites of Sheep and Goats in Eastern Ethiopia. Epidemiology and Anthelmintic Resistance and its Management. Faculty of Veterinary Medicine and Animal Science. Department of Biomedical Sciences and Veterinary Public Health Division of Parasitology and Virology Uppsala Sweden Doctoral thesis Swedish University of Agricultural Sciences. 2007.
6. Urquhart GM, Armour J, Duncan JL, Dunn, AM, Jennings FW. *Veterinary Parasitology*, 2nd ed.; Blackwell Science: Oxford, UK, 1996; 307; ISBN 0632040513.
7. Knox MR, Besier RB, Le Jambre LF, Kaplan RM, Torres-Acosta JF, J Miller, et al. Novel approaches for the control of helminth parasites of livestock VI: summary of discussions and conclusions. *Vet Parasitol.* 2012; 186: 143-149.
8. Charlier J, Morgan ER, Rinaldi L, van Dijk J, Demeler J, Höglund J, et al. Practices to optimise gastrointestinal nematode control on sheep, goat and cattle farms in Europe using targeted (selective) treatments. *Vet Rec.* 2014; 175: 250-255.
9. Regassa F, Sori T, Dhuguma R, Kiros Y. Epidemiology of Gastrointestinal Parasites of Ruminants in Western Oromia, Ethiopia. *Intern J Appl Res.* 2006; 4: 51-57.
10. Eke SS, Omalu ICJ, Ochaguba JE, Urama AC, Hassan SC, Otuu CA, et al. Gastrointestinal Helminths Infections in Small Ruminants Slaughtered in Minna Modern Abattoir, North Central, Nigeria. *J Vet Anim Sci.* 2020; 7: 13-18.
11. Tembely S, Lahlou-kassi A, Rege JEO, Sovani S, Diedhiou ML, Baker RL. The epidemiology of nematode infections in sheep in a cool tropical environment. *Vet Parasitol.* 1997; 70:129-141.
12. Bekele T, Woldeab T, Lahlou-Kassi A, Sherngtion J. Factors affecting morbidity and mortality on farm and on station in Ethiopia highland sheep. *Acta Trop.* 1992; 52: 99-109.

13. Tembely S. Development and survival of infective larvae of nematode parasites of sheep on pasture in a cool tropical environment. *Vet Parasitol.* 1998; 79:81-87.
14. Menkir S, Uggla A, Waller PJ. Prevalence and seasonal incidence of nematode parasites and fluke infections of sheep and goats in eastern Ethiopia. *Trop Anim Health Prod.* 2007; 39:521-531.
15. Menkir S, Asefa A, Uggla A, Waller PJ. Assessment of anthelmintic resistance in nematode parasites of sheep and goats owned by smallholder farmers in eastern Ethiopia. *Trop Anim Health Prod.* 2006; 38: 215-222.
16. Keyyu JD, Kyvsgaard NC, Monrad J, Kassuku AA. Epidemiology of gastrointestinal nematodes in cattle on traditional, small-scale dairy and large-scale dairy farms in Iringa district, Tanzania. *Veterinary Parasitology.* 2005; 127:285-294.
17. Kaplan RM, Vidyashankar AN. An inconvenient truth: global worming and anthelmintic resistance. *Vet Parasitol.* 2012; 186: 70-78.
18. Aga TS, Tolossa YH, Terefe G. Epidemiology of gastrointestinal nematodes of Horro sheep in Western Oromiya, Ethiopia. *J Vet Med Anim Health.* 2013; 5: 296-304.
19. Wakayo BU, Dewo TF. Anthelmintic resistance of gastrointestinal parasites in small ruminants: a review of the case of Ethiopia. *J Veterinar Sci Technol.* 2015; S 10.
20. Kelemework S., Tilahun A, Benalfew E, Getachew A. A study on prevalence of gastrointestinal helminthiasis of sheep and goats in and around Dire Dawa, Eastern Ethiopia. *J Parasitol Vector Biol.* 2016; 8: 107-113.
21. Bedada H, Gizaw F, Negash W. Preliminary study on small ruminant GIT helminthiasis in select arid and semi-arid pastoral and agro-pastoral areas of Afar region, Ethiopia. *J Vet Adv.* 2018; 7: 210-217.
22. Farooq UB, Abdulla A, Chaudhary R, Abera M. A Cross-sectional Study on the Prevalence and Associated Risk Factors of Gastrointestinal Nematodes of Sheep in and around Hirna, South East Ethiopia. *Int J Livest Res.* 2018; 8: 89-97.
23. Handiso T, Yohannes B, Alemu B. Prevalence of Gastrointestinal Nematodes in Sheep at Yabello Town, Borena, Ethiopia. *Dairy and Vet Sci J.* 2019; 11: JDVS.M.S.ID.555813.
24. Kuma B, Abebe R, MekbibB, Sheferaw D, Abera M. Prevalence and intensity of gastrointestinal nematodes infection in sheep and goats in semi-intensively managed farm, South Ethiopia. 2019; *JVMAH* 11: 1-5.
25. Chali AR, Hunde FT. Study on prevalence of major gastrointestinal nematodes of sheep in Wayu Tuka and Diga District, Oromia Regional State. *Vet Med Open J.* 2021; 6: 13-21.
26. Gizaw S, Alemu B, Desta H, Asfaw T, Mersha T, Mekonnen M, Arke A, et al. Community-based approach for the control of gastrointestinal parasites under smallholder sheep farming systems. Nairobi, Kenya: ILRI. 2021.
27. Moje N, Gurmessa A, Regassa G. Gastro-intestinal Tract Nematodes of Small Ruminants: Prevalence and Their Identification in and Around Alage, Southern Ethiopia. *Adv Anim Vet Sci.* 2021; 9: 65-72.
28. Dawit I, Weldegebriel M, Dejene D, Israel I. Prevalence of Gastrointestinal Tract Nematodes Parasites in Sheep in Hawasa Town, Southern Ethiopia. *Biomed J Sci & Tech Res.* 2022; 41.
29. Woldemariam DL. Nematode Prevalence, Helminth Management Practices and Anthelmintic Resistance in Small Ruminants in the Mid-Rift Valley of Ethiopia. University of Pretoria etd. 2005.
30. National metrology agency. 2020.
31. Ejersa lefo Livestock Office. 2020.
32. Tokekutaye worda livestock and fisheries rural development office. 2020.
33. Ilugelan worda livestock and fisheries rural development office. 2020.
34. Hansen J, Perry B. The epidemiology, diagnosis and control of helminth parasites of ruminants. A handbook. ILRAD. Nairobi, Kenya. 1994; 17-132.
35. Van-Wyk A, Cabaret J, Michael M. Morphological identification of nematode larvae of small ruminants and cattle simplified. *Vet Parasitol.* 2004; 119: 277-306.
36. Van Wyk JA, Mayhew E. 'Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: A practical lab guide', Onderstepoort. *J Vet Res.* 2013; 80: 539.
37. Kumsa B, Debela E, Megersa, B. Comparative Efficacy of Albendazole, Tetramizole and Ivermectin Against Gastrointestinal Nematodes in Naturally Infected Goats in Ziway, Oromia Regional State (Southern Ethiopia). *J Anim Vet Adv.* 2010; 23: 2905-2911.
38. Frandson RD, Wilke WL, Fails AD. Anatomy and physiology of farm animals. Department of anatomy and neurobiology collage of veterinary medicine and bio medic sciences, 5thEdn. Colorado state University: for Collins, Colorado. 1992; 337-341.
39. Thrusfield MV. *Veterinary Epidemiology*, 3rd edition. Oxford, England: Blackwell Science. 2005; 234-238.
40. Morgan ER, Torgerson PR, Shaikenov BS, Usenbayev AE, Moore ABM, Medley GF, et al. Agricultural restructuring and gastrointestinal parasitism in domestic ruminants on the rangelands of Kazakhstan. *Vet Parasitol.* 2006; 139: 180-191.
41. Ministry of Agriculture, Fisheries and Food (MAFF). *Manual of Veterinary Parasitological Laboratory Techniques.* HMSO, London. 1986; 1-152.
42. Taylor, MA. Protozoal disease in cattle and sheep. In *Practice.* 2000; 22: 604-617.
43. Dohoo I, Martin W, Stryhn H. *Veterinary epidemiologic research*, 2nd ed. AVC, Charlottetown, Prince Edward Island. Pp. 2009; 239-249.
44. Getachew T, Muktar Y, Mekonnen N, Tesma F. Prevalence of gastrointestinal nematodes and efficacy of commonly used anthelmintics in different sheep breeds in Areka Agricultural Research Center, Areka, Ethiopia. *LRRD.* 2016; 28.
45. Moje N, Gurmessa A, Regassa, G. Gastro-intestinal Tract Nematodes of Small Ruminants: Prevalence and Their Identification in and Around Alage, Southern Ethiopia. *Adv Anim Vet Sci.* 2021; 9: 65-72.
46. Jatau ID, Abdulganiyu A, Lawal AI, Okubanjo OO & Yusuf KH. Gastrointestinal and haemoparasitism of sheep and goats at slaughter in Kano, northern-Nigeria. *Sokoto J Vet Sci.* 2011; 9: 7-11.
47. Asif M, Azeem S, Asif S, Nazir S. Prevalence of Gastrointestinal Parasites of Sheep and Goats in and around Rawalpindi and Islamabad, Pakistan. *J Vet Anim Sci.* 2008; 1: 14-17.
48. Abebe W, Esayas G. Survey of ovine and caprine gastro-intestinal helminthosis in eastern part of Ethiopia during the dry season of the year. *Revue Med Vet.* 2001; 152: 379-384.
49. Tesfaheywet Z, S Murga. Prevalence, Species Composition and Worm Burden of Abomasal Nematodes of Small Ruminants Slaughtered in Hawassa, Southern Ethiopia. *Afr J Food Agric Nutr Dev.* 2019; 19: 14916-14931.
50. Owusu M, Sekyere JO, Adzitey F. Prevalence and burden of gastrointestinal parasites of Djallonke sheep in Ayeduase, Kumasi, Ghana. *Vet World.* 2016; 9: 361-364.

51. Yan X, Liu M, He S, Tong T, Liu Y, Ding K, et al. An epidemiological study of gastrointestinal nematode and Eimeria coccidia infections in different populations of Kazakh sheep. *PLoS One*. 2021; 16: e0251307.
52. Ousman A, Meribo A. Study on prevalence of gastrointestinal nematodes of small ruminant in Adami Tulu Jiddo Kombolcha District, East Shoa Zone of Oromia, Ethiopia. *Int J Adv Res Biol Sci*. 2022; 9: 72-81
53. Abebe R, Gebreyohannes M, Mekuria S, Abunna F, Regassa A. Gastrointestinal nematode infections in small ruminants under the traditional husbandry system during the dry season in southern Ethiopia. *Trop Anim Health Prod*. 2010; 42:1111-1117.
54. Belina D, Giri A, Mengistu S, Eshetu A. Gastrointestinal Nematodes in Ruminants: The Parasite Burden, Associated Risk Factors and Anthelmintic Utilization Practices in Selected Districts of East and Western Hararghe, Ethiopia. *J Vet Sci Technol*. 2017; 8: 433.
55. Fayisa O, Duguma A, Temesgen M, Lemma F. Gastrointestinal Parasites of Sheep and Goat In and Around Gondar Town, Northwest, Ethiopia. *Biotechnology in Animal Husbandry*. 2020; 36: 371-380.
56. Poddar PR, Begum N, Alim MA, Dey AR, Hossain MS, Labony SS. Prevalence of gastrointestinal helminths of sheep in Sherpur, Bangladesh. *JAVAR*. 2017; 4:274-280.
57. Mahlehla MA, Molapo MS, Phoofolo MW, Matebesi PA, Phalatsi M, Moiloa MJ. Prevalence and Faecal Egg Counts of Gastrointestinal Parasites of Merino Sheep in Lesotho. *World Vet J*. 2021; 11: 85-91.
58. Martins NS, Santos CC, Motta SP, Moreira AS, Farias NAR, Ruas JL. Gastrointestinal Parasites in Sheep from the Brazilian Pampa Biome: Prevalence and Associated Factors. *Braz J Vet Med*. 2022; 44: e001522.
59. Troncy PM. Helminths of Livestock and Poultry in Tropical Africa. In: *Manual of Tropical Veterinary Parasitology*. CTA: CAB International. 1989; 23-50.
60. Waruiru RM, NC Kyvsaard, SM Thamsborg, P Nansen, HO Bogh, WK. Munyua, et al. Prevalence and intensity of helminth and coccidial infections in dairy cattle Kenya. *Vet Res Commun*. 2000; 24: 39-53.