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Research Article

Prevalence of Haemoparasites in *Clarias* Species (Cat Fish) In Lake Chad

Mohammed KM¹, Mbaya AW², Ahmed MI², Konto Mohammed^{2,3}, and Falmata Kyari²

¹Department of Veterinary Public Health and Preventive Medicine, University of Ilorin, Nigeria

²Department of Veterinary Parasitology and Entomology, University of Maiduguri, Nigeria

³North-East Zonal Biotechnology Centre of Excellence, University of Maiduguri, Nigeria

Abstract

*Corresponding author

Konto Mohammed, Department of Veterinary Parasitology and Entomology, University of Maiduguri, Borno State, Nigeria, Tel: +2348103702600; Email: kaltum.km@ gmail.com

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The prevalence of haemoparasites of *Clarias* species in Lake Chad was determined using standard parasitological techniques. Out of the 220 *Clarias* species collected, 37 [16.8%] were positive for various haemoparasitic infections. The parasites encountered in *Clarias* were *lchtyobodo* species 25 [11.4%] and *Dactylosoma* species 12 [5.5%]. Infection with lchtyobodo species was 11[8.3%] among the male *Clarias* while 14 [15.9%] was encountered among the female. Similarly, no statistical variation was observed between sexes. However, infection with *Dactylosoma* species was found to be 4 [3.0%] in males and 8 [9.1%] among the females. *lchthyobodo* infection was observed to be 16 [13.3%] and 9 [9.0%] among adult and juveniles *Clarias*, respectively. Meanwhile, *Dactylosoma* infection were 7 [5.8%] and 5 [5.0%] among the adult and juvenile *Clarias*, respectively. However, general Sex-wise prevalence showed that female *Clarias* were significantly [0.001%] more infected 22 [25.0%] than the male 15 [11.4%]. According to age, adults were equally more infected than the juveniles 14 [14.0%]; however, no statistical variation was observed between age group. It is highly imperative to use other screening methods like molecular and serological techniques for more sensitivity and specificity.

INTRODUCTION

A substantial proportion of Nigeria's fish supply is still derived from feral fishes [1] which are at higher risk of diseases compared to cultured fish. Documented information on fish parasite taxonomy is few in Nigeria [2]. Fish is an important dietary protein in many parts of the world [3]. They are low in fat [4] and serves as a source of omega-3 (n-3) fatty acids. Fishing occur in a wide range of Nigerian cultures [5,6], Native Americans [7,8] and in the other parts of the world [9,10].

Fish diseases are not easily recognizable in the wild [11] because ill fishes often become prey to other fish predators or die quickly due to their inability to compete favourably for food and survival. The complexity of bionomics, arrays of biological vectors and scarce reference literature made studies on piscine parasites in Nigeria to be scanty and mostly relates to brackish and freshwaters in the south and the Kainji and Lake Chad [12-15]. The studies done in the Lake Chad basin were limited to Tilapia and *Clarias* species [16-18] Many facets of fish health however, remained hazy and more work is required to elucidate the epidemiology and menace of piscine parasitism in the study area on a broader perspective.

The African catfish *Clarias* gariepinus is one of the economically important freshwater fishes in Lake Chad basin

with great dominancy in spatial distribution among other feral fish species. Its distribution spans from the Maghreb region to the South-West Cape [16-18]. It has been reared for nearly 30 years in Africa with poor performance due to the absence of reliable production techniques [17-19] and numerous diseases afflicting its health. It is found in all inland waters of Nigeria and recognized as an important food fish [15]. It has been a suitable choice for research due to its aquacultural potentials [19,20] and ability to survive for a prolonged period outside its habitat.

Fisheries constitutes up to 60% of total protein intake in adults of rural habitats and used as medications (fish oils), in recreations and vital inclusions of livestock feeds. Fish constitute 40% of animal protein intake in Nigeria [21] and had been observed to be a source of serious foreign exchange drain on Nigerian economy [22] a total of N100.5 million was spent in Nigeria on importation of fish and fisheries products as at 1986. Fish serve as a good source of animal protein for man and his livestock [5,23]. The role of fish in nutrition is recognized, as it supplies a good balance of protein, vitamins and minerals [22].

Parasitic diseases in aquatics features prominently in the tropics comparable to what is seen in terrestrial animals [17,18] knowledge on the prevalence of these parasites in the aquatic system will provide useful information on the potential hazards in fish health and the ecosystem and to enhance control measures

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MATERIALS AND METHODS

Study Area

Lake Chad lies between Latitude 120N-140 20'N: Longitude 130-150 30'E and is located in the Sahelian vegetation zone south of the Sahara, with less than 600mm of rain annually [24] The vegetation terrain around the lake consists of thorny grasses and shrubs in the highlands and swampy grasses in low land areas.

Fish Sampling and Preservation

Clarias species were collected randomly at the Custom fish market in Maiduguri. Sampling was conducted from the catches of five fish vendors who are known to procure their stocks from Lake Chad waters.

A total of 120 adult and 100 juvenile feral Clarias were collected at Custom fish market, consisting of 132 males and 88 females. Live fishes were transported to the laboratory in covered plastic buckets with perforations at the top to provide for aeration.

Laboratory Identification of Clarias species

Live Clarias were collected at landing and identified using taxonomic guides to Nigerian freshwater fishes as described by Reed et al., [25-27]. Sampling was done on weekly basis for five months. Each Claria species was examined morphometrically for proper classification. Order and Family characteristics were first applied followed by taxonomic features of individual species. Fins, mouth, teeth, nostrils, gills and lateral line were the features employed in distinguishing specie from another.

Identification of parasites

Blood samples were collected by salvaging the fish and blood collected in heparinised capillary bottles. Thin blood Giemsa stained smear and haematocrit centrifugation technique [28] were employed for the detection of haemoparasites. Slides were examined for parasites with the aid of binocular light microscope using immersion objective (x100). Parasites were identified using taxonomic features as described by [15] and photomicrographs taken using digital camera with 10.0 Mega pixels of resolution and Optical image stabilizer (Canon IXUS 900IS).

Statistical Analysis

One-way analysis of variance with Tukey-Kramer multiple comparisons (Post-Hoc) test was employed to compare means for standard threshold alpha of less than 0.05. GraphPad InStat, Version 3.00 (1998) and Microsoft office excel (2010) software packages were employed in all statistical analyses and graphical presentations of results respectively.

RESULTS

Out of the 220 Clarias species collected, 37(16.8%) were positive for haemoparasitic infections (Table 1). Infection in females 22(25.0%) was significantly higher than in Male Clarias 15(11.4%) with a two sided P-value of 0.01. Two parasites were observed to be responsible for the infection in *Clarias* species in the study area, which comprise of *Ichthyobodo* sp (Plate 1) responsible for 25(11.4%) of the infection and Dactylosoma sp (Plate 2) responsible for 12(5.5%). Infection with *Ichthyobodo* sp. was 11(8.3%) in male Clarias while 14(15.9%) was recorded in females.

On the other hand, infection in adult Clarias accounted for 23(19.2%) when compared with the infection in juveniles 14(14.0%). There was no significant difference in infections observed between the juvenile when compared with the adult *Clarias* species (Table 2). Infection with *Dactylosoma* sp was observed to be 4(3.0%) in males and 8(9.1%) in female whose differences were insignificant P> 0.05. Ichthyobodo infection was observed to be 16(13.3%) and 9(9.0%) in adult and juvenile Clarias, respectively. While for Dactylosoma sp., infection rates were 7(5.8%) and 5(5.0%) in adult and juveniles, respectively.

DISCUSSION

Reports of piscine haemoparasites in Nigeria are scanty. In this study, two parasites were observed to be responsible for infecting Clarias sp. which comprises of the Ichthyobodo sp

Sex	Number examined	Number positive (%)	Specie positive (%)		Relative risk	95% Confidence	
			Ichthyobodo sp.	Dactylosoma sp	Relative risk	interval	
Male	132	15(11.4) ^a	11(8.3)ª	04(3.0) ^a	0.4545	0.0651 -0.1807	
Female	88	22(25.0) ^b	14(15.9)ª	08(9.1) ^a	0.4545	0.1638 - 0.3531	
Total	220	37(16.8)	25(11.4)	12(5.5)			

Rows with different superscripts are statistically significant.

Table 2: Distribution of haemoparasites in adult and Juvenile <i>Clarias</i> species in the Lake Chad.											
Sex	Number examined	Number positive	Specie positive (%)		Relative risk	95% Confidence interval					
			Ichthyobodo sp.	Dactylosoma sp		Lower	Upper				
Adult	120	23	16(13.3)	07(5.8)	1.369	0.1254 - 0.2732					
Juvenile	100	14	09(9.0)	05(5.0)		0.0786 - 0.2237					
Total	220	37	25(11.4)	12(5.5)							



Plate 1 Ichthyobodo sp. from skin lesions (Giemsa x1000).

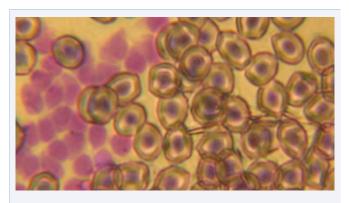


Plate 2 *Dactylosoma sp*; numerous extra-erythrocytic cystozoites in peripheral blood.

and *Dactylosoma* species. *Ichthyobodo* sp and *Dactylosoma* sp collectively constituted for about 16.8% prevalence which is higher than previous findings [14,15,18] The higher prevalence recorded in this study may not be unconnected with the constant proliferation of the parasite in the study area couple with paucity of awareness by the populace and low research interest on fish parasites by researchers as a whole.

As reported by earlier reports, the verse complexity of bionomics [17] the wide range of biological vectors and other intermediate host, coupled with scarce reference on literatures on piscine parasites in Nigeria [12,15] led to the paucity of information on fish parasites in the study area and the country at large. The low prevalence of protozoan parasites in piscivorous species such as *Clarias* indicated that endogenous stages of protozoan parasites do not contribute significantly in the transmission of the infection.

It is also worthy of note that this study indicates that female *Clarias* appeared to be more at risk than their male counter parts. This may be as results of the female's domiciliation in the lithoral region of the water body particularly during nesting as earlier reported by Karshima and Ahmed [17] in a similar

related scenario. The preponderance of leeches in these water bodies may also have been responsible for the propagation of the parasite in fish population. In a similar survey *Ichthyobodo* sp and *Dactylosoma* sp were reported in freshwater fishes (*Synodontis* species) of Lake Victoria with 19.6% and 23% prevalence rates, respectively [10].

CONCLUSION

This study has at least help in contributing to the existing body of knowledge concerning the parasitic diseases in aquatics by demonstrating the presence of two parasitic species of the *lchthyobodo* and *Dactylosoma* in Lake Chad, region of Nigeria. This study also indicated sex related risk of infection in which female *Clarias* are more vulnerable to infection than their male counterparts. It is further recommended that more research on other aspect of fish parasites, more especially the *Clarias* sp. be conducted in order to provide more knowledge on the prevalence of parasites in the aquatic system and provide useful information on the potential hazards in fish health and the ecosystem looking at the growing demand of fish as good source of animal protein for man and his livestock.

REFERENCES

- Falaye PE. Utilization of agro-industrial wastes as fish feedstuffs in Nigeria. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta. 1992; 47-57.
- 2. Paperna I. Parasites, infections and diseases of fish in africa. CIFA Technical Paper. 1980; 7: 216.
- 3. Burger J, Gochfeld M, Jeitner C, Burke S, Stamm T. Metal levels in flathead sole (*hippoglossoides elassodon*) and great sculpin (*Myoxocephalus polyacanthocephalus*) from Adak Island, Alaska: Potential risk to predators and fishermen. Environmental Research, (Corrected proof In Press). 2006.
- Anderson PD, Wiener JB. Eating fish. In: Graham JD, Wiener JB. Edition, Risk versus Risk: Tradeoffs in Protecting Health and the Environment, Harvard University Press, Cambridge, MA, USA. 1995.
- Fagade SO. Production, Utilization and marketing in fisheries status and opportunities. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta, 16th-20th November. 1992; 8-17.
- Kumar JS. Management of super-intensive farming of African cat fish. Akin Tee-Jay Prints, Animal Health care Konsult, Nig. Ltd.v. 2004; 1-55.
- 7. Harris SG, Harper BL. Native American exposure scenarios and tribal risk model. Risk Anayst. 1997; 17: 789-795.
- 8. Burger J. American Indians, hinting and fishing rates, risk and the Idaho National Engineering and Environmental Laboratory. Environmental Research. 1999; 80: 317-329.
- 9. Burger J, Fleischer J, Gochfeld M. Fish, shellfish and meat meals of the public in Singapore. Environmental Research. 2003; 92: 254-261.
- 10.Bristol ET, Tuckland F. Seroprevalence of piscine haemoparasites in Lake Victoria. Dutch periodicals. 2004; 23: 234-238.
- 11. Tatner MF. Fish vaccines; In: Vaccines for Veterinary Application, Peter AR. ed., Ballemouth Heinemann, Oxford. 1993; 199-223.
- 12.Aderounmu EA, Adeniyi F. Cestodes in fish from a pond at Ile-Ife, Nigeria. The African Journal of Tropical Hydrobiology and fisheries. 1971; 2: 151-152.
- 13.0kaeme AN, Obiekezie AI, Ogbondeminu FS. The economic impact

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of diseases and parasitic problems in freshwater fish production. Proceedings of the 5th annual conference of the fisheries society of Nigeria, Ilorin, September 22th-25th. 1998; 368-374.

- 14. Ahmed MI, Ambali AG. Freshwater fish parasites in the Lake 133 Chad. Proceedings of the 21stNVMA Conference, Maiduguri, Nigeria. 2005; 134: 27-30.
- 15. Ahmed MI, Ambali AG, Baba SS. Fresh water fish parasites in the Lake Chad basin. Journal of Food Agriculture & Environment. 2008; 6: 113-117.
- 16. Yaya JK. Prevalence of Helminth parasites of Clarias and Gymnarchus species in Maiduguri, Borno State. 2002; 1-120.
- 17. Karshima SN, Ahmed MI. Prevalence of coccidian oocysts in Clarias and Tilapia fish from Lake Alau, Maiduguri, Nigeria. Sahel Journal of Veterinary Sciences. 2012; 11: 65-67.
- 18. Ahmed MI, Ambali AG, Baba SS. Tissue responses of *Clarias gariepinus* (Catfish) to experimental Eimeria sub-epithelialis infection. Veterinary Parasitology. 2010; 171: 181-184.
- 19. Huisman EA, Richter CJJ. Reproduction growth, health control and aquacultural potential of the African catfish *Clarias gariepinus* (Burchell, 1822). Aquaculture. 1987; 63: 1-14.
- 20.Salami AA, Oluayo-Bello O, Olasogba LP. The effects of dietary treatment on the morphometric and haematological characteristics in *Clarias gariepinus*. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta, 16th-20th November. 1992; 147-151.

- 21. Olatunde AA. Approaches to the study of fisheries Biology in Nigerian inlandwaters. Proceedings of the National Conference of Two Decade of research on Lake Kainji. 1989; 538-541.
- 22. Akande GR, Tobor JG. Improved utilization and increased availability of fishery products as an effective control of aggravated animal protein deficiency-induced malnutrition in Nigeria. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta, 16th-20th November. 1992; 18-31.
- 23.Eyo AA. Utilization of freshwater fish species in Nigeria. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta, 16th-20th November; 1992; 32-38.
- 24. Troncy PM. Helminths of livestock and poultry. 1989; 126-199.
- 25. Reed W, Burchard J, Hopson AJ, Jenner J, Yaro I. Fish and fisheries of Northern Nigeria. Gaskiya Corporation, Zaria, Nigeria. 1965; 2-226.
- 26. Holden M, Reed W. West African freshwater fish. Longman Group Ltd., London. 1978; 2-66.
- 27.Adekeye AO. Age and growth of Heterotis niloticus around Pategi in the middle river Niger, Kwara state. Proceedings of the 10th annual conference of the fisheries society of Nigeria, Abeokuta, 16th-20th November. 1992; 152-158.
- 28.Woo PTK. The haematocrit centrifuge for the detection of trypanosomes in blood. Can J Zoo. 1969; 47: 921-923.

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